HISTOLOGICAL EFFECTS OF AQUEOUS EXTRACT OF Mentha spicata ON LIVER IN ALBINO MICE L. H. Ali Lecturer Anbar University\ College of Education for Pure Sciences. Hatemloay179@yahoo.com

ABSTRACT

This study was conducted to investigate the effects of high concentration of *Mentha spicata* on liver tissues. Fourteen mature male albino mice were used in this study and divided into two groups. Control group received 1 ml normal saline daily for 7days through oro- gastric tube. The remained animals received 1ml of aqueous extract of *Mentha spicata* daily for 7 days through oro-gastric tube. At the end of experiment, all animals were dissected. Liver Samples were fixed in 10% buffered formalin and embedded in paraffin. Sections were prepared and stained with hematoxylin and eosin. The slides were examined macroscopically. The results showed that coagulative degeneration, degeneration of hepatocytes with necrosis, inflammation vacuolization, inflammatory cell infiltration and fatty degenerative changes as also hemorrhage and increased size of Kupffer cells and vacuolated cytoplasm. From this study it has been concluded that high concentration of aqueous extract of *Mentha spicata* have a side effect on the structure and function of liver tissue.

Key word: hepatocytes, degeneration, infiltration, nercrosis, kupffer cells

مجلة العلوم الزراعية العراقية – 48: (عدد خاص): 86- 91 /2017 على المختبرية التأثيرات النسيجية من المستخلص المائي لنبات النعناع على انسجة كبد الفئران المختبرية لؤي حاتم علي مدرس كلية التربية للعلوم الصرفه – جامعة الانبار Hatemloay179@yahoo.com

المستخلص

Ali

طبقت هذه الدراسة لمعرفة تاثير التراكيز العالية من نبات النعناع Mentha spicata على انسجة الخلايا الكبدية. استخدمت في هذه الدراسة 14 ذكر من الفئران المختبرية وقسمت الى مجموعتين. مجموعة السيطرة جرعت ب 1 مل من المحلول الملحي خلال 7 يوم بواسطة انبوية التجريع الفموي. المجموعة الثانية جرعت ب 1 مل من المستخلص المائي من نبات النعناع ولمدة 7 يوم بواسطة انبوية التجريع الفموي. في نهاية التجرية تم قتل الحيوانات وتم تشريحها وعملت مقاطع نسيجية للكبد وفحصت تحت المجهر الضوئي. اوضحت النتائج ظهور عمليات تنكس للخلايا الكبدية مع حدوث تنخر للخلايا والتهاب وعائي وارتشاح في الخلايا الالتهابية مع حدوث تنكس دهني وايضا نزف مع زيادة في اعداد خلايا كوبر وتفجي السايتوبلازم. نستنتج من هذه الدراسة ان التراكيز العالية من المستخلص المائي يانتيج والوظيفي لنسيج الكبد.

كلمات مفتاحية: الخلايا الكبدية، التنكس، الارتشاح، التنخر، خلايا كوبر

86

INTRODUCTION

Medicinal plants have always played an important role in the treatment of human diseases all over the world (16, 31). About 80% of the world's population relies on plantderived medicines for their healthcare (21). Plants contain a variety of organic compounds as secondary metabolites, and a large number of them display pharmacological properties. However, most medicinal plants are used as self medication without knowing their possible side effects. Therefore, traditional plants need further scientific investigation on their toxic side effects (22). It was reported that Iraq has many diverse herbal plants and traditional medicine is widely used for treatment of many diseases in this country (5). Mentha (Family, Lamiaceae) is a genus of 6 species in the flora of Iraq (7, 8). These species have been used in folk medicine and are available in traditional medicinal plant stores and local markets (9). commonly Mentha spicata, known as spearmint, is one of these species. It is a perennial herb with a characteristic spearmint odor with a good flavor and fragrance (6, 18). The plant traditionally used worldwide in pharmaceutical preparations, confectionery and food industries, and also in hygiene and cosmetic products (11, 24). The leaves are well recognized for making herbal tea (1, 14). Some studies have been shown that spearmint oil has anti-fungal, anti-microbial, antiinflammatory, anti-tumor and antioxidant activity (19, 27, 32, 33). These activities are due to the presence of active constituents like menthone, menthol, rosmaninric acid and carvone (4, 15). However, there is some evidence which show that despite its beneficial effects, spearmint has some toxic and adverse effects. Severe histopathological changes in kidney, liver and uterus tissue (3, 20). The data of the toxicity studies on medicinal plants or preparations derived from them should be obtained in order to increase the confidence in their safety to humans, particularly for use in the development of pharmaceuticals. This study was therefore undertaken to evaluate the possible toxicological effect of the Mentha spicata extract on liver tissue of mice.

MATERIAL AND METHODS

Preparation of aqueous extract of peppermint: Leaves of *Mentha spicata* were cleaned, and standardized in "Iraqi National Herbarium". Peppermint leaves were washed and left for air-dry and ground into powder using an electric grinder. 10 gm of powder was macerated in 100ml boiling distilled water, mixed by a glass rod, and was left in steam bath for 60 minutes. The aqueous extract was filtrated by using filter paper (28).

Laboratory animals

Fourteen Swiss albino mice of either sex (10-12 weeks old) weighing 25-30 grams were used in this study. Mice were kept in the University's animal house under standard environmental conditions (temperature: $22 \pm 2^{\circ}$ C and 12/12 h (light/dark) period) and fed with standard pellets and water *ad libitum* throughout the experiments. Mice were divided randomly into two groups:

A- Control group: animals received 1ml distilled water daily for 7 days through oro-gastric tube.

B- Experimental mice: animals administered with 1ml of aqueous extract of *Mentha spicata* daily for 7 days through oro- gastric tube (23). The animals were observed daily for abnormal clinical signs and death. Body weights were measured and recorded at the beginning and the end of the experiment. At the end of the study, all animals were sacrificed by spinal dislocation and dissected.

Histological Examination

For histological examination, the liver samples were removed and fixed in 10% of buffered formalin solution for 24 hour. The tissues were processed (dehydration, cleaning and infiltration). Then, they were embedded in paraffin wax and sectioned with microtome to produce 5 μ m paraffin wax tissue sections. After that, the sections were stained with Haematoxylin & Eosin followed by mounting with DPX mounting media. Next, the sections were examined using light microscope (12).

Statistical Analyses

Statistical analysis of data was performed using the Statistical Package for Social Sciences (SPSS for Windows, V. 13.0, Chicago, USA). Statistical significance was determined by one way analysis of variance (ANOVA) followed by Duncan's multiple range test (DMRT). Results were presented as mean \pm SD. *p* values < 0.05 were regarded as statistically significant (17).

RESULT AND DISCUSSION

1- Body weight: The weights of mice were not significantly different between control and treated group following administration of the extract

Table 1.Indicate the body weig	ht	ıt	igł	wei	odv	bo	the	idicate	1.Ind	1	Table]
--------------------------------	----	----	-----	-----	-----	----	-----	---------	-------	---	-------	---

	Mean of body weig	ght (gm)±SE
Groups	Before treatment	After7-days
		treatment
Control	14.3±0.65	17.60 ± 0.55
Treated	14.5±0.70	18.2±0.57

Histological Examination of Liver Tissue Histological examination of the liver tissues showed varied damage when compared with the baseline-control group (Figure 1).

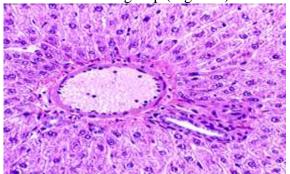


Figure 1. Histological section of liver tissue in normal group show central vein, column of hepatocytes radiating from the central vein, liver sinusoids. (H a& E stain) (200X).

The effects of mentha spicta are shown coagulative degeneration, degeneration of hepatocytes with necrosis, , inflammatory cell infiltration and fatty degenerative changes as also hemorrhage and increased size of Kupffer cells and vacuolated cytoplasm. Mice that were treated with Mentha spicata (group 2) experienced the greatest damage which was mainly irreversible and included nuclear changes (pyknosis, karyolysis and), cytolysis and massive congestion, Inflammatory cellular infiltration was abundant around the central vein in group2. In recent years, an increasing percentage of people from developed countries complementary have been using and alternative medicines .Peppermint (Mentha spicata) is widely used in food, cosmetics and medicines and pharmaceutical industries (10). In this study the administration of aqueous extract of Mentha spicata affects the structure of hepatocytes in liver. Liver is the gateway of body as well as the largest metabolic organ, containing an enzymatic system. This allows the metabolism of protein, fat and carbohydrate and maximum excretion of all foreign substances from the body (11).

Cytotoxicity of some medicinal plants has been well known (2). According to the results of this study, the aqueous extract of M. spicata leaves has cytotoxic effects on liver tissues. These changes might due to high concentration of menthol which is the principle components of the Mentha extracts (30). The cytotoxic activity of Mentha essential oils could be due to the major compositions including menthol, limonene, carvone. Mentha arvensis had major component of menthol (92.38%), and Mentha spicata of limonene (44.12%) and carvone (41.31%)(29,36) Once at all. this concentration of Mentha extract caused several changes in the plasma membrane and these changes could be explained due to the effect of essential oils which act by means of their lipophilic fraction reacting with the lipid parts of the cell membranes, and as a result, modify the activity of the calcium ion channels. The volatile oils saturated the membranes and can interact by means of their physiochemical properties and molecular shapes, and can influence their enzymes, carrier, ion channels and receptors (13). Several toxicological effects have been associated with the oil components of Mentha arvensis and Mentha spicata. In particular, menthol has been found to interact with cytosolic Ca₂, probably through an intracellular Ca₂ store release and Ca₂ channel blocking (30). Menthone has been reported to be a growth inhibitor, whereas abortifacient. pulegone, a potent is metabolized hepatic by microsomal monooxygenases to a series of hepatotoxins that cause liver cancer(30). Similarly, Spindler reported Madsen(34) that and all hematological and biochemical parameters as well as absolute and relative weights of the organs were within normal range, whereas the histopathological examination revealed alteration in the rat brain following the administration of peppermint oil (oil from Mentha piperita for a 86-day experimental period). Also the pathological changes may lead to impaired liver function which interferes with the secretion of plasma proteins, This leads to decreased blood osmotic pressure, with subsequent decreased drainage of tissue fluids, which explains the oedema and congestion observed in the different tissue (25, 26).

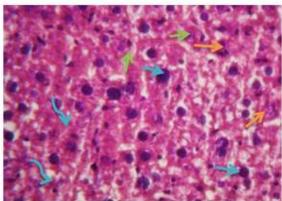


Figure (2) Histological section of liver tissue in second mice group show degeneration of some nuclei , fragmentation of nuclei , karyolysis , inflammatory Infiltration (H a& E stain) (200X).

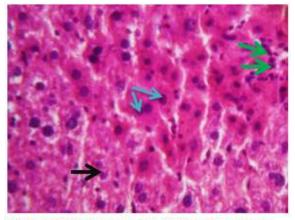


Figure (3) Histological section of liver tissue In mice second group show coagulative degeneration, degeneration of some nuclei, inflammatory infiltration (H a& E stain) (200X).

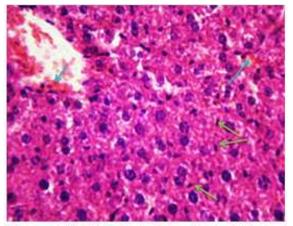


Figure (4) Histological section of liver tissue in mice second group show hemorrhage , increase of kuffper cell (H a& E stain) (200X).

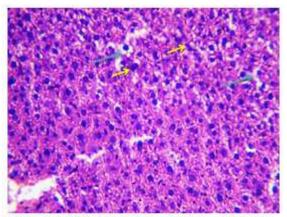


Figure (5) Histological section of liver tissue in mice second group show degeneration of some nuclei, pyknosis

\Rightarrow (H a& E stain) (200X).

REFRENCES

1. Adsersen A, Gauguin B, Gudiksen L and Jäger A. K. 2006. Screening of plants used in Danish folk medicine to treat memory dysfunction for cetylcholinesterase inhibitory activity.J Ethnopharmacol 104:418-422.

2. Afifi-Yazar FU, V. Kasabri, R. Abu-Dahab 2011. Medicinal Plants from Jordan in the Treatment of Cancer: Traditional Uses vs. In vitro and In Vivo valuations - Part 1. Planta Med.

3. Akdogan M, M. Ozguner, G. Aydin, and O. Gokalp 2004. Investigation of biochemical and histopathological effects of *Mentha piperita* Labiatae and Mentha spicata Labiatae on liver tissue in rats. Hum Exp Toxicol, 23: 21–28.

4. Akhilesh Kumar, Sharmila Chaatopadhyay. 2007. DNA damage protecting activity and antioxidant potential of pudina extracts. Food Chem. 100: 1377-1384.

5. Al-Douri N.A, and L. Al-Essa, 2010. A Survey of plants used in Iraqi traditional medicine. Jordan J.Pharmaceut. sci., 3(2):100-110.

6. Ali M.S, Ahmed W, Saleem Mand Khan T 2006. Longifoamide-A and B: Two new ceramides from Mentha longifolia (Lamiaceae). Nat Prod Res 20:953-960.

7. Al-Musawi, Ali, H. E. 1987 . Taxonomy of plant . Department of Biology, College of Science, University of Baghdad.

8. Al-Okbi S., H. Fadel, and D. Mohamed, 2015. Phytochemical constituents, antioxidant and anticancer activity of *Mentha citrata* and *Mentha longifolia*. RJPBCS 6(1) pp: 739.

9. Amin G. 2005. Popular medicinal plants of Iran. Tehran University of Medical Sciences.

10. Arumugam P., and A. Ramesh, 2011. Invivo antioxidant effects of ethyl acetate fraction of *Mentha spicata* L. on 4nitroquinoline-1-Oxide Injected Mice. servicesIranian. Journal of Pharmaceutical Research, 10 (4): 787-793.

11. Bajaj P., and B. Tabassum, 2013. Protective role of *Mentha piperita* against cadmium induced alteration in transaminases of albino rats. Ind J Biol Stud Res Vol. 2 (2): 129-133.

12. Bancroft, J. D. and A. A. Stevens, 1982. Theory and Practice of Histological Techniques. 1st.edn., Churchill, Livingstone, Edinburgh; Melbourne and New York, pp. 189-190, 326-370.

13. Buchbauer , G. and L. jirovetz, 1994. Aromatherapy use of fragrances and essential oils as medicaments. Flav. Frager. J. 9:217-222.

14. Carmona M. D, R. Llorach, C. Obon and D. Rivera 2005. "Zahraa", a Unani multicomponent herbal tea widely consumed in Syria: Components of drug mixtures and alleged medicinal properties. J Ethnopharmacol 102:344-350.

15. Daferera D. J, B. N. Ziogas, and M. G. Polissiou 2003. The effectiveness of plant essential oils on the growth of Botrytis cinerea, Fusarium sp and Clavibacter michiganensis. Crop. Prot.; 22: 39-44.

16. Daswani, G. P., S. Brijesh, and J. T. Birdi, 2006. Preclinical Testing of Medicinal Plants: Advantages and Approaches. Workshop Proceedings Approaches towards on Evaluation of Medicinal Plants Prior to Trial. Foundation Clinical for Medical Research at Yashwantrao Chavan Academy of Development Administration (YASHADA), Pune, 60-77.

17. Duncan, R. C.; R. G. Knap, and M. C. Miller, 1983. Introductory Biostatistics for the Health Sciences, A Wiley Medical Publication, John Wiley and Sons, London. pp: 161-179.

18. Fletcher R. S.; T. Slimmon and L. S. Kott 2010. Environmental factors affecting the accumulation of rosmarinic acid in spearmint (*Mentha spicata* L.) and peppermint (*Mentha* *piperita* L.). The Open Agriculture Journal. 4: 10-16.

19. Guimaraes R, J. C. Barreira, L. Barros, A. M Carvalho, and I. C. Ferreira, 2011. Effects of Oral Dosage Form and Storage Periodon the Antioxidant Properties of Four Species Used in Traditional Herbal Medicine. Phytother Res, 25: 484–492.

20. Guney M, Oral B, N. Karahanl, T. Mungana, and M. Akdogan, 2006. The effect of *Mentha spicata* Labiatae on uterine tissue in rats. Toxicol Ind Health, 22: 343–348.

21. Hajjaj G., A. Bounihi, M. Y. Tajani, Cherrah, and A. Zellou, 2013. Evaluation OF CNS Activities Of *Matricaria Chamomilla* L. Essential Oil in Experimental Animals From Morocco. International Journal of Pharmacy and Pharmaceutical Sciences, Vol 5, Issue 2.

22. Inamul, H. 2004. Safety of medicinal plants. Pakistan Journal of Medical Research, 43, 4.

23. Jormut, S. F. 2004. Study of Action of Crude Aqueous Extract of some Medical Plants in Uterine Contractions and Induction of Abortion in Albino Mice. M.Sc. Thesis Baghdad University.

24. Kumar V, M. R Kural, B. M. J. Pereira, and P. Roy 2008. Spearmint induced hypothalamic oxidative stress and testicular anti-androgenicity in male rats – altered levels of gene expression, enzymes and hormones. Food Chem Toxicol, 46: 3563–3570.

25. Lacroix I, M. Lapeyre-Mestre, H. Bagheri, A. Pathak, and J. L. Montastruc 2004. On steroidal anti-inflammatory drug-induced liver injury: a case control study in primary care. Fundam Clin Pharmacol; 18(2):201-206.

26. Lapeyre-Mestre M, A. M. de Castro, M. P. Bareille, J. G. Del Pozo, A. A. Requejo, L. M. Arias, J. L. Montastruc, and A. Carvajal 2006. Non-steroidal anti-inflammatory drug-related hepatic damage in France and Spain: analysis from national spontaneous reporting systems. Fundam Clin Pharmacol; 20(4):391-395.

27. Lixandru B. E, N. O. Drăcea, C. C. Dragomirescu, E. C. Drăgulescu, I. L. Coldea, L. Anton, E. Dobre, C. Rovinaru, and I. Codiță. 2010. Antimicrobial activity of plant essential oils against bacterial and fungal species involved in food poisoning and/or food decay. Roum Arch Microbiol Immunol, 69: 224–230.

28. Arumugam P, N. Gayatri Priya, M. Subathra and A. Ramesh 2008. Antiinflammatory activity of four solvent fractions of ethanol extract of *Mentha spicata* L. investigated on acute and chronic inflammation induced rats. Environ. Toxicol. Pharmacol. 26: 92-95.

29. Nozhat F., A. Alaee, K. Behzadi, and N. Chegini., 2014. Evaluation of possible toxic effects of spearmint *(Mentha spicata)* on the reproductive system, fertility and number of offspring in adult male rats. AJP, Vol. 4, No. 6.

30. Odeyemi O., M. Yakubu , P. Masika, and A. Afolayan, 2009. Toxicological evaluation of the essential oil from *Mentha longifolia* L. subsp. capensis leaves in rats. J Med. Food 12 (3), 669–674.

31. Ogbonnia, S., A. A. Adekunle, M. K. Bosa, and V. N. Enwuru, 2008. Evaluation of acute and sub-Acute toxicity of alstonia congensis engler (Apocynaceae) bark and xylopia aethiopica (Dunal) A. rich (Annonaceae) fruits mixtures used in the treatment of diabetes. African Journal of Biotechnology, 7, 701-705.

32. Pearson W, R.S. Fletcher, L. S. Kott 2012.

Oral rosmarinic acid-enhanced *Mentha spicata* modulates synovial fluid biomarkers of inflammation in horses challenged with intraarticular LPS. J Vet Pharmacol Ther, 35: 495–502.

33. Soković M. D, J. Vukojević, P. D. Marin, D. D. Brkić, V. Vajs, and L. J. van Griensven 2009. Chemical composition of essential oils of *Thymus* and *Mentha* species and their antifungal activities. Molecules, 14: 238–249.

34. Spindler P, C. Madsen, 1992. Subchronic toxicity study of peppermintoil in rats. Toxicol Lett;62:215–220.

35. Spirling L. I and I. R. Daniels 2001. Botanical perspectives on health peppermint: more than just an after-dinner mint. J R Soc Health, 121; 62–63.

36. Weecharangsan W., W. Sithithaworn, and P. Siripong, 2014. Cytotoxic activity Of essential oils of *Mentha* Sps. On human carcinoma cells . J. Health Res. vol.28 no.1.

37. Wojcikowski K., D. JohnsoNn, and G. Gobe, 2004. Medicinal herbal extracts – renal friend or foe? Part one:The toxicities of medicinal herbs. Nephrology; 9, 313–318.