

RELATIONSHIP OF THE LEPTIN GENE POLYMORPHISM ON THE PRODUCTION PERFORMANCE IN LOCAL AWASSI LAMBS

E. R. Abbas

S. F. Mohamed

Lecturer

Prof.

Coll. Agric. Eng. Sci, University of Baghdad

enas.r@coagri.uobaghdad.edu.iq

ABSTRACT

This study was conducted at the Animal Farm of the College of Agricultural Engineering Sciences, University of Baghdad during the period from 22/1/2022 to 2/4/2022. This study was conducted to investigate the effect of leptin gene polymorphism on the weight gain, digestibility and rumen fermentations of Awassi lambs. Forty Awassi lambs were selected at age 4-5 months, with an initial weight mean of 25.4 kg, and distributed randomly in individual pens. The results showed that there were three variations in SNP (G>A), GG, GA and AA it was 66.67, 30.56 and 2.7%, respectively, and the differences between them were highly significant ($P \leq 0.01$). No significant differences of leptin gene polymorphism were noticed on the percentage of volatile fatty acids (acetic, propionic and butyric) as well as on the percentage of ammonia nitrogen at all times (0,3 and 6 hours). There were highly significant differences ($P \leq 0.01$) in pH of the rumen fluid in GG variations at the times of 0 and 6 hours compared with GA it was 6.78 ± 0.03 and 6.76 ± 0.02 , respectively. The results also showed that there were no significant differences in the leptin gene polymorphism on digestibility and weight gain. It can be concluded that the study of genetic variations of leptin gene had no effect on the performance of Awassi lambs.

Keywords; weight gain , digestibility , rumen fermentations

عباس ومحمد

مجلة العلوم الزراعية العراقية- 2025 :56 (5):1630-1637

علاقة تعدد المظاهر الوراثية لجين اللبتين مع الأداء الانتاجي للحملان العواسية المحلية

سندس فاروق محمد

ايناس رشيد عباس

استاذ

مدرس

كلية علوم الهندسة الزراعية ، جامعة بغداد

المستخلص

اجريت هذه الدراسة في الحقل الحيواني التابع لكلية علوم الهندسة الزراعية / جامعة بغداد للفترة من 22/1/2022 الى 2/4/2022 . هدفت هذه الدراسة بيان تأثير التغيرات الوراثية لجين اللبتين على معدل الزيادة الوزنية ومعامل الهضم وتخمرات الكرش للحملان العواسي. استخدمت في هذه التجربة 40 حملا ذكرا من الأغنام العواسي بعمر 4-5 أشهر وبوزن ابتدائي 25.4 كغم، تم توزيعها عشوائياً في حظائر منفردة. اظهرت النتائج وجود ثلاث طرز وراثية للتغيرات (G>A) وهي GG و GA و AA وكانت 66.67 و 30.56 و 2.7% على التوالي وكانت الاختلافات بينهم عالية المعنوية ($P \leq 0.01$), حيث اظهرت النتائج عدم وجود فروق معنوية في تأثير التغيرات لجين اللبتين في نسبة الاحماض الدهنية الطيارة (الاسيتيك و البروبيونيك و البيوتيريك) وكذلك في نسبة نيتروجين الامونيا و لجميع الاوقات (0,3,6 ساعة). بينما لوحظ وجود فروق عالية المعنوية ($p \leq 0.01$) في الاس الهيدروجيني (pH) في سائل الكرش لصالح التغيرات GG في الاوقات 0 و 6 ساعة, اذ بلغت 6.78 و 6.76 على التوالي . كذلك بينت النتائج عدم وجود فروق معنوية للتغيرات الوراثي لجين leptin في معامل هضم العناصر الغذائية ومعدل الزيادة الوزنية . يمكن الاستنتاج بان دراسة التغيرات الوراثية لجين اللبتين ليس لها تأثير في اداء الحملان العواسي.

الكلمات المفتاحية: الزيادة الوزنية، معامل الهضم، تخمرات الكرش.



This work is licensed under a Creative Commons Attribution 4.0 International License. Copyright© 2025 [College of Agricultural Engineering Sciences](#) - [University of Baghdad](#)

Received: 25/3/2023, Accepted: 9/7/2023, Published : October 2025

INTRODUCTION

Sheep are one of the most economic important local animals. Sheep were initially raised by following the method of natural selection or by relying on their productivity during the period of their rearing in the field, but this approved system slowed down the productivity of sheep and the possibility of benefiting from its economic characteristics to improve the nutritional and productive efficiency of the animal through genetic improvement techniques, many genes and their genetic variants have been identified that have an association with phenotypic traits and have economic importance (12,2). A protein hormone called leptin is released by white adipose tissue. It has a significant impact on energy metabolism and may have an impact on food intake, body energy homeostasis, and nutrient partitioning amongst tissues(5). Leptin also regulates lactogenesis, which is another benefit (11,19). Growth is an important economic trait in sheep (1,26,31). Evidence indicates a significant diversity in the leptin gene in sheep, with clear associations with growth traits and carcass characteristics (2, 32, 23, 28). The single nucleotide polymorphism (SNP) of the leptin gene in intron 2 was significantly associated with birth weight and average daily weight gain in Saanen goats (36). These results demonstrate that LEPSau3AL can be used as an effective genetic marker in growth improvement programs in goats (6, 7, 27, 37). Selecting genes is an important and effective method for genetic improvement of flocks (3, 23) and among these genes is leptin gene, as it is associated with a number of productive traits that are related to nutrition especially in sheep, such as gender, puberty, reproduction and milk production. It also has a fundamental role in growth and feed conversion efficiency (1). Other studies have shown that the leptin gene has a regulatory function in body weight, the amount of feed intake and growth in sheep (4,13, 21), In a study, it was found that the leptin gene influences meat quality and fat content, which affects carcass characteristics in both cows and bulls (14, 17, 25, 30, 38, 42). The results of the study also showed that the birth weight, weaning weight, and weight gain rate were significantly ($P \leq 0.05$) related to the

maternal genotype, and the best performance was in lambs from mothers carrying two genetic combinations, AA and AB. Therefore, the leptin gene is a promising genetic indicator that can be adopted in genetic selection programs for local sheep (16, 33, 39). The objective of this study was done to determine the relationship of leptin gene polymorphism with weight gain, digestibility and rumen fermentations of local Awassi lambs.

MATERIALS AND METHODS

This study was conducted at the College of Agricultural Engineering Sciences, Animal Production Department, University of Baghdad for the period from 22/1/2022 to 2/4/2022. Forty Awassi ram lambs were used to detect the leptin gene polymorphism and its association with digestibility, daily gain, live weight gain and rumen fermentation in local Awassi lambs. Five milliliters of blood were collected via jugular venipuncture in a collection tube supplemented with (K2 EDTA) anticoagulant, and transmitted in a cool box to the Laboratory of Agricultural Molecular Biology at the College of Agricultural Engineering Sciences, University of Baghdad for cryopreservation under -4°C , then DNA extraction from blood was done by using a special kit of extraction of DNA. The genomic DNA's integrity was assessed using electrophoresis on agarose gels. The leptin gene polymerase chain reaction (PCR) technique was focused on the major polymorphism observed exon 3 of the leptin gene. The 627 bp segment Gene, while the volume of whole genome for leptin gene was 443534bp observed in sheep. Genomic DNA was amplified using specific primers
F:5'AGAAGTAAGGGTCCAGGAAG3
R:5'CTGAGAGGAGCAAGAGAGA3'

Annealing temperature was started at 58°C and ending at 48°C (Touchdown PCR) from Bayer company at 30 cycle for 30 sec. The primer sequence as follows (41). After the end of the PCR-program, the samples were sent to the Republic of North Korea (Nappo Company) for DNA Sequencing. The data were analyzed statistically using Statistical Analysis System (SAS) (35). To study the effect of leptin gene Chi-square test (χ^2) was used to compare the percentages of genotypes distribution for the leptin gene in the sheep sample. Allelic

frequency was calculated according to the Falconer and Mackay (18) equation for Leptin gene. As for the chemical analysis of (the concentrate, the roughage, and the waste for the digestion experiment), the dry matter, organic matter, crude protein, crude fiber, ether extract, and nitrogen-free extract were estimated in the postgraduate nutrition laboratory of the Animal Production Department in the College of Agricultural Engineering Sciences(8). Rumen fermentation and volatile fatty acid concentrations were measured after thawing the frozen rumen fluid, filtering it, and then conducting the necessary analyses in the laboratories of the Ministry of Science and Technology / Environment and

Water Department using a chromatography device (40). To study the effect of LEPTIN gene polymorphism on the reproduction performance (the first mathematical model) in the studied traits Significant differences between averages by Duncan (15) multiple range test with the application of least square means.

The statistical model was as follows:

$$Y_{ij} = \mu + T_i + e_{ij}$$

Where:

Y_{ijk} = The observed value,

μ = Overall mean

T_i = The effect of leptin gene polymorphism

e_{ij} = Error term

$$PA = \frac{2 * \text{No. of Homozygous} + 1 * \text{No. of Heterozygous}}{2 * \text{Total number of samples}}$$

First allele replication: PA Since: $P + q = 1$, then the frequency of the second allele is: Qb

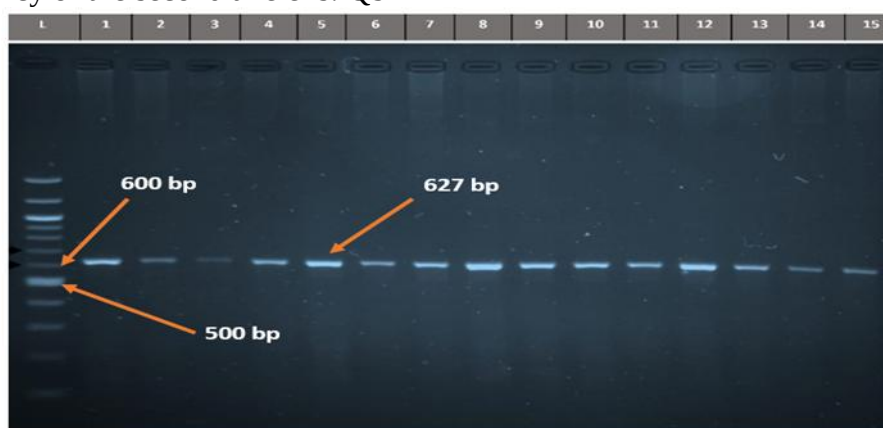


Figure 1.The electrophoresis of PCR for the Leptin gene ,agarose concentration 1.5%,5volt/cm for1.5hour,ladder(100-1000)bp,lanes1-1,5represent samples with sizes of approximately 627bp

RESULTS AND DISCUSSION

PCR amplification of exon 3 for leptin gene

PCR amplification of the target gene genomic DNA. The results showed the amplification of a fragment size 627 bp of the Leptin gene. The PCR products were separated successfully on 1.5% agarose gel with the DNA ladder (100 bp). The samples were amplified successfully by PCR reaction, and a single band stained with Ethidium Bromide was obtained, as shown in the Figure (1). The gene polymorphism, its percentages, and the allelic frequency of the Leptin gene are shown in Table (1). Three polymorphism appeared and

the percentages of GG,GA,AA were 66.67,30.56 and 2.77% respectively. Calculator of allele frequency of LEPTIN gene according of Hardy Weinberg's equilibrium(18).

The sequences of the nitrogenous bases of the leptin gene: The nitrogen base sequencing technique was used, and the results revealed that the studied segment of the leptin gene (627base pairs) had in the target coding region of the leptin gene, as three polymorphism appeared in this variation (G>A SNP) which are GG,GA and AA,(18) as shown in Figure (2).

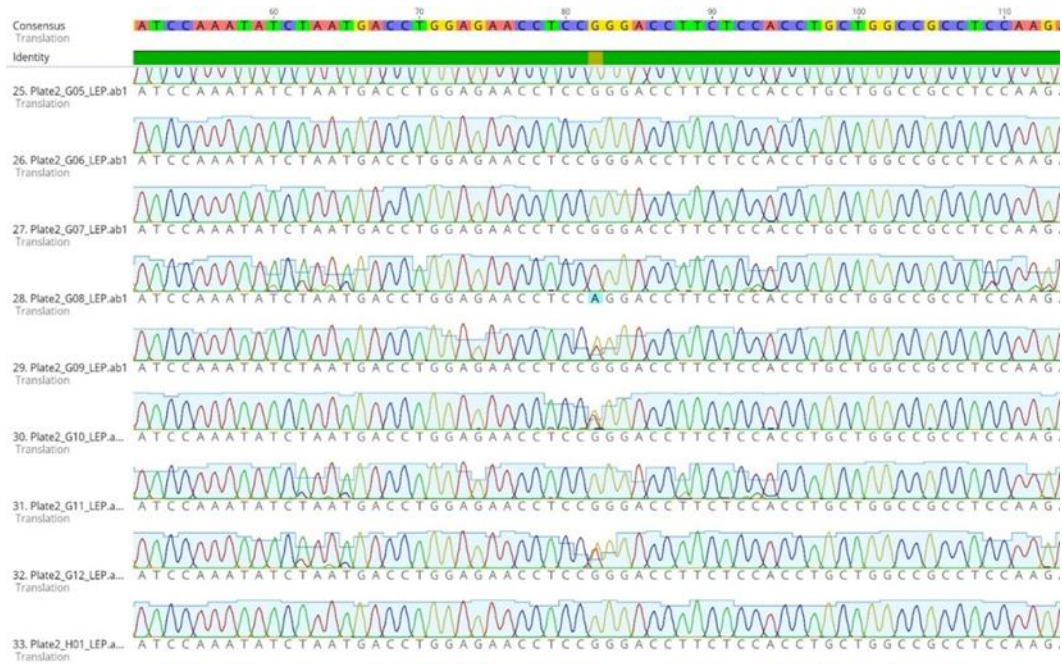


Figure 2. The sequences of the nitrogenous bases of the LEPTIN gene
Table 1. Distribution and allele frequency of the leptin gene G >A SNP polymorphism in a number of Awassi lambs

Leptin gene /G >A SNP Polymorphism	Number	Percentage (%)
GG	24	66.67
GA	11	30.56
AA	1	2.77
Total	36	%100
Chi-Square (χ^2)	p-Value	0.84
Allele		Frequency
G		0.82
A		0.18

($P \leq 0.01$)**

Table 2 shows the effect of the Leptin gene polymorphism on the rumen fermentations of lambs. The results show that there were no significant differences between the genotypes and the rumen fluid characteristics at different times (0, 3, 6 hours), but there was significant effect of the variants of the Leptin gene on rumen PH at 0 and 6 hours for the same genotype GA, and the ratios were 6.59 ± 0.03 at 0 hr. and 6.61 ± 0.03 at 6 hr. However, it is within the normal limits of the ruminal pH in sheep is between 6.4 and 6.8. Ruminal pH values less than 5.5 or greater than 7.0 are considered abnormal; ruminal pH of less than 5.5 is defined as subacute rumen acidosis (24). Awassi lambs were chosen to evaluate the effects of leptin gene polymorphisms on

rumen fermentations and ammonia nitrogen ($\text{NH}_3\text{-N}$). The results of the current investigation indicated that volatile fatty acids have no apparent effects. This might be a result of a similar diet with the same quantity of protein and energy for all animals; therefore, they may have prevented major variations in feed intake (29). This outcome was linked to the same study's no significant differences on weight gain and digestibility. The Leptin gene Polymorphism did not have any significant effects on dry matter intake (9,34). While polymorphisms of the leptin gene were associated with significant differences among genotypes in feed intake in cattle (10).

Table 2. Association of the leptin gene G>A SNP polymorphism with rumen fermentation

Leptin gene SNP (G>A) polymorphism	Means ± SE				
	PH	NH ₃ -N Mg/dl	Acetic Acid	Propionic acid	Butyric acid
0 hr.					
GG	6.78±0.03a	13.77±1.48a	11.94±0.10a	5.90±0.17a	3.95±0.19a
GA	6.59±0.03b	14.59±1.21a	11.65±0.14a	5.37±0.18a	3.41±0.28a
Level of significance	P≤0.01	NS	NS	NS	NS
3 hr.					
GG	6.63±0.05a	11.76±1.09a	16.35±0.42a	8.68±0.32a	6.90±0.18a
GA	6.45±0.03a	13.62±1.11a	15.09±0.82a	7.65±0.49a	6.37±0.03a
Level of significance	NS	NS	NS	NS	NS
6 hr.					
GG	6.76±0.02a	11.23±1.14a	15.26±0.37a	8.03±0.31a	6.35±0.21a
GA	6.61±0.03b	13.35±1.04a	13.97±0.5a	7.15±0.54a	6.09±0.22a
Level of significance	P≤0.01	NS	NS	NS	NS
Means with different letters in the same column differed significantly, NS: Non-Significant.					

Table 3 shows the effect of genotypes the Leptin gene on the digestibility of different nutrients. The results showed that there were no significant differences among the

genotypes and the traits studied. The results of this study disagreed with the findings of (21) who mentioned that the Leptin gene is one of the genes that is associated with growth traits.

Table 3. Association of the leptin gene /G>A SNP polymorphism with digestibility (%) and nutritive ratio

Leptin gene SNP (G>A) polymorphism	Means ± SE					
	Dry matter %	Organic matter %	Crude protein %	Crude fiber %	Ether extract %	Nitrogen free extract % Nutritive ratio %
GG	67.36±0.36	77.88±0.66	75.39±0.98	57.63±0.64	66.46±0.68	66.97±0.61
GA	66.11±1.41	78.30±0.61	74.77±1.41	55.78±0.65	65.52±1.15	65.54±1.44
Level of significance	NS	NS	NS	NS	NS	NS
Means with different letters in the same column differed significantly, NS: Non-Significant.						

Table (4) shows the effect of the genetic variation of the Leptin gene on the rate of weight gain, but there were no significant differences between the different genotypes and the trait studied. Leptin has a variety of purposes, including controlling appetite, body weight, and glucose homeostasis(13). In many sheep breeds, the LEP gene has been associated with growth (1) . In conclusion, the

results show a positive influence of GG SNP Polymorphism on pH of Rumen liquor and without effect on other studied traits. The results of the current study differed from previous studies due to the different conditions of the experiment and the animals used in the experiment, as well as the different environmental conditions of the experiment from previous stud

Table 4. Association of Leptin gene /G>A SNP Polymorphism with initial weight, final weight, total gain, total intake and feed conversion ratio

Leptin gene SNP (G>A) polymorphism	Means ± SE				
	Initial weight (kg)	Final weight (kg)	Total gain (kg)	Total intake (kg)	Feed conversion ratio %
GG	25.26±0.77	34.63±.76	9.36±0.63	71.83±1.67	8.11±0.42
GA	25.75±1.29	34.25±1.31	8.50±0.62	70.37±3.51	8.53±0.56
Level of significance	NS	NS	NS	NS	NS
NS: Non-Significant.					

Conclusion

The results showed the presence of one SNPS for each of the Leptin gene (G>82 A) and (G<292 A) in the blood sample taken from the Awassi male lambs used in the study. Based

on the results obtained, there is no significant effect between the genetic compositions of the Leptin gene and the studied traits, except for PH.

CONFLICT OF INTEREST

The authors declare that they have no conflicts of interest.

DECLARATION OF FUND

The authors declare that they have not received a fund.

REFERENCES

1. Abd Kadhim, J., A. Z. EK, and N. N. AL-Anbari, 2019. Relationship between leptin gene and some growth traits in local Awassi sheep. *J. Kerbala Agric. Sci.*, 4(4), 180-188. <http://doi.org/10.52113/mjas04/9.2/21>
2. Abdali, N. A. A. and A. H. Salim., 2023, Study the leptin gene polymorphism and its relationship with a number of carcass traits in Iraqi sheep. *IOP Conf. Ser.: Earth Environ. Sci.*, 1262, 072027. <http://doi.org/10.1088/17551315/1262/7/072027>
3. Ajafar, M. H., A. H. Kadhim and T. M. Al-Thuwaini., 2022. The reproductive traits of sheep and their influencing factors. *Rev. Agric. Sci.*, 10: 82-89. https://doi.org/10.7831/ras.10.0_82
4. Al-Burkat, H. A. H., H. M. J. Al-Khafaji and J. K. Al-Gharawi., 2023. Polymorphism of the leptin gene and its relationship to the body dimensions of the Awassi sheep. *Revis Bionatura* 2023; 8 (1), 73. <https://doi.org/10.21931/RB/2023.08.01.73>
5. Al-Hussaniy, H. A., A. H. Alburghaif and M. A. Naji., 2021. Leptin hormone and its effectiveness in reproduction, metabolism, immunity, diabetes, hopes and ambitions. *J. Med. Life*, 14(5), 600. <https://doi.org/10.25122/jml-2021-0153>
6. Alim, M. A. M. M. K. Hossain, J. Nusrat, M. Salimullah, Z. Shu-Hong and J. Alam., 2019. Genetic effects of leptin receptor (LEPR) polymorphism on litter size in a Black Bengal goat population. *Anim. Biol.*, 69(4), 411-420. <https://doi.org/10.1163/15707563-00001079>
7. Almaamory, Y. A. and N. N. Al-Anbari., 2025. Relationship of the FASN gene polymorphism with milk production and its components in local Awassi sheep. *Iraqi J. Agric. Sci.*, 56(3), 985-992. <https://doi.org/10.36103/vtjmb057>
8. AOAC, 2015. (Association of Official Analytical Chemists). Official methods of analysis. 18th ed. AOAC International, Gaithersburg, Maryland, USA.
9. Avondo, M., A. Di Trana, B. Valenti, A. Criscione, S. Bordonaro, A. De Angelis, D. Giorgio and P. Di Gregorio., 2019. Leptin gene polymorphism in goats fed with diet at different energy level: Effects on feed intake, milk traits, milk fatty acids composition, and metabolic state. *Animals*. 9 (7): 424. <https://doi.org/10.3390/ani9070424>
10. Banos, G., J. Woolliams, B. Woodward, A. Forbes and M. Coffey., 2008. Impact of single nucleotide polymorphisms in leptin, leptin receptor, growth hormone receptor, and diacylglycerol acyltransferase (DGAT1) gene loci on milk production, feed, and body energy traits of UK dairy cows. *J. Dairy Sci.* 91(8): 3190-3200. <https://doi.org/10.3168/jds.2007-0930>
11. Barbagallo, F., R. A. Condorelli, L. M. Mongioi, R. Cannarella, L. Cimino, M. C. Magagnini, A. Crafa, S. La Vignera and A. E. Calogero., 2021. Molecular mechanisms underlying the relationship between obesity and male infertility. *Metabolites*, 11(12), 840. <https://doi.org/10.3390/metabo11120840>
12. Bordoni, L. and R. Gabbianelli., 2019. Primers on nutrigenetics and nutri (epi) genomics: Origins and development of precision nutrition. *Biochimie*, 160, 156-171. <https://doi.org/10.1016/j.biochi.2019.03.006>
13. Caron, A., S. Lee, J. K. Elmquist and L. Autron .2018. Leptin and brain adipose crosstalks. *Nat. Rev. Neurosci.* 19(3): 153-165. <https://doi.org/10.1038/nrn.2018.7>
14. Darwish, A. M., M. A. Abdelhafez, Z. G. Abdel-Hamid, S. I. Othman, I.E. Mohamed and A. A. Allam., 2023. Correlation analysis between polymorphism of leptin and IGF1 genes and body measurements in Barki and Farafra sheep. *Beni-Suef University Journal of Basic and Applied Sciences*, 12(1), 119.
15. Duncan, D. B., 1955. Multiple range and multiple F tests. *Biometrics*. 11(1): 1-42. <https://doi.org/10.2307/3001478>
16. Enas, R. A. and S. F. Mohamed., 2023. Effect of genetic variation of leptin gene on intake of roughage and concentrate diet of Awassi Lambs. *IOP Conf. Ser.: Earth Environ. Sci.*, 1262, 072099. <https://doi.org/10.1088/1755-1315/1262/7/072099>

17. Ermawati, D., P. Panjono, S. Bintara, A. Agus, B. Widyobroto, B. Yudistyra, R. Mustofa and T. Hartatik., 2024. Association of leptin gene polymorphism with growth in crossbred cattle through PCR-RFLP analysis. Iraqi J. Vet. Sci., 38(4), 771-779. <https://doi.org/10.33899/ijvs.2024.148741.3610>
18. Falconer, D. S. and T. F. Mackay., 1996. Introduction to Quantitative Genetics (4th ed.). Longman, Harlow, UK.
19. Feuermann, Y., S. Mabjeesh and A. Shamay., 2004. Leptin affects prolactin action on milk protein and fat synthesis in the bovine mammary gland. J. Dairy Sci. 87(9): 2941-2946. [https://doi.org/10.3168/jds.S0022-0302\(04\)73425-6](https://doi.org/10.3168/jds.S0022-0302(04)73425-6)
20. Frühbeck, G., S. Jebb and A. Prentice., 1998. Leptin: physiology and pathophysiology. Clin. Physiol. 18(5): 399-419. <https://doi.org/10.1046/j.1365-2281.1998.00129.x>
21. Gebreselassie, G., H. Berihulay, L. Jiang and Y. Ma., 2019. Review on genomic regions and candidate genes associated with economically important production and reproduction traits in sheep (*Ovis aries*). Animals 10(1), 33. <https://doi.org/10.3390/ani10010033>
22. Girmay, S., N. Ijaz, N. Hashmi, M. I. Ullah, G. Afzal, A. Nasir, S. Perween, A. Sami. N. Tara, S. Abbas and S.A. Muhammad., 2023. Functional genomics analysis of leptin-melanocortin system genes reveals candidate genes associated rapid growth and high carcass yield in sheep. J. King Saud University-Sci., 35(8), 102853. <https://doi.org/10.1016/j.jksus.2023.102853>
23. Herdis, H., I. Inounu, S. Santoso, R. I. Anwar, S. Y. Hayanti, M. F. Hudaya, D. .A. .Mahari, F.B.I. Lupitasari, A. Hafid, M.A. da Costa and N. Adianto., 2025. Reproductive integration of leptin and Kisspeptin in small ruminants: Mechanisms, biomarker potential, and prospects for precision breeding. Veterinary World, 18(6), 1614 <https://doi.org/10.1186/s43088-023-00450-0>
24. Kaur, R., S. Garcia, A. Horadagoda and W. Fulkerson., 2010. Evaluation of rumen probe for continuous monitoring of rumen pH, temperature and pressure. Anim. Prod. Sci. 50(2): 98-104. <https://doi.org/10.1071/AN09048>
25. Kibar, M. and İ. Aytekin., 2024. Associations between leptin gene polymorphism and some reproductive traits in Holstein-Friesian dairy cattle. J. Hellenic Vet. Med. Soc., 75(4), 8163-8172. <https://doi.org/10.12681/jhvms.32289>
26. Loor, J. J., M. Vailati-Riboni, J. C. McCann, Z. Zhou and M. Bionaz., 2015. Triennial Lactation Symposium: Nutrigenomics in livestock: Systems biology meets nutrition. J. Anim. Sci., 93(12), 5554-5574. <https://doi.org/10.2527/jas.2015-9225>
27. Maruszak, A., P. Sablik, E. Janus and A. Dybus., 2023. Relationship between the leptin gene polymorphism and the productivity and health traits in Holstein-Friesian cattle. Acta Scientiarum Polonorum. Zootechnica, 22(4). <https://doi.org/10.21005/asp.2023.22.4.06>
28. Meira, A. N., G. C. M. Moreira, E. N. Coutinho, L. L., Mourão, G. B., Azevedo, H.C., Muniz, , A. L. Machado, L.P.S. Junior, V. B. Pedrosa, and L. F. B. Pinto, 2018. Carcass and commercial cut yield of Santa Inês sheep affected by polymorphisms of the LEP gene. Small Rumin. Res., 166, 121-128 <https://doi.org/10.1016/j.smallrumres.2018.06.012>
29. Naeemah, A. G. and N. N. Al-Anbari 2022. FASN gene polymorphism and its relationship with milk yield and composition in the Iraqi Awassi sheep. J. Kerbala Agric. Sci., (JKAS). 9(2): 34-45. <https://doi.org/10.59658/jkas.v9i2.960>
30. Nkrumah, J. D., C. Li, J. B. Basarab, S. Guercio, Y. Meng, B. Murdoch, C. Hansen, and S. S. Moore, 2004. Association of a single nucleotide polymorphism in the bovine leptin gene with feed intake, feed efficiency, growth, feeding behavior, carcass quality and body composition. Canadian J. Anim. Sci., 84(2), 211-219. <https://doi.org/10.4141/A03-033>
31. Nugroho, T., T. S. M. Widi and D. Maharani., 2022, February. The potency of leptin gene as a selection marker of economic traits for Madura cattle: preliminary study. In 9th Int. Semin. Trop. Anim. Prod., (ISTAP 2021), 231-237. <https://doi.org/10.2991/absr.k.220207.048>
32. Prihandini, P.W., A. P. Z. N. L. Sari, Tribudi, Y. A., Robba, D. K. and Wibowo,

T.B., 2024, May. polymorphisms of the leptin gene in Jabres cattle. IOP Conf. Ser: Earth Environ. Sci., 1341, 012008.

<https://doi.org/10.1088/1755-1315/1341/1/012008>

33. Raza, S.H.A., G.Y. Liu, L. Zhou, L. S. Gui, R. Khan, Y. Jinmeng, M. Chugang, N. M. Schreurs, R. Ji and L. Zan., 2020. Detection of polymorphisms in the bovine leptin receptor gene affects fat deposition in two Chinese beef cattle breeds. *Gene*, 758, 144957.

<https://doi.org/10.1016/j.gene.2020.144957>

34. Saleem, A. H., K. Javed, M. E. Babar, T. Hussain, A. Ali, A. Afzal, A. Nisar, M. Z. Farooq and M. Dawood., 2018. Association of leptin gene polymorphism with growth rate in Lohi sheep. *Pak. J. Zool.*, 50(3):1029-1033.

<http://dx.doi.org/10.17582/journal.pjz/2018.50.3.1029.1033>

35. SAS, 2018, Statistical Analysis System, User's Guide. Statistical. Version 9.4th ed. SAS. Inst. Inc. Cary. N.C. USA.

36. Sedykh, T. A., L. A. Kalashnikova, R. S. Gizatullin and V. I. Kosilov., 2020. Effects of leptin gene polymorphism on beef cattle performance. *Russian Agric. Sci.*, 46(6), 614-618.

<https://doi.org/10.3103/S1068367420060166>

37. Senturk, N., T. N. Selvi, M. Demir, H. Ustuner, H. Samli and S. Ardicli., 2024. The impact of LEP gene polymorphisms located at exon 2 (LEP-Hin fI) and intron 2 (LEP-Sau 3AI) on growth and reproductive traits in Saanen goats. *Archives Animal Breeding*, 67(4), 523-531.

<https://doi.org/10.5194/aab-67-523-2024>

38. Suárez-Mesa, R., R. Ros-Freixedes, R. N. Pena, J. Reixach and J. Estany., 2024. Impact of the leptin receptor gene on pig performance and quality traits. *Sci. Rep.*, 14(1), 10652.

<https://doi.org/10.5194/aab-67-523-2024>

39. Triantaphyllopoulos, K. A., I. Ikonomopoulos, and A. J. Bannister, 2016. Epigenetics and inheritance of phenotype variation in livestock. *Epigenetics and chromatin*, 9(1), 31.

<https://doi.org/10.1186/s13072-016-0081-5>

40. Ueta, I., Y. Nakamura, S. Kawakubo and Y. Saito., 2018. Determination of aqueous formic and acetic acids by purge-and-trap analysis with a needle-type extraction device and gas chromatography barrier discharge ionization detector. *Anal. Sci.*, 34(2), 201-205.

<https://doi.org/10.2116/analsci.34.201>

41. Yang, D., H. Chen, X. Wang, Z. Tian, L. Tang, Z. Zhang, C. Lei, L. Zhang and Y. Wang., 2007. Association of polymorphisms of leptin gene with body weight and body sizes indexes in Chinese indigenous cattle. *J. Genet Genomics*. 34(5): 400-405.

[https://doi.org/10.1016/S1673-8527\(07\)60043-5](https://doi.org/10.1016/S1673-8527(07)60043-5)

42. Yi, X., S. Yi, J. Wang, B. Ye, X. Zhu, Q. Meng, H. Wu, and Z. Zhou., 2025. Differential lipid metabolism in beef cattle: A comparative study of high and low residual feed intake bulls. *Anim. Nut.* 22, 214-229.

<https://doi.org/10.1016/j.aninu.2025.03.019>