

## EFFECT OF SELENIUM AND MANGANESE OXIDE AND THEIR COMBINATION IN INDUCING RESISTANCE IN PEPPER PLANTS INFECTED WITH PATHOGENIC FUNGUS *FUSARIUM SPP.*

Baraa A. A. Aljubory  , Mena W. Hatem    
Dept. Plant Prot., Coll. Agric. Eng. Sci., University of Baghdad

### ABSTRACT

This experiment was conducted in a greenhouse belonging to the Department of Plant Protection, College of Agricultural Engineering Sciences, University of Baghdad - Jadiriya during the 2023-2024 growing season. The aim was to study the effect of selenium and manganese oxide in inducing resistance in pepper plants (*Capsicum annuum*) against the pathogenic fungus *Fusarium solani* while enhancing certain growth parameters. The results revealed that applying these elements in combination at the optimal concentration of 200 mg/L inhibited fungal growth in vitro by 77.71% compared to the pathogenic fungus control, which showed 0% inhibition. This treatment also improved growth parameters such as plant height, fresh weight, and dry weight in pot experiments. Moreover, it reduced disease severity and infection rate in both pot and field experiments, in addition to enhancing the activity of enzymes such as peroxidase, Phenylalanine ammonia-lyase (PAL), and catalase in pepper plants under field conditions.

**Key words:** Se, Mn<sub>2</sub>, *F. solani*, *Capsicum annuum*.



Copyright© 2025. The Author (s). Published by College of Agricultural Engineering Sciences, University of Baghdad. This is an open-access article distributed under the term of the Creative Commons Attribution 4.0 International License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cite.

**Received: 22/11/2024, Accepted: 16/2/2025, Published: 30/5/2026**

### INTRODUCTION

The pepper plant (*Capsicum annuum* L.) is one of the economically significant main crops, rich in various vitamins, minerals, carbohydrates, and other nutrients. It is cultivated in several countries worldwide and belongs to the Solanaceae family (Ismail *et al.*, 2011). Pepper plants are affected by several soil-borne pathogens, the most notable of which are those causing wilt and seedling damping-off (Al-aamel & Al-maliky, 2023). Various factors contribute to wilt in peppers, leading to significant losses, including the pathogen *Fusarium spp.* (Al-Rubaie, 2022). Among the primary causes of wilt and root rot diseases affecting numerous crops are the fungus *Fusarium solani* (Avery *et al.*, 2019). Abiotic elements can be utilized to control certain pathogens and enhance plant resistance, thereby mitigating the negative environmental impacts of chemical pesticides (Hatem & Ali, 2023). Selenium is an essential

micronutrient for plants, known to enhance their resistance to various environmental stresses such as drought, salinity, high temperatures, and bacterial and fungal pathogens. It also boosts the activity of antioxidant enzymes in plants (Schiavon *et al.*, 2017). Selenium can be applied to plants through various methods, including soil application (Hussein *et al.*, 2019). Studies have demonstrated that selenium application promotes plant growth and productivity (Bano *et al.*, 2021). Research has shown that selenium at a concentration of 20 mg/kg inhibited the growth of *F. proliferatum* isolated from rice in vitro (Troni *et al.*, 2021). Additionally, a study revealed that selenium in the form of sodium selenate at a concentration of 120 ppm reduced the growth of *Fusarium spp.* by 87.14% (Özer *et al.* 2024). Manganese is an essential element in plant fertilization, enhancing plant nutrition, photosynthetic efficiency, and crop productivity (Mousavi *et*

al., 2007). A study demonstrated that manganese oxide improved grain production through foliar spraying ( $3.74 \text{ mg ha}^{-1}$ ) and seed soaking ( $3.57 \text{ mg ha}^{-1}$ ), showing effectiveness in increasing productivity and biological fortification of bread wheat grains in alkaline soils (Ullah *et al.*, 2018). Increasing manganese levels in the soil stimulates plant growth and reduces the impact of root pathogens due to its significant role in enzyme-catalyzed reactions (Schmidt & Husted, 2019). Given the severe damage caused by seed rot and seedling damping-off diseases to pepper seedlings worldwide, this study aimed to induce systemic resistance in plants against damping-off disease using environmentally safe substances to mitigate disease risks and reduce associated losses. This was achieved by applying selenium and manganese oxide individually or in combination to combat the pathogen and enhance certain growth parameters. The study also aimed to analyze the biochemical reactions by measuring the activity of enzymes such as PAL (Phenylalanine ammonia-lyase), peroxidase, and catalase in plants.

$$\text{Inhibition Percentage} = \frac{\text{Mean Colony Diameter in Control} - \text{Mean Colony Diameter in Treatment}}{\text{Mean Colony Diameter in Control}} \times 100$$

## 2. Testing the Susceptibility of Pepper Cultivars to *Fusarium solani* Infection:

This experiment was conducted in the greenhouse of the Department of Plant Protection, College of Agricultural Engineering Sciences, University of Baghdad. The susceptibility of three pepper cultivars (Charisma, King Green, and Key Man) to the pathogenic fungus *F. solani* was tested. The

The severity of infection was evaluated using a 5-grade disease index, adapted slightly from Sun *et al.* (24), as follows:

- 0 = Healthy plant with no symptoms.
- 1 = Less than 25% of the roots showing light brown discoloration.
- 2 = 25–50% of the roots showing dark brown discoloration.

$$\text{Infection Severity Percentage} = \frac{\sum(\text{Disease Grade} \times \text{Number of Plants in Grade})}{\text{Maximum Disease Grade} \times \text{Total Number of Examined Plants}} \times 100$$

## MATERIALS AND METHODS

### 1. Testing the Effect of Selenium and Manganese Oxide at Three Concentrations on the Inhibition of *Fusarium solani* Growth In Vitro:

Three concentrations of selenium and manganese oxide (100, 150, and 200 mg/L) were tested, both individually and in combination, against *Fusarium solani*, which had been morphologically and molecularly identified. The optimal concentration was selected for subsequent experiments. These concentrations were applied to PDA (Potato Dextrose Agar) medium in three 9 cm Petri dishes for each treatment. The center of each dish was inoculated with a 0.5 cm diameter disc of *F. solani* mycelium from a 6-day-old culture. The control treatment consisted of PDA medium inoculated with the pathogenic fungus disc without any additives. The plates were incubated at  $25 \pm 2^\circ\text{C}$ , and after six days, the percentage of fungal inhibition was calculated using the following formula (Kim *et al.*, 2012):

cultivars were planted in 2 kg sterilized pots containing a sterile soil mixture combined with peat moss in a 2:1 (w/w) ratio. The pathogenic fungus inoculum was applied to the pots (50 mL per pot) at a concentration of  $1 \times 10^6$  spores  $\text{ml}^{-1}$ . Five 40-day-old pepper seedlings were transplanted into each pot. After 30 days, the infection rate was calculated using the following formula:

$$\text{Infection Percentage} = \frac{\text{Number of Infected Plants}}{\text{Total Number of Plants}} \times 100$$

3 = 50–75% of the roots showing dark brown discoloration, including discoloration of the crown area.

4 = 75–100% of the roots showing dark brown discoloration, with crown discoloration and complete plant death.

The percentage of infection severity was calculated using McKinney's formula:

**3. Evaluation of the efficiency of the optimal concentration of materials in greenhouse pot experiments:** This experiment was conducted in the greenhouse of the Department of Plant Protection, College of Agricultural Engineering Sciences, University of Baghdad. A mixture of soil was sterilized using an autoclave at a temperature of 121°C and a pressure of 1.5 kg cm<sup>-2</sup> for 20 minutes, repeated twice with a 48 hour interval between sterilizations. The sterilized soil was mixed with peat moss in a 2:1 ratio (w/w) and placed in 1 kg pots. The pots were planted with seeds of the pepper cultivar Charisma, which was identified as the most susceptible to infection

by the pathogenic fungus *F. solani*. Three seeds were sown per pot. Treatments were applied to the soil, and three days later, the fungal inoculum was added at a concentration of 1×10<sup>6</sup> spores ml<sup>-1</sup>, except for the control treatments, which received only distilled water without the fungus. The most effective concentration identified in the in vitro inhibition experiments (200 mg L<sup>-1</sup>) was used. The materials were applied at a volume of 30 ml per pot. After 30 days of adding the fungal inoculum, plant height, fresh weight, dry weight, and the percentage and severity of infection were measured (Table 1).

**Table 1. Treatments used in pot experiments and their concentrations**

No.	Treatment	Additive material Conc. mg l <sup>-1</sup>
1	Control (Plant only)	0
2	Control (Pathogenic fungus only)	0
3	Fungus + Selenium	200
4	Fungus + Manganese Oxide	200
5	Fungus + Selenium + Manganese Oxide	100+100
6	Selenium	200
7	Manganese Oxide	200
8	Selenium + Manganese Oxide	100+100
9	Chemical pesticide Uniform	2.5

**4. Evaluation of the efficiency of the best treatments in field pot experiments:**

Six treatments were selected from the pot experiments that demonstrated effectiveness in controlling *F. solani* infection in pepper plants. The field experiment was conducted in one of the greenhouses of the Plant Protection Department of the College of Agricultural Engineering Sciences, University of Baghdad, Jadiriya, during the 2023 winter growing season. The land was plowed, finely tilled, and divided into five blocks with furrows, maintaining a distance of 100 cm between each furrow. The experiment followed a randomized complete block design (RCBD) with three replicates per treatment. Pepper

seedlings with three true leaves were planted at 40 cm intervals. A 3 cm diameter circle, 10 cm deep, was created around each seedling, into which 100 mL of the treatments were applied, except for the control treatments (with and without the fungus). The treatments were applied at a concentration of 200 mg L<sup>-1</sup>. Three days after treatment application, fungal inoculum (spore suspension) was added to the circles around the seedlings at a concentration of 1×10<sup>6</sup> spores ml<sup>-1</sup>, with 50 mL of the suspension applied per seedling. For the control treatment, only sterile distilled water was added without the pathogenic fungus. The treatments used in the experiment are detailed in Table (2).

**Table 2. Treatments used in field experiments and their concentrations**

No.	Treatment	Conc. mg l <sup>-1</sup>
1	Control (Plant)	0
2	Control ( <i>Fusarium solani</i> )	0
3	Se + <i>F. solani</i>	200
4	MnO <sub>2</sub> + <i>F. solani</i>	200
5	MnO <sub>2</sub> + Se + <i>F. solani</i>	100+100
6	Uniform + <i>F. solani</i>	2.5

**5. Evaluation of the materials' effectiveness on enzyme activity in plants:** The activity of the enzymes PAL (Phenylalanine Ammonia-Lyase), Peroxidase, and Catalase was assessed in the laboratory of the Environmental and Water Directorate, Ministry of Science and Technology. Pepper leaves were collected 7, 14, and 21 days after the treatments were applied. The third leaf from the growing apex of 10 plants per replicate was sampled, placed in polyethylene bags, stored in a cooled container, and transported to the laboratory. The enzymes were analyzed according to standard laboratory procedures. Peroxidase activity was determined following the methods of Al-Sufi (2001) and Hamid (2002). PAL activity was measured using the method described by Al-Jarrah (2011), while catalase activity was assessed based on the method of Kato and Shimizu.

**Calculation of infection percentage and severity:** Five plants were randomly uprooted from each replicate after 60 days to calculate the infection severity. The roots were thoroughly washed with running water and dried. The infection percentage and severity were calculated as described in section (2).

**6. Statistical analysis:** Statistical analysis was conducted using GenStat program, using the completely randomized design (CRD) in the laboratory and greenhouse experiment, while using the completely randomized block design (RCBD) in implementing the plastic house

experiment. The averages were compared according to the least significant difference (LSD) at the probability level (0.05).

## RESULTS AND DISCUSSION

**1. The effect of selenium and manganese oxide on the inhibition of *F. solani* growth in vitro:** The results, as shown in Table (3), indicate that the combined treatments of selenium and manganese oxide at a concentration of 200 mg L<sup>-1</sup> provided the best inhibition of the pathogenic fungus *F. solani*. The percentage of inhibition in the combined treatment (Se + MnO<sub>2</sub> + *F. solani*) at the highest concentration was 77.17%. The treatment with manganese oxide (MnO<sub>2</sub> + *F. solani*) resulted in 75.13% inhibition, while selenium treatment (Se + *F. solani*) achieved 76.43%. The chemical fungicide, used as a comparison, showed the highest inhibition rate of 100%. Based on these results, the concentration of 200 mg L<sup>-1</sup> was selected as the optimal treatment for application in greenhouse and field experiments (Figure 1). These results align with those reported by Mao *et al.*, (2020), who found that ordinary selenium at a concentration of 20 mg/L inhibited *Fusarium graminearum*, the causal agent of wheat blight. Similarly, selenium at the same concentration inhibited *Fusarium proliferatum* in vitro. Eskandari *et al.*, (2020) also demonstrated that ordinary manganese contributed to reducing cotton plant infections caused by *Fusarium spp.*

**Table 3. Effect of different concentrations of selenium and manganese oxide on the inhibition of *F. solani* growth in vitro on PDA medium**

No.	Treatment	Concentration mg L <sup>-1</sup>	Fungus inhibition %
1	Control ( <i>Fusarium solani</i> )	0	0
2	Se + <i>F. solani</i>	100	73.67
3	Se + <i>F. solani</i>	150	74.4
4	Se + <i>F. solani</i>	200	76.43
5	MnO <sub>2</sub> + <i>F. solani</i>	100	73.1
6	MnO <sub>2</sub> + <i>F. solani</i>	150	74.4
7	MnO <sub>2</sub> + <i>F. solani</i>	200	75.13
8	Se + MnO <sub>2</sub> + <i>F. solani</i>	100	73.83
9	Se + MnO <sub>2</sub> + <i>F. solani</i>	150	74.97
10	Se + MnO <sub>2</sub> + <i>F. solani</i>	200	77.17
11	Uniform + <i>F. solani</i>	2.5	100
LSD 0.05			1.237

Each number in the table represents the mean of 3 replicates.

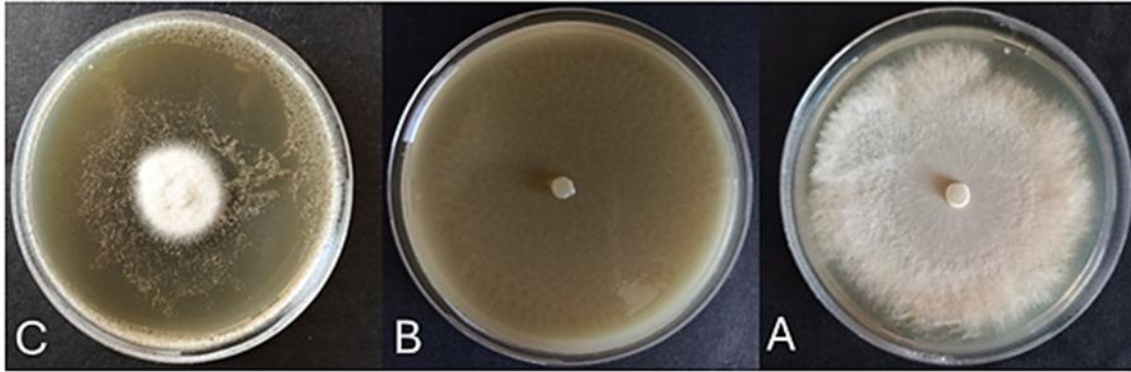


Figure 1. inhibition zone treatment: A: *Fusarium solani*, B: Chemical Fungicide Uniform, C: Combined Treatment at 200 mg/L

**2. Testing the Susceptibility of Pepper Cultivars to *F. solani* Infection:** The results for the infection percentage and severity caused by the fungal pathogen on three pepper cultivars indicated that the cultivar Charisma was significantly more susceptible than the other cultivars. The infection severity for Charisma was 84.6%, showing a substantial difference. On the other hand, the cultivars

Key Man and King Green did not exhibit a significant difference in infection severity, which was 41.7% and 35.4%, respectively. However, the infection percentage was uniform across all cultivars, at 100%, as shown in Table (4). Therefore, the Charisma cultivar was selected for use in subsequent experiments.

Table 4. Susceptibility of some pepper cultivars to the pathogenic fungus *F. solani*

No.	Cultivars	Infection severity	Infection percentage
1	Charisma CA	84.6	100
2	King Green KG	35.4	100
3	Key Man KM	41.7	100
	L.S. D <sub>0.05</sub>	12.49	

Each number in the table represents the mean of 3 replicates

**3. Evaluation of the efficiency of the optimal concentration of materials in greenhouse pot experiments:** The results of adding the optimal concentration of materials in the pot experiments demonstrated a significant improvement in several growth parameters. The plant height measurements showed that all treatments exhibited significant differences compared to the control treatments (plant and *F. solani* only). The highest average plant height was observed in the combined treatment without the fungal pathogen (MnO<sub>2</sub> + Se), with an average height of 32.61 cm. This was followed by the combined treatment with the pathogen (MnO<sub>2</sub> + Se + *F. solani*) and manganese oxide without the pathogen, both with an average height of 31.13 cm, as detailed in Table (5). For fresh weight, all treatments

significantly differed from the control treatments. The highest average fresh weight was recorded for the combined treatment without the fungal pathogen (MnO<sub>2</sub> + Se), with an average fresh weight of 12.32 g. Other treatments followed with close averages, as shown in Table (5). For dry weight, the treatments (MnO<sub>2</sub> + Se without the fungal pathogen, Uniform + *F. solani*, and MnO<sub>2</sub> + Se + *F. solani* with the pathogen) showed significant differences compared to the control treatments (plant only and *F. solani* only). Their average dry weights were 4.6 g, 4.33 g, and 4.1 g, respectively. Additionally, all treatments demonstrated significant differences compared to the fungal pathogen control, as shown in Table (5).

**Table 5. Effect of selenium, manganese oxide, and their combination on growth parameters (plant height, fresh weight, and dry weight) of pepper plants under greenhouse conditions (pot experiments)**

No.	Treatment	Concentration mg L <sup>-1</sup>	Plant height cm	Fresh weight gm	Dry weight gm
1	Control (Plant)	0	27.83	8.4	2.67
2	Control ( <i>F. solani</i> )	0	25.73	5.13	1.67
3	Se + F. s	200	29.2	10.2	3.50
4	MnO <sub>2</sub> + F. s	200	29.86	10.64	3.33
5	MnO <sub>2</sub> + Se + F. s	100+100	31.13	11.26	4.1
6	Se	200	30.53	11.2	3.77
7	MnO <sub>2</sub>	200	31.13	11.37	3.93
8	MnO <sub>2</sub> + Se	100+100	32.61	12.32	4.6
9	Uniform + F. s	2.5	30.21	11.03	4.33
	L.S.D <sub>0.05</sub>		0.1666	0.4168	1.424

Each number in the table represents the mean of 3 replicates

The results for infection severity and percentage showed that all treatments had significant differences compared to the fungal pathogen control (*F. solani* only). Treatments without the fungal pathogen performed better due to the absence of the disease-causing agent. Among the treatments with the pathogen, the combined treatment of manganese oxide and selenium with the fungal pathogen (MnO<sub>2</sub> + Se + *F. solani*) achieved the best results, with an average infection severity

of 16.7% and an infection percentage of 29.97%. Other treatments followed in performance, as shown in Table (6). The increase in growth parameters in plants can be attributed to the added materials stimulating plant defenses to combat the pathogen, in addition to their inhibitory effect on the pathogenic fungus. These materials also enhanced photosynthesis and improved plant growth parameters (Rashed *et al.*, 2019; El-Ramady *et al.*, 2022; Shahbaz *et al.*, 2023).

**Table 6. Effect of Selenium, Manganese Oxide, and their combination on infection severity and percentage in pepper plants under greenhouse conditions (pot experiments)**

No.	Treatment	Concentration mg L <sup>-1</sup>	Infection severity	Infection percent %
1	Control (Plant)	0	0	0
2	Control ( <i>F. solani</i> )	0	100	100
3	Se + F. s	200	35	56.64
4	MnO <sub>2</sub> + F. s	200	36.7	74
5	MnO <sub>2</sub> + Se + F. s	200	16.7	29.97
6	Se	200	0	0
7	MnO <sub>2</sub>	200	0	0
8	MnO <sub>2</sub> + Se	200	0	0
9	Uniform + F. s	2.5	0	0
	L.S.D <sub>0.05</sub>		11.20	5.067

Each number in the table represents the mean of 3 replicates

#### 4-1 Evaluation of the efficiency of the best treatments in field pot experiments:

**4-1-1. Effect of induction factors on Peroxidase enzyme activity in pepper plants:** The results of peroxidase enzyme activity measurements showed a significant increase in enzyme activity during the second week after treatment with all materials. The three-week average results indicated that the combined treatments with the fungal pathogen

and the Uniform fungicide with the pathogen significantly outperformed the control treatments (plant only and *F. solani* only). The combined treatment with the fungal pathogen demonstrated the highest enzyme activity, with a three-week average of 78.4 U/ml, followed by the Uniform fungicide with the pathogen treatment, which had an average of 72.9 U/ml over three weeks. Other treatments did not show significant differences compared to the

fungal pathogen control, whereas all treatments exhibited significant differences compared to the plant-only control without the fungal pathogen, as shown in Table (7). The

enzyme value decreases after a while because the resistance induction does not last for a long time and requires adding materials to the soil again.

**Table 7. Effect of selenium, manganese oxide, and their combination on peroxidase enzyme activity in pepper plants in the field**

No.	Treatment	Peroxidase Enzyme Activity (U/ml) after 7 Days	Peroxidase Enzyme Activity (U/ml) after 14 Days	Peroxidase Enzyme Activity (U/ml) after 21 Days	Mean
1	Control (Plant)	32.44	33.79	34.33	33.5
2	Control ( <i>F. solani</i> )	55.50	60.07	51.35	55.6
3	Se + <i>F. solani</i>	58.55	64.12	52.18	58.3
4	MnO <sub>2</sub> + <i>F. solani</i>	44.23	73.47	64.23	60.6
5	MnO <sub>2</sub> + Se + <i>F. solani</i>	75.87	88.36	70.92	78.4
6	Uniform + <i>F. solani</i>	71.12	79.28	68.27	72.9
	L.S.D <sub>0.05</sub>	0.686	6.541	1.441	11.35

Each number in the table represents the mean of 3 replicates

**4-1-2. Effect of induction factors on Phenylalanine Ammonia-Lyase (PAL) enzyme activity in pepper plants:** The results of PAL enzyme activity measurements indicated a significant increase in enzyme activity during the second week after treatment with all materials. The three-week average results showed that the treatments with selenium and the fungal pathogen, as well as the combination of selenium and manganese oxide with the fungal pathogen, had

significantly higher enzyme activity compared to the fungal pathogen control. The three-week average enzyme activity for these treatments was 81.41 and 80.76 (U/ml), respectively. Other treatments did not show significant differences compared to the fungal pathogen control, whereas all treatments showed significant differences compared to the plant-only control without the fungal pathogen, as presented in Table (8).

**Table 8. Effect of Selenium, Manganese Oxide, and Their Combination on PAL Enzyme Activity in Pepper Plants in the Field**

No.	Treatment	PAL Enzyme Activity (U/ml) after 7 Days	PAL Enzyme Activity (U/ml) after 14 Days	PAL Enzyme Activity (U/ml) after 21 Days	Mean
1	Control (Plant)	41.31	44.72	42.48	42.84
2	Control ( <i>F. solani</i> )	67.64	76.56	53.37	65.86
3	Se + F. s	82.35	91.55	70.32	81.41
4	MnO <sub>2</sub> + F. s	74.46	85.65	62.34	74.15
5	MnO <sub>2</sub> + Se + F. s	84.36	93.46	64.47	80.76
6	Uniform + F. s	66.56	74.55	57.5	66.20
	L.S.D <sub>0.05</sub>	0.3411	1.363	0.4187	8.81

Each number in the table represents the mean of 3 replicates

**4-1-3. Effect of induction factors on Catalase enzyme activity in pepper plants:** The results of catalase enzyme activity measurements showed an increase in activity during the second week, similar to the previous sections, after treatment with all materials. The three-week average results indicated that the chemical fungicide (Uniform) with the fungal pathogen and the combined treatment of selenium and manganese oxide with the fungal pathogen

exhibited significantly higher enzyme activity compared to the untreated control and the fungal pathogen control. The average enzyme activity for these treatments over three weeks was 33.86 and 23.23 U/ml, respectively. All treatments showed significant differences compared to the plant-only control without the fungal pathogen, as shown in Table (9). The increase in enzyme activity can be attributed to the fact that these enzymes are naturally present in plants but are stimulated by external

factors such as infection by a pathogen, as seen in the fungal pathogen control (Hussein *et al.*, 2019; Khan *et al.*, 2021, Shahbaz *et al.*, 2023). Additionally, the induction materials, including selenium, manganese oxide, and

their combination, further stimulated these enzymes. These materials enhanced the plant's defense mechanisms and increased the levels of defensive enzymes such as peroxidase, PAL, and catalase (Schmidt & Husted, 2019).

**Table 9. Effect of Selenium, Manganese Oxide, and Their Combination on Catalase Enzyme Activity in Pepper Plants in the Field**

No.	Treatment	Catalase Enzyme Activity (U/ml) after 7 Days	Catalase Enzyme Activity (U/ml) after 14 Days	Catalase Enzyme Activity (U/ml) after 21 Days	Mean
1	Control (Plant)	7.46	8.18	8.38	8.01
2	Control ( <i>F. solani</i> )	13.17	17.2	11.56	13.98
3	Se + <i>F. s</i>	16.27	25.39	11.8	17.82
4	MnO <sub>2</sub> + <i>F. s</i>	14.18	19.21	10.32	14.57
5	MnO <sub>2</sub> + Se + <i>F. s</i>	22.49	27.24	19.96	23.23
6	Uniform + <i>F. s</i>	33.15	37.39	31.04	33.86
	L.S.D <sub>0.05</sub>	0.4941	0.7075	0.6399	4.141

Each number in the table represents the mean of 3 replicates

**4-1-4 Calculation of Infection Percentage and Severity:** The results of the added treatments on the soil to combat the fungal pathogen *F. solani* showed that the combined treatment of materials with the fungal pathogen and the Uniform fungicide with the fungal pathogen were the most effective in reducing infection severity and percentage. Both treatments achieved infection severity and percentage of (0, 0%), respectively. These were followed by the combined treatment of conventional materials with the fungal pathogen, which resulted in an infection severity and percentage of (10, 36.7%), respectively. Other treatments followed in

performance, as shown in Table (10). The reduction in infection severity in the treatments can be attributed to the effects of the added materials and their combinations, as well as the chemical fungicide, on the fungal pathogen and the plant. These factors stimulated the plant to produce enzymes, proteins, and induced resistance, ultimately leading to the death of the pathogen in the Uniform fungicide treatments. Moreover, the other treatments significantly reduced the growth of the pathogen to varying degrees (Eskandari *et al.*, 2020; Troni *et al.*, 2021, Wulandari *et al.*, 2022).

**Table 10. Effect of selenium, manganese oxide, and their combination on infection severity and percentage in pepper plants in the field.**

No.	Treatment	infection	infection
1	Control (Plant)	0	0
2	Control ( <i>F. solani</i> )	95	100
3	Se + <i>F. solani</i>	16.67	40
4	MnO <sub>2</sub> + <i>F. solani</i>	23.33	46.7
5	MnO <sub>2</sub> + Se + <i>F. solani</i>	10	36.7
6	Uniform + <i>F. solani</i>	0	0
	L.S.D <sub>0.05</sub>	8.79	9.20

Each number in the table represents the mean of 3 replicates

#### CONFLICT OF INTEREST

The authors declare that they have no conflicts of interest.

#### AUTHOR/S DECLARATION

We confirm that all Figures and Tables in the manuscript are original to us. Additionally, any Figures and images that do not belong to

us have been incorporated with the required permissions for re-publication, which are included with the manuscript.

Author/s signature on Ethical Approval Statement.

Ethical Clearance and Animal welfare Funds:

## REFERENCES

- Al-aamel, A. N. A., & Al-maliky, B. S. A. (2023). Control pepper fusarium wilting by biocontrol agent *Trichoderma harzianum* and chelated iron Fe-EDDHA. *Baghdad Science Journal*, 20(6), 24.  
doi:  
<https://dx.doi.org/10.21123/bsj.2023.8037>.
- Al-Jarrah, N. S. A. (2011). The Effect of the Biological Mixture EMI and Magnetic Field in Protecting Cucumber Plants from Decay and Seedling Damping-Off Pathogens. Ph. D. Dissertation, College of Agricultural Engineering Sciences, University of Baghdad, pp: 138.
- Al-Rubaie A. R. (2022). Induction of Resistance Against Damping-Off and Root Rot Diseases in Pepper Plants Using Some Biological and Chemical Agents. M.Sc. Thesis. College of Agricultural Engineering Sciences, University of Baghdad. pp: 119.
- Al-Sufi, M. A. A. (2001). Isolation, Purification, and Characterization of Peroxidase Enzyme from the Latex of *Calotropis Procera* and Its Potential Applications. M.Sc. Thesis, Department of Food Industries, College of Agriculture, University of Baghdad, pp: 96.
- Avery, S. V., Singleton, I., Magan, N., & Goldman, G. H. (2019). The fungal threat to global food security. *Fungal biology*, 123(8), 555-557.  
<https://doi.org/10.1016/j.funbio.2019.03.006>
- Boubakri, H., Chong, J., Poutaraud, A., Schmitt, C., Bertsch, C., Mliki, A., Masson, J.E., & Soustre-Gacougnolle, I. (2013). Riboflavin (Vitamin B2) induces defence responses and resistance to *Plasmopara viticola* in grapevine. *European Journal of Plant Pathology*, 136(4), 837-855.  
<https://doi.org/10.1007/s10658-013-0211-x>
- El-Ramady, H., El-Sakhawy, T., Omara, A. E. D., Prokisch, J., & Brevik, E. C. (2022). Selenium and nano-selenium for plant nutrition and crop quality. In *Selenium and nano-selenium in environmental stress management and crop quality improvement* (pp. 55-78). Cham: Springer International Publishing.  
[https://doi.org/10.1007/978-3-031-07063-1\\_4](https://doi.org/10.1007/978-3-031-07063-1_4)
- Eskandari, S., Höfte, H., & Zhang, T. (2020). Foliar manganese spray induces the resistance of cucumber to *Colletotrichum lagenarium*. *Journal of plant physiology*, 246, 153129.  
<https://doi.org/10.1016/j.jplph.2020.153129>
- Hamid, F.R. (2002). Study of The Efficiency of *Trichoderma* spp. Isolates in Inducing Resistance Against *Rhizoctonia Solani* and Stimulating Growth in Four Cotton Cultivars. M.Sc. Thesis, Department of Plant Protection, College of Agriculture, University of Baghdad, pp: 80.
- Hatem, M. W., & Ali, H. A. (2023, December). Methods of Inducing Systemic Resistance in plants by Using Abiotic Agents to Control Powdery Mildew Pathogens. In *IOP Conference Series: Earth and Environmental Science* (Vol. 1252, No. 1, p. 012004). IOP Publishing.  
<https://doi.org/10.1088/1755-1315/1252/1/012004>
- Hussein, H. A. A., Darwesh, O. M., & Mekki, B. B. (2019). Environmentally friendly nano-selenium to improve antioxidant system and growth of groundnut cultivars under sandy soil conditions. *Biocatalysis and agricultural biotechnology*, 18, 101080.  
<https://doi.org/10.1016/j.bcab.2019.101080>
- Bano, I., Kašparová, E. Š., Sajjad, H., & Reza, R. (2021). Importance of micro-nutrient supplementation for livestock a mini-review. *Acta Scientific Veterinary Sciences* (ISSN: 2582-3183), 3(7).  
<https://www.researchgate.net/publication/352903340>
- Ismail, F., Anjum, M. R., Mamon, A. N., & Kazi, T. G. (2011). Trace metal contents of vegetables and fruits of Hyderabad retail market. *Pakistan journal of nutrition*, 10(4), 365-372.  
<https://www.researchgate.net/profile/Fatima-Ismail-4/publication/230788723>
- Khan, M., Salman, M., Jan, S. A., & Shinwari, Z. K. (2021). Biological control of fungal phytopathogens: A comprehensive review based on *Bacillus* species. *MOJ Biol. Med*, 6, 90-92.  
<https://www.academia.edu/download/108188466/MOJBM-06-00137.pdf>

- Kim, S. W., Jung, J. H., Lamsal, K., Kim, Y. S., Min, J. S., & Lee, Y. S. (2012). Antifungal effects of silver nanoparticles (AgNPs) against various plant pathogenic fungi. *Mycobiology*, 40(1), 53-58.  
<https://doi.org/10.5941/MYCO.2012.40.1.053>
- Mao, X., Hua, C., Yang, L., Zhang, Y., Sun, Z., Li, L., & Li, T. (2020). The effects of selenium on wheat fusarium head blight and DON accumulation were selenium compound-dependent. *Toxins*, 12(9), 573.  
<https://www.mdpi.com/2072-6651/12/9/573>
- Mousavi, S. R., Galavi, M., & Ahmadvand, G. (2007). Effect of zinc and manganese foliar application on yield, quality and enrichment on potato (*Solanum tuberosum* L.).  
doi:  
<https://doi.org/10.3923/ajps.2007.1256.1260>
- Nawrocka, J., Małolepsza, U., Szymczak, K., & Szczech, M. (2018). Involvement of metabolic components, volatile compounds, PR proteins, and mechanical strengthening in multilayer protection of cucumber plants against *Rhizoctonia solani* activated by *Trichoderma atroviride* TRS25. *Protoplasma*, 255(1), 359-373.  
<https://doi.org/10.1007/s00709-017-1157-1>
- Özer, G., Türkkan, M., Sönmez, F., Kabakcı, H., Alkan, M., & Derviş, S. (2024). In vitro evaluation of selenium against some plant pathogenic fungi. *Akademik Ziraat Dergisi*, 13(1), 99-110.  
<https://dergipark.org.tr/en/pub/azd/issue/86009/1452105>
- Rashed, M. H., Hoque, T. S., Jahangir, M. M. R., & Hashem, M. A. (2019). Manganese as a micronutrient in agriculture: crop requirement and management. *Journal of Environmental Science and Natural Resources*, 12(1-2), 225-242.  
<https://www.researchgate.net/profile/Mmr-Jahangir/publication/349342170>.
- Schiavon, M., Lima, L. W., Jiang, Y., & Hawkesford, M. J. (2017). Effects of selenium on plant metabolism and implications for crops and consumers. In *Selenium in plants: Molecular, physiological, ecological and evolutionary aspects* (pp. 257-275). Cham: Springer International Publishing.  
[https://doi.org/10.1007/978-3-319-56249-0\\_15](https://doi.org/10.1007/978-3-319-56249-0_15)
- Schmidt, S. B., & Husted, S. (2019). The biochemical properties of manganese in plants. *Plants*, 8(10), 381.  
<https://www.mdpi.com/2223-7747/8/10/381>
- Shahbaz, M., Akram, A., Mehak, A., Haq, E. U., Fatima, N., Wareen, G., & Sabullah, M. K. (2023). Evaluation of selenium nanoparticles in inducing disease resistance against spot blotch disease and promoting growth in wheat under biotic stress. *Plants*, 12(4), 761.  
<https://www.mdpi.com/2223-7747/12/4/761>
- Sun, R., Zhang, X. X., Guo, X., Wang, D., & Chu, H. (2015). Bacterial diversity in soils subjected to long-term chemical fertilization can be more stably maintained with the addition of livestock manure than wheat straw. *Soil Biology and Biochemistry*, 88, 9-18.  
<https://doi.org/10.1016/j.soilbio.2015.05.007>
- Troni, E., Beccari, G., D'Amato, R., Tini, F., Baldo, D., Senatore, M. T., ... & Covarelli, L. (2021). In vitro evaluation of the inhibitory activity of different selenium chemical forms on the growth of a *Fusarium proliferatum* strain isolated from rice seedlings. *Plants*, 10(8), 1725.  
<https://www.mdpi.com/2223-7747/10/8/1725>
- Ullah, A., Farooq, M., Rehman, A., Arshad, M. S., Shoukat, H., Nadeem, A., & Nadeem, F. (2018). Manganese nutrition improves the productivity and grain biofortification of bread wheat in alkaline calcareous soil. *Experimental Agriculture*, 54(5), 744-754.  
<https://doi.org/10.1017/S0014479717000369>
- Wu, Q., Chen, Y., Zou, W., Pan, Y. B., Lin, P., Xu, L., & Que, Y. (2023). Genome-wide characterization of sugarcane catalase gene family identifies a ScCAT1 gene associated disease resistance. *International Journal of Biological Macromolecules*, 232, 123398.  
<https://doi.org/10.1016/j.ijbiomac.2023.123398>
- Wulandari, E., Prasetyo, J., Nurdin, M., & Maryono, T. (2022). Mildew and the growth of corn plant. *Jurnal Agrotek Tropika*, 10(1), 43-49.  
<https://www.academia.edu/download/107086821/3904.pdf>

تأثير عنصر السيلينيوم وأوكسيد المنغنيز والخلط بينهما في استحثاث المقاومة في نبات الفلفل المصاب بالفطر الممرض  
*Fusarium spp.*

براء عباس احمد الجبوري ، مينا وليد حاتم  
قسم وقاية النبات، كلية علوم الهندسة الزراعية، جامعة بغداد، العراق.

المستخلص

نفذت التجربة في احد البيوت البلاستيكية التابعة لقسم وقاية النبات/ كلية علوم الهندسة الزراعية/ جامعة بغداد- الجادرية في الموسم الزراعي 2023-2024 وذلك لبيان تأثير عنصر السيلينيوم واوكسيد المنغنيز في استحثاث مقاومة نبات الفلفل ضد الفطر الممرض *Fusarium solani* فضلاً عن زيادة بعض معايير النمو في النبات، إذ بينت نتائج التجربة ان إضافة هذه العناصر بخلطها معاً بالتركيز الأفضل 200 ملغم/ لتر أدى الى تثبيط نمو الفطر في المختبر بنسبة 77.71% مقارنة مع معاملة الفطر الممرض فقط والتي بلغت نسبة التثبيط فيها 0%، كما أدت هذه المعاملة الى زيادة بعض معايير النمو (اطوال النبات، الوزن الطري، الوزن الجاف) في تجربة الاصص، كما أدت الى خفض شدة ونسبة الإصابة في تجارب الاصص والتجربة الحقلية، فضلاً عن زيادة الانزيمات (البيروكسيداز، PAL، الكاتليز) في نبات الفلفل في الحقل.

الكلمات المفتاحية: *Capsicum annum* ، *F. solani* ،  $MnO_2$  ، Se.