

STIMULATIVE RESPONSE TO QUANTITY AND QUALITY OF JOJOBA OIL BY GIBBERELIC ACID AND BENZYL ADENINE

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ABSTRACT

This study was aimed to evaluate five clones of jojoba plant under the influence of salinity stress which was sprayed by gibberellin and benzyl adenine on growth yield and oil. Two field experiments were carried out at Magharah Research and dmtng Production which follow Desert Research Center, Agriculture Ministry, in North Sinai governorate, Egypt, 2015/2016 and 2016/2017 respectively, the experimemts wene consist of five clones (S-L, S-610, S- 700, S-B and S-G), (were sprayed by, GA₃ 200ppm, BA 200ppm and GA₃+BA and control) under effect of salinity stress. The experiment was conducted as split plot design having varieties in main plot and IAA, GA₃ sub plots three replicates. The jojoba plant was affected by increased GA₃, BA or/and GA₃+BA, and it was clear in all clones. The effect of GA₃, BA or/and GA₃+BA varied from one clone to another and the lowest was s-700. All studied variables were confirmed that clones of jojoba had a significant effect with GA₃, BA or/and GA₃+BA

Key words: oil yield, water salinity, spray acid, sinai, water stress.

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استجابة كمية وجودة زيت الجوجوبا للتحفيز بحمض الجبريليك والبنزويل أدنين

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المستخلص

تهدف هذه الدراسة إلى تقييم خمس سلالات من الجوجوبا تحت تأثير الإجهاد الملحي برش النباتات بحامض الجبريلين وحامض بنزويل أدنين وأثر ذلك على النمو وإنتاجية الزيت. تم إجراء تجربتين حقليتين في محطة المغارة للأبحاث والإنتاج التي تتبع مركز بحوث الصحراء، وزارة الزراعة التي تقع في محافظة شمال سيناء، مصر، خلال السنتين 2016/2015 و2017/2016، لخمسة سلالات جوجوبا هي (SL، S-610، S-700، SB و SG)، وتم رشها بتركيز (GA₃ 200ppm، BA 200ppm و GA₃ + BA) بالإضافة إلى المقارنة تحت تأثير ملوحة مياه الري. نفذت التجربة بتصميم الألواح المنشقة ووزعت السلالات في الألواح الثانوية والمعاملات (IAA و GA₃) في القطع الفرعية وتم توزيعها في ثلاث مكررات. أشارت النتائج إلى تأثير الجوجوبا بزيادة GA₃ أو BA أو / و GA₃ + BA ، وكان ذلك واضحاً في جميع السلالات. وتباينت تأثيرات GA₃ أو BA أو / و GA₃ + BA من سلالة لآخرى وكان أقلها استجابة السلالة s-700. وأكدت جميع الصفات التي تم دراستها أن سلالات الجوجوبا كانت متأثرة بشكل كبير بالرش بكل من GA₃ أو BA أو / و GA₃ + BA.

كلمات مفتاحية: حاصل الزيت، ملوحة المياه، ملوحة التربة ، سيناء.

INTRODUCTION

Jojoba, *Simmondsia chinensis* (Link) Schneider, is native to the Sonoran desert. The seeds of jojoba contain a light-gold-colored wax ester (jojoba oil) that makes up 50–55% of the seed weight. Jojoba is an extremely salinity water stress species and is gaining worldwide attention, for extraction of oil which is used in pharmaceuticals and lubricant industries as a replacement of sperm whale oil. Jojoba is a high cash crop and Industrial, and it is need to low production costs for its quality of growth in all circumstances. In recent years, renewed interest in commercial cultivation of jojoba has begun as the useful properties of the liquid wax obtained from the seed have been confirmed(20, 29) Salinization threatens the productivity of agricultural land and agricultural sustainability. Salinity is a major problem that negatively affects agricultural activities in many regions in the world, salinity problems increase with increasing salt concentration in irrigation water. Crop growth reduction due to salinity is generally related to the osmotic potential of the root zone soil solution, (2). Water used for irrigation can vary greatly in quality depending on type and quantity of dissolved salts. Almost 50% of the irrigated land is affected by high salinity, often resulting in secondary salinization due to inappropriate use of saline irrigation water. Despite the essentiality of chloride as a micronutrient for all higher plants and of sodium as mineral nutrient for many halophytes and some species, salt accumulation may convert agricultural areas in unfavorable environments, reduce local biodiversity, limit growth and reproduction of plants, and may lead to toxicity in nonsalt-tolerant plants, known as glycophytes (6, 33). The effects of salinity are generally summarized as water stress, salt stress and stress due to ionic imbalance (14). Therefore at least one part of salt stress is associated with water stress, which is a general condition, and it can be expected that plant adaptation to salinity may show features similar to those characteristic of adaptation to water stress. It was believed to be future arid plant, in general, jojoba developed reasonably well under salinities of 8 dSm⁻¹ (9). Salinity and drought are considered to be the most serious growth-

limiting factors for crop plants (11, 41). Increasing salinity of irrigation water has contributed to progressive salinization of agricultural soils inhibiting agricultural productivity in many semi-arid and arid regions of the world (35). Sodium chloride (NaCl) is the most commonly encountered source of salinity (24). Much of the strain in salinity stress is related to water stress arising from excessive uptake of salts by the plants and the resulting reduction in water potential. For this crop to be an economically profitable alternative for arid and semi-arid zones, it is necessary to first select plants of high productivity that also possess sufficient resistance to abiotic stresses. Salinity is considered to be the most serious growth limiting factors for crop plants (4, 41). In spite of information showing that jojoba tolerates fairly high levels of salinity (9) and water stress the selections to date have not been intended for use in regions with extremely high levels of salinity and water stress (10). Therefore, the aim of this study was to evaluate the effect of salinity concentrations on shoot and root growth, leaf measurements and chemical constituents of jojoba leaves, (18). The stimulative response of gibberellic acid, which known to be one of the endogenous growth regulators, and could be attributed to its unique roles in plant growth and development as reported by many investigators. (3) suggested that GA₃ has the capability of modifying the growth pattern of treated plants.(39) Gibberellic acid is used to regulation plant growth through increasing affecting the DNA and RNA levels, cell division and expansion, biosynthesis of enzymes, protein, carbohydrates and photosynthetic pigments. GA₃ treatment increased significantly shoot growth (shoot length, number of leaves, number of branches and leaf area) and root growth (root length, number of roots & root thickness) compared to the control, (53) and (22). The effect of benzyl adenine on the plant growth and chemical constituents of different plants have mentioned by (153); (33); (34); (32) and (51). Studies of exogenous applications of various plant growth regulators (PGRs) showed that PGRs play important roles in floral development (27); (19); (47). Exogenous cytokinin (CK)

application has been shown to increase meristem activity and promote floral initiation in several species (54); (55); (24). Ohkawa (40) and (23) found that 6-benzyladenine (BA), a synthetic compound with CK activity) treatment had a significant influence on increasing flower numbers of *Lilium speciosum*, particularly when combined with gibberellins. Chen (13) showed that flower bud differentiation of lychee (*Litchi chinensis*) was significantly promoted by exogenous kinetin application after bud dormancy. The total number of flowers on jojoba (*Simmondsia chinensis*) was also significantly increased by treatment with BA (45); (43). Most of the criteria of vegetative growth expressed as number of leaves/plant, leaf area and petiole length and were significantly affected by application of GA₃ and BA. GA₃ and BA separately promoted all the aforementioned characters in this study compared with control plants; Parviz rahbarian, et al., (42). The aim of this study is investigation of five clones of jojoba plants (S-L, S-610, S- 700, S-B and S-G), were spray by (control, GA₃ 200ppm, BA 200 ppm and GA₃+BA) that growing under effect of water salinity stress.

MATERIALS AND METHODS

Plant and Treatments

Jojoba plants were cultivated in Almaghara Research and Production Station (latitude: 30,717993"N, longitude: 33, 329103 E) followed to Desert Research Center,

Table 1a. Main physical and chemical properties of the soil at 2015/2016 and 2016/2017

Character	At 0-30cm	At 30- 60 cm
Fine sand (%)	81.4	84.8
Silt (%)	8.2	4.5
Clay (%)	11.28	10.7
Texture	Sandy	Sandy
Organic matter (%)	0.93	0.85
pH	7.9	7.88
EC (mmhos/cm ²)	2.84	2.46
CaCO ₃	1.38	1.11
SO ₄	4.89	4.56
HCO ₃	1.24	1.11
Total N (%)	5.6	4.1
Available P (%)	4.7	4.3
K (%)	4.55	5.6
Mg %	3.21	3.78
Fe (ppm)	1.43	1.3
Zn (ppm)	0.5	0.35
Mn (ppm)	1.3	1.2
Na+	20.41	19.66
Cl-	22.15	21.25
Ca++	2.27	2.14

Agriculture Ministry, Egypt at 2015/2016 and 2016/2017 respectively. Two field experiments were carried out for five clones (S-L, S-610, S- 700, S-B and S-G), to study the effect of spray Gibberellic acid GA₃ 200 ppm and Benzyl adenine BA 200 ppm and interaction between them on jojoba plants at four years old from planting to investigation content seed from oil and other chemical contents.

Experimental device

The experimental rows (5 plants each) were assigned for each clone in each replication. Distances between rows and within plants in rows were 3 and 2 m. respectively. Plants (mixed males and female seedlings) derived from the open population, as a source of pollen, were repeated one row every six female (clone) rows. Additional border mixed seedling rows were planted around each replication and no free space was left between rows within each replication to ensure homogeneity within each replication. The main physical and chemical properties of the soil of the experiment) and water irrigation analysis are shown in Table 1(a and b). A drip irrigation system was installed in the experimental areas and weed control were done as necessary applied in the course of the study. All clones were treatment at three times (October, March and April) with concentration of Gibberellic acid and Benzyl adenine.

Table 1b. Chemical characteristics of water weal used for the present study

Parameters	pH	EC(dSm ¹)	Ca ⁺⁺	Mg ⁺⁺	Na ⁺	K ⁺	HCO ₃ ⁻	Cl ⁻	SO ₄ ⁻
Values	7.49	3.77	5.44	3.56	21.83	3.21	1.96	28.21	3.22

Determination of growth/ yield parameters of plants:

Vegetative and reproductive measurements: Data were collected from tagged plants of each clone for each block: plant height; branch characters: Two branches were selected from the mid- level of the plants and the average of two branches characters were calculated for detailed analysis. From each branch the following data were collected; number of main branches/plant, stem diameter (cm), number of nodes/stem, length of node (mm), leaf number of leaves/plant, leaf width (mm), length (mm) and leaf area (cm²), and this last character was calculated as mentioned by Koller (22). Leaf area (cm²) was estimated from the following equation:

$$\text{Leaf area} = 0.717 X - 0.095,$$

which X is the product of length by width, Kohorn, (4). Chlorophyll contents were assayed in the commercial harvest stage. They were determined according to Wintermans and Mats (45) as follows: half gram of fresh leaves was extracted by about 15 ml. of 85% acetone with 0.5 g calcium carbonate, the mixture was through a glass funnel and the residue was washed with a small volume of acetone and completed to 25 ml. The optical density of Journal of a constant volume of filtrate was measured at a wave length of 622 nm for chlorophyll A, 644 nm and for chlorophyll B using spectrophotometer

Seed yield: Seed were harvested from the previous tagged plants by hand at full maturity. Harvested seeds were cleaned, dried and weighted (g). Seeds were hand-harvested every year in July and August from five plants per clone and used for determining seed yield plant⁻¹ (g) and analyzed for main components. Oil content was determine oil content, seeds of each genotype were randomly selected, weighed, and dried at 50 °C. The drying process was continued until the difference between the two successive weights was less than 1 mg five replications were used for this characteristic. The oil was extracted for 16 h with hexane with a Soxhlet apparatus. Total carbohydrates: Total carbohydrates were estimated by the difference in the mean values, i.e., 100 - (sum of concentrations of protein, ash and lipid). Determination of total

carbohydrates was determined spectrophotometrically (as glucose) after acid hydrolysis using phenol sulphuric acid reagent, according to (11). Mineral content: To remove carbon, approximately 5gm of each dry sample was ignited in a porcelain container and incinerated in the muffle furnace at about 550oC. Mineral content was expressed as a percentage of dry matter. Total Nitrogen content determined using Micro-Kjeldahl method Jackson (20). Protein content was determined by the Kjeldahl method for the calculation of all proteins which equal nitrogen content multiplied by 6.25, AOAC (3). Potassium content was extracted according to Chaudhary et al. (12). Phosphorous content using chlorostannous reduced molybdo-phosphoric acid blue color method, in hydrochloric described system as described by Jackson (20). Magnesium Mg %, Zinc Zn m/kg, copper Cu m/kg and iron Fe m/kg assay by atomic absorption.

Statistical analysis

The experiment was conducted as split plot design having varieties in main plot and IAA, GA₃ sub plot and distributed in three replicates. Statistical analyses and mean comparisons were conducted using MSTAT-C software. (38).

RESULTS AND DISCUSSION

All data in Tables 2, 3 and 4 show that the foliar application of gibberellic acid or/and benzyl adenine lead to increase of all growth characters as plant height (cm), number of main branches/plant, stem diameter (cm), number of nodes/stem, length of node (mm), leaf number of leaves/plant, leaf width (mm), leaf length (mm) and leaf area (cm²) and, yield characters as total number of branches, number of leaves, weight of seeds/plant (kg), weight of seed (gm.), weight 100 seeds (gm.) and oil content of the seeds %, Some chemical contents in leaves as chlorophyll content (chl. A, Chl. B and carotene); total Carbohydrates, nitrogen%, phosphorus%, Potassium%, Fe ppm, Zn ppm, Cu ppm and Mg ppm. All characters affected by increase in GA₃ and BA under clone S-700. Data presented in Table 1 and 2 show that, the foliar application of different concentrations of gibberellic acid

(GA₃) and benzyl adenine (BA) had significantly stimulatory effect on growth parameters of jojoba plants in term of plant height, number of branches, and leaves/plant and leaf area compared with the untreated plants, as this respect Afifi et al., (3), (5, 37).

Table 2. Effect of rates of GA₃ and BA and clones of jojoba plants on growth characters

Clones Treatments	plant height (cm)	shoot characters			leaves characters				
		number of main branches/plant	stem diameter (cm)	n umber of nodes/stem	Length of node (mm)	leaf number of leaves/plant	leaf width (mm)	length (mm)	leaf area (cm ²)
0	73.21	2.448	1.282	15.04	11.19	87.46	27.68	38.11	7.672
S-L GA ₃ 200ppm	76.77	2.569	1.345	16.43	12.30	92.60	28.72	38.36	7.994
BA200ppm	78.14	2.975	1.557	16.67	12.42	99.57	28.20	39.04	8.146
GA ₃ +BA	79.74	3.036	1.589	17.04	12.67	101.64	28.78	39.84	8.316
0	74.78	2.521	1.319	15.54	11.59	90.035	27.73	38.19	7.712
S-610 GA ₃ 200ppm	78.36	2.646	1.385	16.78	12.53	101.61	28.78	38.43	8.054
BA200ppm	79.70	3.064	1.603	16.92	12.64	108.39	29.44	38.60	8.448
GA ₃ +BA	81.33	3.127	1.637	17.39	12.91	110.60	30.04	39.39	8.614
0	76.25	2.598	1.359	15.80	11.81	92.805	27.79	38.26	7.786
S-700 GA ₃ 200ppm	79.97	2.725	1.425	16.78	12.53	104.47	30.11	38.51	8.491
BA200ppm	81.38	3.156	1.651	17.80	13.34	111.88	29.50	38.67	8.528
GA ₃ +BA	83.04	3.221	1.686	18.27	13.61	114.16	30.11	39.47	8.706
0	75.53	2.572	1.346	15.67	11.68	91.479	27.76	38.22	7.746
S-B GA ₃ 200ppm	79.18	2.699	1.412	16.91	12.60	100.32	28.81	38.47	8.091
BA200ppm	80.48	3.125	1.636	17.42	12.97	106.45	29.55	38.63	8.482
GA ₃ +BA	82.14	3.189	1.669	17.89	13.29	108.63	30.07	39.43	8.668
0	74.01	2.497	1.307	15.29	11.43	88.749	27.70	38.15	7.680
S-G GA ₃ 200ppm	77.63	2.620	1.371	16.66	12.42	97.752	28.75	38.40	8.021
BA200ppm	78.97	3.034	1.588	16.89	12.53	103.34	29.40	38.56	8.410
GA ₃ +BA	80.59	3.096	1.620	17.166	12.801	105.46	30.01	39.35	8.582
LSD C	13.222	0.8137	0.0833	2.0332	2.866917	15.867	7.4419	8.7928	0.84686
LSD T	11.413	0.4603	0.0485	4.3042	2.76033	13.504	4.7233	8.0646	0.57374
LSD C x T	8.098	0.2757	0.0373	4.1717	1.814712	10.080	2.8561	6.3975	0.20779

Table 3. Effect of rates of GA₃ and BA and clones of jojoba plants on yield

Clones Treatments	total number of branches,	No. leaves	W, Seeds harvest (kg)	Weight of seed gm	Weight (100 seed gm)	oil content of the seeds %
0	7.87	125.62	66.64	0.6758	67.58	40.05
S-L GA ₃ 200ppm	8.26	132.97	67.99	0.7155	71.55	45.42
BA200ppm	9.47	141.51	67.33	0.8097	80.97	47.62
GA ₃ +BA	9.76	145.89	69.41	0.8348	83.48	49.09
0	8.11	129.29	70.59	0.6957	69.61	41.44
S-610 GA ₃ 200ppm	8.51	145.89	72.04	0.7354	73.70	46.41
BA200ppm	9.75	153.65	71.35	0.8386	83.42	49.36
GA ₃ +BA	10.05	158.41	73.55	0.8646	86.00	50.88
0	8.35	133.56	74.74	0.6957	69.68	42.44
S-700 GA ₃ 200ppm	8.76	150.85	76.11	0.7354	73.78	48.10
BA200ppm	10.04	159.24	75.36	0.8386	83.50	50.42
GA ₃ +BA	10.35	164.18	77.70	0.8646	86.08	51.98
0	8.27	131.98	72.57	0.6857	68.99	41.64
S-B GA ₃ 200ppm	8.68	144.30	74.07	0.7354	73.05	46.81
BA200ppm	9.95	151.44	73.36	0.8290	82.68	49.74
GA ₃ +BA	10.25	156.12	75.63	0.8546	85.24	51.28
0	8.03	127.41	67.62	0.6857	68.93	40.85
S-G GA ₃ 200ppm	8.42	140.32	69.00	0.7254	72.98	45.91
BA200ppm	9.66	146.91	68.34	0.8290	82.59	48.49
GA ₃ +BA	9.95	151.45	70.45	0.8546	85.15	49.99
LSD C	2.09783	31.0123	9.29889	0.31861	9.8492	9.2233
LSD T	1.95182	22.9740	8.73443	0.00998	8.3286	7.9494
LSD C x T	1.54885	17.1969	7.13716	0.00222	7.5144	8.0442

The most effective treatment which had the tallest plants, the largest leaf area and the highest number of leaves was (GA₃) when applied at concentration of 200 ppm. The results are in agreement with (37) and (15); (39) and Soad (2005) on Jojoba plant. However, (GA₃) is used to regulating plant

growth through increased meristematic activity due to enhance cell division and elongation Treatment with BA at 150 ppm gave the highest value of the number of branches/plant compared with the other treatments and control.

Table 4. Effect of rates of GA₃ and BA on clones of jojoba plants on content of leaves from chemical content

Clones		Chlorophyll 51Content			Total	Nitrogen%	phosphorus%	Potassium%	Fe	Zn	Cu	Mg
Treatments		Chl.A	Chl. B	Carot.	Carbohydrates				ppm	ppm	ppm	ppm
	0	2.006	1.503	2.857	9.987	2.484	0.235	1.513	1218.2	4275.3	1207.3	0.7474
	GA ₃											
S-	200ppm	3.045	1.671	3.283	11.964	3.155	0.244	1.949	1436.6	4709.6	1217.2	0.8887
L	BA200ppm	3.352	1.840	3.613	13.170	3.474	0.268	2.146	1581.8	5185.2	1340.8	0.9784
	GA ₃ +BA	3.836	1.693	4.005	13.250	3.187	0.276	2.022	1635.4	5153.6	1228.1	0.9591
	0	2.956	1.563	3.075	10.679	2.731	0.261	1.877	1232.8	4291.6	1219.3	0.7831
	GA ₃											
S-	200ppm	3.440	1.738	3.322	13.151	3.214	0.272	2.182	1453.8	4728.5	1229.3	0.9314
61	BA200ppm	3.787	1.914	3.662	14.480	3.539	0.299	2.402	1600.7	5206.0	1353.8	1.0255
0	GA ₃ +BA	4.469	1.801	4.884	13.448	3.305	0.296	2.221	1655.0	5174.1	1240.3	1.0056
	0	2.956	1.563	3.075	10.679	2.731	0.261	1.877	1232.8	4291.6	1219.3	0.7831
	GA ₃											
S-	200ppm	3.440	1.738	3.322	13.151	3.214	0.272	2.182	1453.8	4728.5	1229.3	0.9314
70	BA200ppm	3.787	1.914	3.662	14.480	3.539	0.299	2.402	1600.7	5206.0	1353.9	1.0255
0	GA ₃ +BA	4.469	1.801	4.884	13.448	3.305	0.296	2.221	1655.0	5174.1	1240.3	1.0056
	0	2.857	1.595	3.085	11.173	2.692	0.273	1.900	1544.0	5194.0	1210.4	0.7883
	GA ₃											
S-	200ppm	3.124	1.773	3.411	13.350	3.237	0.284	2.091	1468.4	4742.6	1221.3	0.9360
B	BA200ppm	3.439	1.952	3.751	14.699	3.567	0.312	2.302	1616.7	5221.6	1344.6	1.0390
	GA ₃ +BA	4.301	1.835	4.450	13.547	3.318	0.305	2.215	1671.6	5189.7	1250.3	1.0113
	0	2.827	1.533	3.045	10.383	2.641	0.240	1.844	1525.6	5173.4	1198.4	0.7657
	GA ₃											
S-	200ppm	3.094	1.705	3.381	13.052	3.176	0.271	2.030	1451.1	4723.8	1209.6	0.9089
G	BA200ppm	3.406	1.877	3.721	14.369	3.495	0.298	2.235	1598.5	5200.9	1331.3	1.0016
	GA ₃ +BA	4.261	1.764	4.409	13.349	3.254	0.284	2.151	1651.7	5169.0	1238.0	1.0133
	LSD C	0.0867	0.0227	0.0599					153.86	923.16	2.3173	0.0093
	LSD T	0.0782	0.0221	0.0941					138.52	831.19	0.1895	0.0085
	LSD C x T	0.0027	0.0033	0.1296					131.99	792.04	0.1904	0.0081
		2	8	4	4.24344	0.10151	0.00557	0.10725	7	1	1	8

According to Table 4 it was found that the leaves content of three photosynthetic pigments chlorophyll (chl. a and chl. b) and carotenoids were gradually increased as the concentration of GA₃ and BA were increased. The highest value of Chl (a and b) and carotenoids was obtained at concentration of GA₃ (200 ppm). The results herein are agreement with the finding of Mousa et al., (30) on *Nigella sativa*, (37) and (5) they mentioned that GA₃ treatments were more effective than kinetin in increasing photosynthetic pigments in croton leaves. Application of GA₃ and BA with 200ppm concentrations were favorable for accumulation of total soluble sugars in leaves of the tested plants compared with the control. The greatest content of total soluble sugars occurred in the leaves of plants treated with GA₃ and BA at 200 ppm. In harmony with these results were those obtained by (37) and (31) and Nahed (39) on flax plants recorded

that GA₃ and BA application resulted in an increase of total soluble sugars content. These results are in line with those obtained by Lobna et al., (25). This study clearly demonstrated that growth hormones, whether alone or in combination, have a major impact in the stimulation of various growth parameters in jojoba plants. It was concluded that plant growth hormones could be successfully employed for enhancement of seed yield, directly or indirectly, through its components. Increase oil content by application of plant growth regulators on jojoba. The salt stress has a significant impact on the productivity of jojoba. The treatment of growing plants under the influence of Sinai salinity with gibberellic acid GA₃ and benzyl adenine used as growth regulators induces the accumulation of substances such as antioxidants and photosynthesis products which reduces the effect of salinity, especially with S-700 clone.

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