EFFECT OF BIOFILM WITH BIOFERTILIZER OF Pseudomonas fluorescens AND Rhizobium leguminosarum, CHEMICAL FERTILIZER LEVEL AND ADDATION TECHNIQUE ON SOME GROWTH AND YIELD TRAITS OF

WHEAT(Triticum asetivum L.)

HA.Abdul-Ratha	S. J. Jasim
Prof.	Assist Lecturer
Dept . Soil Science and water Resource	Dept. Plant Protection
Coll. of Agric. of Univer . of Baghdad	Coll. of Agric. of Univer . of Al-Muthanna
hasan_a_abd@yahoo.com	Sofiajasem2017@gamail.com

ABSTRACT

This Research was performed to evaluate the efficiency of Biofilm Biofertilizer produced from local isolates of *Pseudomonas fluorescens, Rhizobium leguminosarum*, with two addition technique include with bentonite as carrier and with liquid inoculant(without carrier) and three levels of chemical fertilizers (zero,25,50)% of recommendation on some growth and yield traits of wheat variety Rashid Biofilm Biofertilizer and chemical fertilizer affect significantly on all growth characters of wheat plant, Bentonite gave the best result compare with liquid inoculant, use of Biofilm Biofertilizer with 50% of the chemical fertilizer recommendation gave highest growth and yield of wheat Jlant .The duple Biofilm Biofertilizer of *P. fluorescens* plus *R. leguminosarum* was superior in enhancing growth and yield of wheat plant .The treatment of Biofilm Biofertilizer of *P. fluorescens* and *R. leguminosarum* with Bentonite and 50 chemical fertilizer recommendation gave45.89cm², 47.77 spad,70.00 gm,39.81mgN g⁻¹ 4.931mgP g⁻¹35.471mgK g⁻¹ of leaf area, chlorophyll content, weight of 1000 grains, Nitrogen, Phosphorus, Potassium uptake in the vegetative part respectively.of plant

Key words: Biofilm Biofertilizer, carrier, chemical fertilizer and wheat Part of Ph.D Thesis of second author

مجلة العلوم الزراعية العراقية -2018 :2019 :2016 -656 تأثير الغشاء الحيوي مع السماد الحيوي لبكتريا Pseudomonas fluorescens و Pseudomonas fluorescens و Rhizobium leguminosaru m ومستويات السماد الكيميائي وطريقة اضافة اللقاح في بعض صفات النمو و الحاصل للحنطة حسن علي عبد الرضا مدرس مساعد قسم علوم التربة والموارد المائية – كلية الزراعة –جامعة بغداد المستخلص

نفذت هذه التجربة لتقييم كفاءة السماد الحيوي المدعم بالغشاء الحيوي المنتج من عزلات محلية لبكتريا Pseudomonas fluorescens و من السماد الكيمياني هي (صفرو25و50) %من التوصية السمادية في بعض صفات النمو والحاصل لمحصول الحنطة صنف رشيد . ادى استعمال من السماد الحيوي المدعم بالغشاء الحيوي الى التأثير معنويا في جميع صفات النمو والحاصل لمحصول الحنطة صنف رشيد . ادى استعمال السماد الحيوي المدعم بالغشاء الحيوي الى التأثير معنويا في جميع صفات النمو المدروسة لنبات الحنطة ، ادى استعمال البنتونايت لإعطاء نتائج افضل عند استعماله كحامل للقاح مقارنة باللقاح السائل، ان استعمال السماد الحيوي مع غشاء الحيوي مع 50% من التوصية السمادية كان قد اعطى نتائج افضل لصفات النمو المدروسة، ادت اضافة السماد الحيوي المزدوج والغشاء الحيوي المتكون من 50% من التوصية السمادية كان قد اعطى نتائج المفضل لصفات النمو المدروسة، ادت اضافة السماد الحيوي المزدوج والغشاء الحيوي المتكون من 50% من التوصية السمادية كان قد اعطى نتائج الفضل لصفات النمو المدروسة، ادت اضافة السماد الحيوي المزدوج والغشاء الحيوي المتكون من 50% من التوصية السمادية كان قد اعطى نتائج المفسل لصفات النمو المدروسة، ادت اضافة السماد الحيوي المزدوج والغشاء الحيوي المتكون من 100% من التوصية السمادية كان قد اعطى نتائج و الفضل لصفات النمو المدروسة، ادت اضافة السماد الحيوي المزدوج والغشاء الحيوي المتكون من 100% معنوي المعنوي العرف و 1.30% معناء المعاملة في تحفيزه لنمو نبات الحنطة. اعطت معاملة السماد الحيوي ذي الغشاء الحيوي لبكتريا R. *الووساسي و 1.30%* التفوق هذه المعاملة في تحفيزه لنمو نبات الحنطة. اعطت معاملة السماد الحيوي ذي الغشاء الحيوي لبكتريا 4.30% م غر و 4.93% ملغم عنوي المعاملة في معاملة المسادية 4.30% ما حربة و النتروجين و الفسفور و البوتاسيوم المحص و 4.93% ملغم عنو علم عنوي المنوي المساحة الورقية ومحتوى الكلوروفيل ووزن الف حبة و النتروجين و الفسفور و البوتاسيوم المحص في الجزء الخضري للنبات بالتتابع.

كلمات مفتاحية: سماد حيوى، الغشاء الحيوى، حامل، اسمدة معدنية، حنطة.

البحث مستل من اطروحة الدكتوراه للباحث الثانى.

*Received:20/6/2017, Accepted:24/10/2017

INTRODUCTION

Agricultural development require optimum use of microorganisms activities to increase the available of nutrient for plants and this technique known as biofertilization which are an alternative, inexpensive and appropriate source for environment in comparison with chemical fertilizer the difficulty Lies in the spread of this technology is in enabling these microorganisms to survive effectively in the plant rhizosphere and soil (20). To solve this problem .researchers carried out manv extend the survival attempts to of microorganisms in soil through use different carriers but despite the remarkable success of this method they sought to find other possibilities to add biofertilizer by focusing on the importance of Biofilm Biofertilizer and developing strategies for the survival of microorganisms alive longer duration (16).Biofilm spread in different environments including soil and their presence, varies between nutrient rich rhizosphere soil to soil poor with nitrogen ,phosphorus , water and other nutrients (28). Different genera and species of bacteria are able to form biofilm on the surface of roots like Rhizobium and Pseudomonas fluorescens and the later play other important role in biocontrol and plant promoting growth Rhizobacteria (11).Pseudomonas fluorescens bacteria present in various plants rhizosphere and act as PGPR that lead to increasing nutrients (19). Rhizobia are important symbiotic nitrogen fixers which produce different hormones, growth factors and vitamins which stimulate seeds germination and increase photosynthesis pathways in addition to protect plant against diseases (15). The aim of this research was to evaluate the efficiency of **Biofilm Biofertilization** of local isolates of Pseudomonas fluorescens and Rhizobium leguminosarum with three levels of chemical fertilizer and two addation technique include with bentonite as carrier and liquid inoculant on some growth and yield traits of wheat (*Triticum asetivum L*).

MATERIALS AND METHODS Bacterial Biofilm Collection

Pseudomonas fluorescens and *Rhizobium leguminosarum* isolates, were tested previously it form biofilm and were grow in test tube contain 50ml of Tryptic soya broth and incubate at 28c°.Biofilm collected under control conditions and put in scrow tubes phosphate Buffer saline(PBS).

Modification of culture media

Different modification were carried out in the current study for KingB and Yeast Extract Manitol Agar medium in order to make it more suitable for the above bacterial isolates when they grow together.

Inoculation of Bentonite

Bentonite slurry suspension was prepared (20%w:v)using Distilled water in flasks and sterilized at 121c°,15b/in² for 20 minutes ,1ml of P. fluorescens inoculum grow in kingB broth medium was added to the first flask and other 1ml of each P.fluorescens and R. grow leguminosarum inoculum in the modified broth medium was added to the second flask ,then 100ml of 10% arabic gum solution was added to all flasks. wheat seeds were sterilized by ethyl alcohol 95% and soudium hypochloride 15% then washed repeatedly by distilled water, seeds were added to the flasks and incubated 72hr. at 28c°then it spreaded over cardboard, collected biofilm power was added to the inoculated seeds and lay aside for half an hour befor planting

Biological Experiment

biological experiment include The the following factors. The first Factor : Biofilm Biofertilizer, include three level denote as follow .P1 Biofilm **Biofertilizer** of Pseudomonas fluorescens,P3 **Biofilm** Biofertilizer of both *P.fluorescens* and *R*. .P0:control leguminosarum (without inoculation). The second factor :addation technique ,include two types denote M1: Biofertilizer with Bentonite as carrier and MO:

denoteto liquid inoculation without carrier. The Third Factor: chemical fertilizer ,include three levels denote F0: without addation, F1 addation of 25% of the recommendation and F2 addation of 50% of the recommendation. The biological factorial experiment was conducted using Complete Randomized Design with replicates experiment three .the unit $=3\times2\times3\times3=54$ plus three experiment units (complete chemical fertilizer recommendation without biofertilizer) so the total experiment units =57, soil were air dryad, 10kg of 4mm diameter of dry sieved soil were put in pots which sterilized previously from inside by ethyl alcohol Eight inoculated wheat seeds were planting in each pot at 24-11-2015, urea (46%N)was used as nitrogen fertilizer and added by two doses ,the first at planting and the other at elongation period with note that the recommendation was 160kgNh⁻¹,one dose of triple super phosphate (20%P)was added at planting note that the recommendation was 100kgPh⁻¹and one dose of potassium sulphate (50%k₂0)was added also at planting period note that it's recommendation was 100kgKh⁻ ¹.After one week of germination five plant only were leave for each pot ,plants were harvested at 28-4-2016. Chlorophyll content (spad) ,leaf area (cm²)weigh of 1000grian (g), Nitrogen, Phosphorus and Potassium uptake (mgg⁻¹)were estimate after harvesting .chemical ,physical and biological properties of pots soil were analyzed in the laboratory of soil Department .College of Agriculture -University of Al-Muthna. Data were analyzed as analysis of variance using a statistical analysis program Genstat while the means were compared using LSD0.05.

Table1. Some chemical, physical and	l biological characteristics of soil before planting
-------------------------------------	--

Adjective		The value
pH In the extract 1:1		7.2
EC	dsm ⁻¹	2.6
Ca ⁺²		1.15
Mg^{+2}		0.57
CO3 ²⁻		Nil
нсоз ⁻	Centimolle L ⁻¹	0.49
CL ⁻		0.98
SO4 ²⁻		1.27
Organic matter O.M(gm kg -1		1.5
N available mg kg ⁻¹		24.2
P available mg kg ⁻¹		11
K available mg kg ⁻¹		165
sand		110
silt	gm kg ⁻¹ soil	480
Clay		410
Soil texture		Silty Clay
Total bacteria		$10^{7} \text{ x} 1.4$
Total fungi	CFUg ⁻¹ dry soil	10 ⁵ x0.32

RESULTES AND DISCULATION

Flag leaf area

Results in Table 2 shows that inoculation with Biofilm Biofertilizer affect significantly in increasing leaf area in comparison with control uninoculuted treatment with the superiority of P3 treatment (37.69cm²)which represent increasing percentage 48.85% compare to control treatment, significant differences were obtained between types of Biofilm Biofertilizer ,P3gave the superiority with increasing %29.64% over P1 treatment ,these results agree with other Researchers (29) that they found significant effect of Biofilm Biofertilizer in increasing leaf area of tea plant and this increasing may due to the increasing in the availability of nutrients like nitrogen which play important role in protoplasm synthesis ,Enzymes ,co enzymes growth regulators and chlorophyll content(2).

According to Table 2 results which, indicates the effect of chemical fertilizer on leaf area ,F2 treatment gave increasing% of 27.74% in compare with control treatment ,these results conform with the results as Al-Haidere (6) when he obtained increasing in wheat leaf area with the increasing of nitrogen levels and this could be due to the role of nitrogen in activation of meristem cells and Auxins (17). Results also shows the superior of Bentonite carrier (M1 treatment) with 28.60cm²leaf area compare with liquid inoculant(M0 in treatment) that gave 27.66cm². The second order interaction shows the superiority of P3F2 treatment when compare with all others of leaf area 44.51cm² and this agree with the results Seneviratne et al (29), they found increase in leaf area after addation of 50% of chemical fertilizer recommendation and this may due to energy source and available phosphorus that increase the proliferation of microorganisms that form biofilm which produce more organic acids reflect in increasing the available nutrients and all these reflect on increasing leaf area (12). On the other hand P3M1 treatment gave leaf area mean of 38.83cm ¹compare with the lowest value of F0M1 treatment 22.83cm²while the statistical results shown in table 2 indicate that the triple interaction treatment P3F2M1 gave the highest value 45.89cm² of leaf area and this may due not to the increasing of nutrient only but to the protection of plant against different diseases (10).

Table 2. Effect of biofilm with biofertilizer, type of carrier and level of chemical fertilizer in
the area of the flag leaf area (cm ²)for wheat plant

Р		М	F	Levels of che	mical fer	tilizati	on kg h ⁻¹		
Type of bacter pollen	ial	Download metho	od	F1	F2		F3	P×M	Rate p
-		M0		17.43	18.87		20.35	18.88	
PO		M1		18.22	20.11		20.72	19.68	
	P× F			17.83	19.49		20.54		19.28
P1		M0		21.28	25.94		29.99	25.74	
P. fluorescen	s	M1		21.75	26.27		33.89	27.30	
	P×F			21.52	26.11		31.94		26.52
P3		M0		29.78	36.72		43.13	36.54	
P. fluorescen Rhizobium	s	M1		32.37	38.23		45.89	38.83	
	P× F			31.08	37.48		44.51		37.69
	RateH	7		23.48	27.69		32.33	RateM	
F+M		M0		22.83	27.18		31.16	27.06	
F +IVI		M1		24.11	28.20		33.50	28.60	
Fertilizer st	andard t	reatment 100%		50.71					
Р	F	Μ	P×F	P×M		F×M		P×F×M	L.S.D=0.05
0.614 0.	.614	0.501	1.0632	0.868		0.868		1.504	L.3.D=0.05

Chlorophyll content

The results were shows that addation of Biofilm Biofertilizer cause significant increase in Chlorophyll content of wheat leafs compare with control treatment.P3 treatment gave Chlorophyll content 46.68 spad (Table 3), with increasing % of 35.07% from control that gave 30.31 spad and this could be due to gibberellins that produce by microorganisms (33), furthermore double Biofilm the Biofertilizer cause increase in nitrogen in particular NH₄and increase root system (13). Table 3 indicates also the role of chemical fertilizer levels especially F2 treatment in obtainment the highest value for chlorophyll content, on the other hand P3F2 treatment gave the highest value 52.40spad and these results reflect the role of Biofilm Biofertilizer support growth regulators that enhance root growth (9)and increasing cells divisition and elongation (5). Concerning the interaction chemical and carriers, between F2M1 treatment was superiored in it's chlorophyll content that was 42.49spad while the third order interaction treatment P3F2M1 gave the highest value 53.50spad of chlorophyll content and this could be due to the role of Biofilm in enhancing the population of microorganisms in soil and rhizosphere more than the liquid inoculum alone (24),(32).

Table 3. Effect of biofilm with biofertilizer, type of carrier and level of chemical fertilizer in								
chlorophyl content of wheat plant(spad).								
Р								

P Type of bacte	rial pollen	M Download method F Levels of chemical fertilization kg h^{-1}				P×M	Rate p
			F1	F2	F3		
		M0	28.56	30.30	31.37	30.08	
P0		M1	28.84	31.02	31.77	30.54	
	P× F		28.7	30.66	31.57		30.31
P1		M0	36.07	38.00	40.61	38.23	
P. fluore	scens	M1	36.76	38.47	42.21	39.15	
	P×F		36.42	38.24	41.41		38.69
Р3		M0	40.05	45.42	51.30	45.59	
P. fluorescens	Rhizobium	M1	41.49	48.33	53.50	47.77	
	P× F		40.77	46.88	52.40		46.68
	RateF		35.30	38.59	41.79	RateM	
F+N	r	M0	34.89	37.91	41.09	37.96	
F +N	1	M1	35.70	39.27	42.49	39.15	
Fertili	izer standard tı	eatment 100%	59.38				
Р	F	M P	×F P×M	F × M	1	P×F×M	L.S.D=0.05
0.598	0.598	0.488 1.	036 0.846	0.84	6	1.465	L.S.D=0.05

Weight of 1000 grains(gm).

Results in the Table 4 indicates that P3 treatment gave the highest value of 1000grains (55.88g) with increasing %59.36% compare with control treatment that gave 22.71g and these could be due to the role of Biofilm in increasing the efficiency of microorganisms to produce growth hormones (18) which effect on cell division and elongation that increase the surface area of roots and this reflect on more uptake of nutrients which gave highest weight of grains (14). Results showed the superiority of F2 level of chemical fertilizer in increasing 29.57% compare to control and this may be due to the role fertilizer in the balance of nutrients and carbohydrate ,protein content of cells (21). Table 4 indicates that M1 treatment gave 39.67g of the Weight of 1000 grain over control and this may due to the high cation exchange capacity of Bentonite(16)which help to survive of bacteria for long time in rhizosphere (4).On the other hand results showed the superiority of P3M1 treatment when it record the highest value 58.90g and this could be due to the efficiency of microorganisms in this carrier that reflect to more profelration and production of growth promoting substance (7)while the triple interaction treatment P3F2M1 gave the best results and the Weight achieved 70.00g and this may be due to the contribution of this treatment in acceleration of female flowering as a result of increasing in synthesis of metabolite that affect on more nutrients uptake and full grain (3,)(30),(34).

Table 4. Effect of biofilm with biofertilizer, type of carrier and level of chemical fertilizer in Weight of
1000 grain for plant wheat

	100	oo gram tor pie	ant wheat			
Р	M Download metho	F Levels of	chemical fertiliza	tion kg h ⁻¹	DVM	Data n
Type of bacterial pollen	M Download metho	F1	F2	F3	P×M	Rate p
P0	M0	17.16	21.86	28.00	22.34	
10	M1	17.44	23.57	28.21	23.07	
P×1	F	17.30	22.72	28.11		22.71
P1	M0	29.77	31.17	37.00	32.65	
P. fluorescens	M1	32.50	35.60	43.00	37.03	
P×I	F	31.14	33.39	40.00		34.84
P3	M0	43.30	53.30	62.00	52.82	
P. fluorescens Rhizobium	M1	48.70	58.00	70.00	58.90	
P×1	F	46.00	55.65	66.00		55.88
Rate	F	31.48	37.25	44.70	RateM	
F+M	M0	30.08	35.44	42.33	35.95	
1, ±111	M1	32.88	39.06	47.07	39.67	
Fertilizer standard	treatment 100%	77.12				
P F	M I	P×F P×M	F×M	P×F	×M	L.S.D=0.05
1.111 1.111	1 0.907 1	.925 1.571	1.571	2.7	22	L.S.D=0.05

Nitrogen uptake

Results in Table 5 shows that Biofilm Biofertilizer cause significant effect on the amount of nutrient uptake by wheat plant ,The P3 treatment record the value 29.20 mg N g⁻¹ with increasing %61.17% over control which was 11.18mg Ng⁻¹ and this could be due to the role of biofilm in regulation of growth promoter production like IAA and gibberelins which contributed in increase the growth of root system (9) and this in turn the increasing of nitrogen uptake and other nutrient ,on the other hand addation of chemical fertilizer increase N uptake with the superioty of F2 that gave 24.77mg Ng⁻¹with treatment increasing %of 38.99%over control treatment while M1 treatment record 20.58mgNg⁻¹at an 9.86% compare to increasing rate **M**0 treatment. The Third order($P \times M \times F$) interaction treatment shows that the treatment P3F2M1 gave the highest nitrogen uptake to 39.81mg Ng⁻¹ and this may due to the role of the combination of biofilm with Bentonite

Table 5. Effect of biofilm with biofertilizer, type of carrier and level of chemical fertilizer
in Weight amount of nitrogen uptake mg Ng ⁻¹ by wheat plant

Р	M Download	F Levels of	chemical fert	ilization		
—			kg h ⁻¹		P×M	Rate p
Type of bacterial pollen	method	F1	F2	F3		-
	M0	10.35	11.11	11.79	11.08	
P0	M1	10.39	11.28	12.13	11.27	
P× F		10.37	11.20	11.96		11.18
P1	M0	12.41	16.04	22.00	16.82	
P. fluorescens	M1	13.59	18.90	26.99	19.83	
P×F		13.00	17.47	24.50		18.32
P3	M0	21.25	26.14	35.86	27.75	
P. fluorescens Rhizobium	M1	22.65	29.47	39.81	30.66	
P× F		21.95	27.81	37.84		29.20
Rate	<u>?</u>	15.11	18.83	24.77	RateM	
T M	M0	14.67	17.76	23.22	18.55	
F+M	M1	15.54	19.88	26.31	20.58	
Fertilizer standard t	reatment 100%	53.874				
P F	M P>	F P×M	F×M	P×	F×M	I S D_0.05
2.387 2.387	1.949 4.1	35 3.376	3.376	5.	.847	L.S.D=0.05

Phosphorus uptake

Results in Table 6 shows the effect of Biofilm Biofertilizer on Phosphorus uptake by plant, P3 treatment gave 3.44mg Pg⁻¹with increasing %75.58% compare to control treatment and this could be due to the role of Biofilm on increasing plant hormons that promote plant and root growth which contributed in Phosphorus uptake (9). The second level F2 of chemical fertilizer gave 2.94 mg Pg⁻¹ with increasing %52.04% compare to control F0 treatment. Bentonite carrier (M1 treatment) gave 2.28mg Pg⁻¹with increasing %12.60% over M0 treatment. Results showed that Biofilm Biofertilizer participate in reduction of chemical fertilizer to 50%, so P3F2 treatment gave 4.68 mg Pg⁻¹ and these results agree with Seneviatne et al(29)whom

found that Biofilm Biofertilizer reduce using of chemical fertilizer to half and this results lead to the true that integration in fertilization by addition of 50% Of recommendation with biofertilizer gave better results in comparison with addition of total chemical fertilizer recommendation with low economic costs(7). Results showed that the third interaction highest treatment P3F2M1 gave the phosphorus uptake 4.93 mgPg-1 and this may due to the role of Biofilm acidity as a results of IAA and in increasing Organic acid that increase the availability of phosphorus(18), and these results come in agreement with Igual et al(23), found that using of microorganisms with chemical fertilizer reduce the amount need to be add to 50% during dissolving the orecipitate(25).

•	amount of Phos	phorus upra	ike ing i g	by whe	ai piani	
P Type of bacterial polle	M Downloa n method		Levels of che ertilization k F2		P×M	Rate p
	M0	0.52	0.79	1.14	0.82	
P0	M1	0.56	0.87	1.17	0.87	
P×	F	0.54	0.83	1.16		0.84
P1	M0	1.21	1.81	2.54	1.85	
P. fluorescens	M1	1.39	2.13	3.44	2.32	
P×	F	1.30	1.97	2.99		2.09
P3	M0	2.25	3.05	4.43	3.24	
P. fluorescens Rhizobiu	m M1	2.53	3.47	4.93	3.64	
P×	F	2.39	3.26	4.68		3.44
Rat	æF	1.41	2.02	2.94	RateM	
T. M	M0	1.33	1.88	2.70	1.97	
\mathbf{F} + \mathbf{M}	M1	1.49	2.16	3.18	2.28	
Fertilizer standard	l treatment 100%	7.18				
P F	Μ	P×F 1	P×M I	F×M	P×F×M	L.S.D=0.05
0.246 0.24	6 0.201	0.426).348 0).348	0.628	L.S.D=0.05

Table 6 . Effect of biofilm with biofertilizer, type of carrier and level of chemicalfertilizer in
amount of Phosphorus uptake mg Pg ⁻¹ by wheat plant

Potassium uptake

Results showed the superiority of the treatment received Biofilm Biofertilizer that in Potassium uptake compare to non-inoculated seeds P3 treatment gave 24.33mg Kg⁻¹and this may due to the role of Biofilm in increasing nitrogen which caused the synthesis of various compounds in the green part of plant that need Potassium for different metabolism pathway which lead to adsorb more Potassium from the soil. On the other hand addition of 50% of chemical fertilizer recommendation increase the K uptake to 70.65 mgKg⁻¹ and this could be due to the increasing of it's availablity in the soil (1)in addition inoculated of Bentonite as carrier increase K uptake to 14.35 mgKg⁻¹with

increasing%14.42%. The results showed also that the interaction treatment P3F 2 gave 32.98 mgKg⁻¹ and this superiority may due to the role of Biofilm Biofertilizer to enhance the Potassium from its unavailable source due to the production of different organic acids (22). The Third order interaction treatment P3F2M1 showed the superiority in comparison with all other treatment when it gave 35.47 mgKg⁻¹. This results shows the role of this treatment to prepare good environment for microorganisms in addition to protect its against different ecological stress (27),(28) and enhance the root system to absorb more Potassium from fertilizer or unavailable source (10).

Table7 . Effect of biofilm with biofertilizer, type of carrier and level of chemical fertilizer in
Weight amount of Potassium uptake mg Kg ⁻¹ by wheat plant

				I.	88	~ ,	Firefore Presso		
Р		M Downloa	d	F Levels of chemical fertilization kg					
P Type of bacterial pollen		M Download	ia		h ⁻¹		P×M	Rate p	
		method		F1	F2	F3		_	
		M0		4.56	5.07	5.95	5.93		
PO		M1		4.71	5.32	6.12	5.38		
$\mathbf{P} \times \mathbf{F}$				4.64	5.20	6.04		5.29	
P1		M0		6.48	8.60	12.01	9.03		
P. fluorescens	5	M1		8.17	10.84	15.83	11.61		
P×F				7.33	9.72	13.92		10.32	
P3		M0		15.45	21.93	30.49	22.62		
P. fluorescens Rhiz	obium	M1		17.95	24.69	35.47	26.04		
P×F				16.70	23.31	32.98		24.33	
	RateF			9.56	12.74	17.65	RateM		
\mathbf{F} + \mathbf{M}		M0		8.83	11.87	16.15	12.28		
		M1		10.28	13.62	19.14	14.35		
Fertilizer star	ndard trea	tment 100%		42.83					
Р	F	Μ	P×F	P×M	F×M		P×F×M	L.S.D=0.05	
1.578	1.578	1.288	2.733	2.231	2.231		3.865		

REFERENCES

1.Abu Dahi,Y.M and M.A.Al-younis .1988 .Plant Nutrition Guide .Ministry of Higher Education and Scientific Resear .pp:411 2.Abu-Nuqta. F, and M.S. AL-Shaatir .2011. Soil fertility and Fertilization. The Theoretical Part. Publications of Damascus Univer, Faculty of Agric .pp:120

3.Al-Bahrane ,E.K.M.2015.Effected of Phosphate Dissolving Bacteria and Humic Acid on Phosphate Equilibrium Nutrient Availability and Yield of Corn (*Zea mays L.*).Ph.D. Dissertation . Coll of Agric –Univer of Baghdad pp: 155

4.Al-Dulami .M.A.G,2004.Isolation and Diagnosis of *Pseudomonas fluorescens* and *Pseudomonas putida* from Soil and Test its Efficiency in Biocontrol . M.S.c. Thesis . Dept. Biology .Coll. Science–Univer of. Anbar pp:

5.Akbari ,P.,Ghalavand, Modarres A.M. Sanavy , and M.Agha Alikhani.2011.The effect of biofertilizers ,nitrogen fertilizer and farmyard manure on grain yield and seed quality of sunflower (*helianthus annus L*).J. Agricu. Techn. 7(1):173 -184.

6.Al –Haidere ,H.KM.A .2003.Effect of Times of Addition of Levels of Nitrogen and Means of Sowing in Growth and Yield of Wheat(*Triticum asetivum. L*).Ph.D Dissertation . Dept. Soil Science and Water Resource.Coll., of Agric. Univer. Baghdad pp:122.

7.Al-Keleel S.M.A.2011. Effected of Integration Between Mineral –Organic and Biofrtilizer on Yield of Tomato (*Lycopersicon esculentum*) in Plastic House .M.Sc. Thesis Dept. Soil Science and Water Resource.Coll., Agric. Univer. Baghdad pp:110.

8.Al-Marjanin.A.H.F.2005. Effect Ground Addition Level by NPK and Spraying on Growth and Yield Wheat (*Triticum asetivum*. *L*). M.Sc. Thesis . Dept. Soil Science and Water Resource.Coll., Agric. Univer. Baghdad pp:117. 9.Bandara W.M.M.S, G Seneviratne and, S.A. 2006 Kulasooriya. Interactions among endophytic bacteria and fungi: effects and J Biosci 31:645-650. potentials. 10.Bashan, Y. and L.E. de- Bashan. 2005.Fresh weigth measurement of root provide inaccurate estimates of the effects of plant growth- promoting bacteria on root growth :a critical examination .Soil Biology and Biochemistry 37:1795-1804.

11.Bianciotto V, S Andreotti, R. Balestini, P. Bonfante and S. Perotto. 2001. Mucoid mutants of the biocontrol strain *Pseudomonas fluorescens CHA0* show increased ability in *biofilm formation* on mycorrhizal and *nonmycorrhizal* carrot roots. Mol. Plant-Microbe Interact. 14:255–60.

12.Browning M, D. B Wallace, C. Dawson, S. R. Alma and J. A. Amador .2006 .Potential of butyric acid for control of soil-borne fungal pathogens and nematodes affecting strawberries; Soil Biol. Biochem. 38 401-404. 13.Buddhika U.V.A., G. Seneviratne and C.L. Abayasekara.2012. Biofilmed Biofertilizers for Maize (Zea mays L.): Effect on Plant Growth under Reduced Doses of Chemical Fertilizers. Proceedings of the Abstracts of International Jaffna University Research Conference (JUICE-2012).

14.Cornejo ,Hexon Angel Contreras ,Lourdes Macl´as_Rodr´guez,Carlos Corte´s _Penagos ,and Jose´ Lo´ pez_Bucio.2009.Trichoderma virens ,a plant beneficial fungus ,enhances biomass production and promotes lateral root growth through an auxin-dependen mechanism in arabidopsis1 .American Society of Plant Biologists .Plant Physiology ,March ,61.149:1579-1592.

15.Dakora ,F.D.2003.Defining new roles for plant and rhizobial molecules in sole and mixed plant cultures involving symbiotic legumes New Physiologist. 158(1):39-49. 16.Gilbert, P.; J. Das and I Foley,.1997 Biofilms susceptibility to antimicrobials. Adv. Dent. Res. 1997, 11, 160–167. 17.Hassan .N.A,H.U .Al-Dulemi and L.A.Al-Ethawi 1990 Soil Fertility and Fertilizer .Al-Hakma pupil .pp:140

18.Jayasinghearachchi H.S. and Seneviratne G. 2004a. Can mushrooms fix atmospheric nitrogen? Journal of Biosciences 23, 293–296. 19.Klopper,J.W., ,M.N Schroth. and T.D Miller. 1988. Effect of rhizosphere colonization by plant growth-promoting rhizobacteria on potato plant development and yield phytopathol. 70 : 1078-1082.

20.Klopper J. W. R Lifshitz. and R. M Zablotowicz. 1989. Free-living bacterial inocula for enhancing crop productivity. Trends in Biotechnology. 7: 39-44.

21.Khairo .O.M.2003.Effect of Integral Spraying with Nitrogen and Potassium in Growth and Yield of Corn (*Zea mays L.*) .M.Sc. Thesis . Soil Science and Water Resource. Coll., Agric. Univ. Baghdad.pp:135 22.Kumar .F.2010 .A lecture about plant Biofrtilizer ,the pertect replacement for chemical fertilizer .General authority for Agriculture Kuwait. Al –Gabas News paper .No.13464

23.Igual, J.M.; A. Vqlverde; E. Eervantes; **E.**Cervartes and E. Velazquez. 2001. Phosphate solubilizing bacteria as an inoculants for agriculture: use of updated molecular techniques in their study. Agronomie, 21 : 561-568.

24.Monier J.M, and S.E Lindow.2003. Differential survival of solitary and aggregated bacterial cells promotes aggregate formation on leaf surfaces. Proc Natl Acad Sci USA, 100:15977-15982.

25.Pradhan, N.L .B.Sukla .2005.Solubilization of inorganic phosphate by fungi isolated from agriculture soil .African J. Biotechnol,5:850-854.

26.Seneviratne G. and I.K. Indrasena. 2006. Nitrogen fixation in lichens is important for improved rock weathering. J. Biosciences 31, 639–643. 27.Seneviratne G., Zavahir, J.S. W.M.M.S.Bandara and M.L.M.A.W Weerasekara. 2007. Fungal-bacterial biofilms: their development for novel biotechnological applications. World J. Microb. and Biotech 24(6), 739–743.

28.Seneviratne G .2008 Biological nitrogen fixation: potential biotechnological applications beyond biofertilizers. Curr Sci 95:107.

29.Seneviratne G, Thilakaratne RMMS, APDA. Javasekara Seneviratne KACN, Padmathilake KRE, De Silva MSDL.2009. Developing Beneficial Microbial Biofilms on Roots of Non-Legumes: a Novel Biofertilizing Technique. In: Khan MS, Zaidi A, Musarrat J. editors. Microbial Strategies for Crop Improvement. Berlin Heidelberg; Springer-Verlag. pp: 51–62.

30.Turan ,M.;N .Ataogh ,and F.Sahin .2006.Evaluation of the capacity of phosphate solubilizing bacteria and fungi on different form of phosphate in liquid culture .Sustuinable Agricultural ,28:99-108.

31.Van de Mortel, M.and L.J Halverson, 2004 Cell envelope components contributing to biofilm growth and survival of *Pseudomonas putida* in low-water-content habitats. Mol. Microbiol., 52, 735–728.

32.Vessey JK 2003 Plant growth promoting rhizobacteria as fertilizers. Plant Soil 255:571–586.

33.Weiss,D,;AJ.Van Tunen ,A.H.Halevy ;J.N.M.Mol and A.G.M. Gerats.1990 .Stamens and gibberellic acid in the regulation of flavonoid gene expression in the corolla of petunia hybrid .plant Williams and Wilkins,U.S.A .94.511-515..

34.Zaidi, A.and M. S Khan, 2006.Co-Incoulation effects of phosphate solubilizing micro-organisms and glomus fasciculatumongreen gram bradyrhizobium symbiosis Turk .J.agric.: 30:223-230.