NEW RECORD SPECIES OF GIANT FRESH WATER SHRIMP FROM THE DOWNSTREAM OF SERAYU RIVER

Baruna K.1S. B. Prayitno2A. Sabdaningsih3P. H. T. Soedibya1S. W. Saputra3ResearcherProf.Assist. Prof.Prof.Prof.1DenAssist. East Fish and Man SaiI. Saedisman Univ. Indenseis

¹Dep. Aqua., Fac. Fish. and Mar. Sci., J. Soedirman Univ., Indonesia.

²Dep. Aqua., Fac. Fish. and Mar. Sci., Diponegoro Univ., Indonesia. ³Dep. Aqua. Res., Fac. Fish. and Mar. Sci., Diponegoro Univ., Indonesia.

Email: barunakusuma@unsoed.ac.id

ABSTRACT

Indonesia is characterized by great diversity of freshwater shrimp, consisting of several islands due to the geographical location on the equator. A recent investigation conducted in Central Java has identified Serayu River as the major habitat of freshwater shrimp genus Macrobrachium. Therefore, this research aimed to determine the type of giant freshwater shrimp found downstream of Serayu River using explanatory method. The experiment was conducted for one year (January - December 2023) and the results found two freshwater giant prawn species, namely *Macrobrachium rosenbergii* and *Macrobrachium spinipes*. Moreover, this research is the first record of *M. spinipes* based on the Cytochrome C oxidase Subunit I (CO1) gene. The mtDNA segment of *M. rosenbergii* was 631 bp (base pairs) (GenBank accession: LC813231.1), while *M. spinipes* was 616 bp (GenBank accession: PP735157.1). *M. spinipes* and *M. rosenbergii* downstream of Serayu River showed similar locations and characteristics. Genetic distance conducted on 15 species of the genus Macrobrachium showed that giant freshwater prawn serayu had a matrix value of 0.09 - 0.18. Genetic analysis conducted with GenBank indicated that *M. spinipes* showed significant differences from other species, suggesting the natural dispersal of juvenile through currents across Wallace Line. This research also found that *M. rosenbergii* was similar to *M. dacqueti* and related to *M. malcolmsonii*, while *M. spinipes* was close to *M. suphanense*. To support these results, recommendation was made to explore further the rivers in the western part of Java, Indonesia.

Key words: Bio geography, DNA Barcoding, M. rosenbergii, M. spinipes, Serayu River, life below water

المستخلص

يوجد في إندونيسيا تنوع كبير لروبيان المياه العذبة. ويرجع ذلك إلى موقعها الجغرافي على خط الاستواء وتكوينها من عديد الجزر. نهر سيرايو هو أحد الأنهار الكبيرة في جاوة الوسطى وهو شيمة لجنس الروبيان *Macrobrachium*. الهدف من هذا البحث هو تحديد أنواع روبيان المياه العذبة العملاق الأنهار الكبيرة في جاوة الوسطى وهو شيمة لجنس الروبيان *Macrobrachium*. الهدف من هذا البحث هو تحديد أنواع روبيان المياه العذبة العملاق الموجود في مجرى نهر سيرايو. الطريقة المستخدمة في هذا البحث هي الطريقة الاستكشافية. تم إجراء البحث في اتجاه مجرى نهر سيرايو. مدة البحث من الروبيان العملاق الذي يعيش في المياه العذبة، وهما *Macrobrachium سنة واحدة (يناير – ديسمبر 2023). وجدت نتائج البحث نوعين من الروبيان العملاق الذي يعيش في المياه العذبة، وهما Macrobrachium spinjes وتعويم واحدة (يناير – ديسمبر 2023). وجدت نتائج البحث هو أول سجل له <i>spinjes الاستكشافية*. تم إجراء البحث في المياه العذبة، وهما *Macrobrachium spinjes وتعوام ويعيش في المياه العذبة وهما Macrobrachium spinjes وعوام spinjes واحدة (يناير – ديسمبر 2023). وجدت نتائج البحث هو أول سجل له <i>spinjes الم spinjes المي إلى والي يعيش في المياه العذبة وهما Macrobrachium spinjes وتعون معيش واحدة (إلي مولا يعيش في المياه العذبة واحدة (إلي منه معنا المعدة الفرعية المحمودة الفرعية <i>orsenbergii mt Sinjes الله ما spinjes الله بين العملاق الذي يعيش في المياه العذبة (Con) ا. كان مقطع An rosenbergii mt DNA وiلوج أساس (أزواج أساسية) (انضمام spinjes الغربي العملاق الزواج أساسية) (انضمام spinjes). يُظهر spinjes المعنية العادية سيرايو السلي ينفي لموقع والخصائص. أظهرت المسافة الوراثية التي أجريت على 15 نوعا من جنس <i>spinjes مع spinjes الم وبيان الما وي والملي والله والي العربي الموري والملي والملي يعيش المورات الموالي الم والي أوريت على 15 مورات الي مي والملي والي أن روبيان الما الطبيعي ليرقات 15 مي جلي والمليق مولية والمطابقات مع Spinjes ألي ألي الما مولا ولي ولي المي والملي والي أل والي والملي والملي والملي والسلي في في المورات الخربي الموراثي التولي الوراثي الذي تم إجراؤ والملالي . وولول ال الموراثي العد ما معداق مع الحزفي في الموقع والخصاص. أظهرت المالمي يعي ليروات Senak. <i>Spinjes هي وحوي وال والي أل وال الي و*

الكلمات المفتاحية: الجغرافيا الحيوبة، ترميز الحمض النووي، M. Spinipes ، M. rosenbergii، نهر سيرايو، الحياة تحت الماء

INTRODUCTION

Rivers have a very vital role in life (6). Rivers can be used for industrial, agricultural, and fisheries (55; 39). One of the rivers in Central Java Province is the Seravu River. This river has a length of 153 km, crosses 5 districts, and empties into Cilacap Regency (52). This river is commonly used to irrigate rice fields. Given the importance of this river for people's lives, it is also very important to know the biodiversity of aquatic organisms in the Serayu River. One of the aquatic organisms present in this river is the giant freshwater shrimp. Macrobrachium is a highly diverse group of freshwater shrimp found in the Indo-Pacific region (13). The genus includes aapproximately 240 identified species, where more than 100 are found in East and Southeast Asia (3). It is also known as one of the freshwater crustacean species with high species variation (63). The comprehensive geographical distribution of the species has shown significant morphological and life history variation, with the most differences found on both sides of Wallace Line, which divides Asian and Australasian fauna (49). Proposed that freshwater shrimps be recognized two distinct subspecies: as Macrobrachium rosenbergii (sensu stricto), which is distributed in eastern Indonesia, New Guinea, Australia and the Philippines, and Macrobrachium rosenbergii dacqueti (Sunier, 1925), which is found in Sunda Southeast Asia, Indochina and South Asia. However, Wowor and Ng (64) provided a different classification, suggesting that the two subspecies should be separately considered due to the non-overlapping geographical distributions and distinctive morphological characteristics. These include relative height of the rostral base, shape of the second to fifth pereiopods in adult shrimp, the shape of the fourth thoracic sternite, and the coloration pattern in live shrimp. Wowor and Ng (65) stated the confusion in naming Palaemon spinipes, considered the same initially as Macrobrachium wallacei. To clarify the confusion, further investigation by Ng and Wowor (35) identified P. spinipes as species with attractive color, which is often caught and traded as an ornamental fish. This led to the division of the genus Macrobrachium into two species based on Wallace Line, comprising west and east species, namely M. rosenbergii and M. spinipes. In Indonesia, the large number of Macrobrachium has the potential to remain molecularly unidentified, showing the need for DNA barcoding to identify cryptic and undescribed species (556). Species identification using DNA provides several science. in biological benefits where cytochrome c-oxidase subunit I (COI) is commonly used as the standard gene. COI gene has been proven to provide convenience in amplifying and providing information to identify organisms at the species level. Very little is known about freshwater shrimp in the Serayu River. According to Siregar (51), 6 species of Macrobrachium were found in the tributaries of the Serayu River (Pelus River and Banjaran River). According to Kusbiyanto (27), the last study left 4 species found in the tributary of the Serayu River (Banjaran River). Both references above identify freshwater shrimp in the upstream tributaries of the River. However, Serayu the genus Macrobrahium's existence in the river's lower reaches has yet to be widely discussed. This study aims to determine the type of freshwater prawns found in the lower reaches of the Serayu River. Information in this study is very important and will be needed in the development of science. Similar research has never been conducted at this location. However, crustacean larval DNA research has been conducted by Winarni et al. (61), whose research location is close but has a different upstream and ecology.

MATERIALS AND METHODS

Study Area: This research was conducted for one year (January to December 2023) downstream of Serayu River with coordinates -7.525722 S, 109.200998 E to -7.579252 S, 109.148062 E (Figure 1). Shrimp were captured at night (18.00-24.00) and morning (06.00-10.00), using nets, fishing, and traps Captured shrimp were put in an aerated coolbox and taken to the laboratory for maintenance as well as morphological identification. The collection results were morphologically identified to determine giant freshwater shrimp species that were caught. Meat tissue samples (\pm 10 g) were preserved in separate 50 mL bottles containing 96% ethanol technical and labeled (47). Subsequently, shrimp samples identified based on morphology were analyzed for DNA

in the BIONESIA laboratory.



Figure 1. Research location on the Serayu River hill (with organic points -7.525722 S, 109.200998 E to -7.579252 S, 109.148062 E with freshwater properties to -7.579954 S, 109.146195 E to -7.686207 S, 109.101048 E with brackish water properties.

Procedures Morphological **Analysis:** Captured shrimp were morphologically differentiated based on rostrum shape, number of rostrum serrations, and color pattern. The catch results were distinguished based on differences in morphology using shrimp identification book. Additionally, morphological identification was carried out in this research following the guidelines of Short (49)

DNA analysis: The extraction method used in this research followed the Qiagen protocol. After DNA was extracted, the next step was amplifying specific DNA segments using PCR (Polymerase Chain Reaction) following **BIONESIA** laboratory guidelines. The primers used for amplification of shrimp samples were jgLCO (5'-TIT CIA CIA AYC AYA ARG AYA TTG G-3') and jgHCO (5'-TAI ACY TCI GGR TGI CCR AAR AAY CA-3'). according to Geller (14). PCR reactions were performed in a total volume of 26 µL, consisting of 2 µL of extracted DNA template, 1.25 µL of each primer with a concentration of 10 mM, 9 µL of ddH2O, and 12.5 µL of PCR buffer. The amplification process used an Applied Biosystems[™] 2720 Thermal Cycler with a temperature and time profile, including initial denaturation at 94°C for 3 minutes, by consisting followed 38 cycles of denaturation at 94°C for 30 seconds, annealing at 52°C for 30 seconds, and extension at 72°C for 60 seconds, and final extension at 72°C for 2 minutes. PCR products were visualized using a 1% agarose gel stained with Nucleic Acid Gel Stain (GelRed®). Visible DNA bands showed successful amplification results and positive samples were sent to PT Genetika Science Jakarta for sequencing using the Sanger method.

Data analysis: The process of editing the forward and reverse sequences of the various morphotypes using BioEdit version 7.2.5 (21) started with removing errors and ambiguities at the ends of the sequences caused by the primers. Subsequently, these sequences were used to create consensus sequences reflecting the various dominant samples. COI coding region was identified using an online ORF finder (https://www.ncbi.nlm.nih.gov/orffinder/) with parameters such as a minimum length of 300 nucleotides and mitochondrial invertebrate genetic code and verified using BLAST to ensure the absence of unwanted stop codons. Sequence arrangement was carried out using ClustalW as done by Alam (4) which was available in BioEdit, to ensure consistency and accuracy between sequences in species barcode analysis (21). The identity and similarity nucleotide of sequences of Macrobrachium species were analyzed using BLAST through NCBI server and BOLD for homology matching. Based on analysis, identified sequences were arranged using CLUSTAL Omega for multiple sequence arrange. Phylogenetic analysis was performed using MEGA X software, with phylogenetic constructed using the maximum trees parsimony method. Confidence in phylogenetic tree topology was evaluated through bootstrap analysis with 1000 replicates to provide an in-depth view of the evolutionary relationships among species in the genus Macrobrachium.

RESULTS AND DISCUSSION Morphological Identification

This research successfully identified two species of giant freshwater shrimp, namely *M. rosenberrgii* and *M. spinipes*, as shown in <u>Figure 2</u>. Therefore, 20 *M. rosenbergii* and 20 *M. spinipes* were successfully collected downstream of Serayu River. This is consistent with the characteristics of *M. rosenbergii* as done by Wowor and Ng (64) and *M. spinipes* Ng and Wowor (35).



Figure 2. Giant Freshwater prawn is found downstream of Serayu River. A) *M. rosenbergii* and B) *M. spinipes*

The results of morphological identification showed significant differences in juvenile *M. rosenbergii* and *M. spinipes*, which included rostral teeth, rostrum, cephalothorax, body, and pole claw color, as presented in <u>Table 1</u>:

Fable 1. Differences between M	rosenbergii and M. spinipes juvenile.
---------------------------------------	---------------------------------------

Characters	Juvenile <i>M. spinipes</i>	Juvenile M. rosenbergii
Rostral ventral teeth	6 - 16 with mode 12	6 - 15 with mode 9
Rostral dorsal teeth	8 - 15 with mode 13	8 - 14 with mode 12
Rostral	Lower rostral crest	Higher rostral peak
Color of rostrum	There is a red line from the base to the tip of	Plain grey
	the rostrum	
Color of cephalothorax	Reddish, there is a longitudinal solid black	Gray has faint longitudinal
	line	black lines
Body color	Reddish, there is a dotted black line	Gray with transverse lines in
	longitudinal to the base of the tail	each segment
Claw color	There is a black and white look pattern on	Grey and occasionally a slight
	the tip of the claw	reddish tint to the segments

Giant Freshwater prawn DNA Barcode The results of the molecular analysis in <u>Table 2</u> showed that the mtDNA segment of *M. rosenbergii* in this research was 631 bp (base pairs) (GenBank accession: LC813231.1), with the percentage of nucleotides T(U): 26.49%, C: 26.49%, A: 26.67%, and G: 20.18%. Meanwhile, the mtDNA segment of *M*. *spinipes* was 616 bp (GenBank accession: PP735157.1), comprising T(U): 28.60%, C: 23.68%, A: 29.12%, and G: 18.60%. The average nucleotide composition of the genus Macrobrachium used as a comparison was T(U): 28.05%, C: 24.97%, A: 28.04%, G: 18.95% (Table 3).

Table 2. DNA barcoding of giant freshwater	prawns in the lower reaches of the Serayu River
DNA Barcoding of <i>M. spinipes</i> Seravu PP735157.1	

DNA Barcoding of M. rosenbergii Serayu LC813231.1

Table 3. Nucleotide composition of the genus Macrobrachium used in this research

GenBank Code and Species	T(U)	С	Α	G
PP735157.1 M. spinipes	28.60	23.68	29.12	18.60
LC813231.1 M. rosenbergii	26.67	26.49	26.67	20.18
MK782941.1 M. rosenbergii	26.67	26.49	26.67	20.18
MT235937.1 M. sintangense	27.02	24.91	28.77	19.30
MF622028.1 M. suphanense	30.70	22.63	28.77	17.89
MK412772.1 M. nipponense	29.47	22.46	29.47	18.60
MW845505.1 M. lanchesteri	30.88	22.28	28.77	18.07
OM273995.1 M. malcolmsonii	27.37	24.91	29.12	18.60
FM958058.1 M. asperulum	30.35	21.40	30.35	17.89
MT235944.1 M. neglectum	28.42	24.04	28.77	18.77
JF792432.1 <i>M. dacqueti</i>	26.67	26.49	27.37	19.47
MN526206.1 M. esculentum	25.96	26.67	27.72	19.65
ON753700.1 M. equidens	28.95	25.44	27.89	17.72
MW845507.1 M. latidactylus	28.60	26.49	25.61	19.30
MW845472.1 M. forcipatum	24.39	30.18	25.44	20.00
Average	28.05	24.97	28.04	18.95

Notes: The red color is the nucleotide composition of Macrobrachium.

Identification of Similarity

The maximum genetic divergence to the reference species in GenBank or BOLD system is less than the predetermined value of 3% (61). These results are consistent with previous decisions on the taxonomic status of the samples, which show identical characteristics to similar references in the

database. Match results with GenBank showed that *M. spinipes* in Serayu River had an identity value of 95.62%, but BOLD system showed no significant value. Meanwhile, *M. rosenbergii* with an identity value of 100% was considered true, as shown in Table 4.

Table 4. Similarit	v of the genus	Macrobrachium	downstream of	f Seravu River
				•/

	Reference and accession	GenBank	K	BOLD			
	number	Query Coverage (%)	Identity (%)	Similarity (%)	Results Summary		
	M. spinipes PP735157.1	100	95.62	-	No match		
	M. rosenbergii LC813231.1	100	100	100	M. rosenbergii		
G	iant Freshwater Prawn I	Phylogenetics	suphanense	e. Similarly,	M. rosenbergii from		

Phylogenetic tree reconstruction results show that *M. spinipes* found downstream of Serayu River is most closely related to *M*. *suphanense*. Similarly, *M. rosenbergii* from this research has the same bootstrap value, with *M. dacqueti* showing a close relationship with *M. malcolmsoni*, as presented in Figure 3.

Genetic Distance

Genetic distance refers to the results of geneticdifferences between species. The analysis conducted on 15 species of the genus Macrobrachium shows a matrix value of 0.09 -0.18. with the closest distance in M. suphanense (0.09) and the farthest distance in M. rosenbergii (0.18), as shown in Table 6. Previous research showed that smaller genetic distance value correlated with more similar COI partial genes between species (8).

Table 6 Constic	distance of the	anus Macrah	rachium hase	d on COI gene
Table 0. Genetic	uistance of the	genus Macron	pracinum base	a on COI gene.

GenBank Code and	1	2	3	4	5	6	7	8	9	10	11	12	13	14
Species			-		-			-		-			-	
PP735157.1 M. spinipes	*	*	*	*	*	*	*	*	*	*	*	*	*	*
LC813231.1 <i>M</i> .	0.18	*	*	*	*	*	*	*	*	*	*	*	*	*
rosenbergii	0110													
MK782941.1 <i>M</i> .	0.18	0.00	*	*	*	*	*	*	*	*	*	*	*	*
rosenbergu														
МТ235937.1 М.	0.09	0.18	0.18	*	*	*	*	*	*	*	*	*	*	*
sintangense	0.05	0120	0120											
MF622028.1 M.	0.09	0.18	0.18	0.10	*	*	*	*	*	*	*	*	*	*
MKA12772 1 M														
	0.12	0.18	0.18	0.11	0.13	*	*	*	*	*	*	*	*	*
mpponense														
NI W 845505.1 M.	0.15	0.14	0.14	0.18	0.15	0.16	*	*	*	*	*	*	*	*
lanchesteri														
OM2/3995.1 <i>M</i> .	0.15	0.14	0.14	0.17	0.17	0.16	0.14	*	*	*	*	*	*	*
maicoimsonii														
FM958058.1 <i>M</i> .	0.15	0.18	0.18	0.15	0.14	0.17	0.15	0.15	*	*	*	*	*	*
asperulum														
MT235944.1 <i>M</i> .	0.12	0.16	0.16	0.14	0.13	0.12	0.15	0.15	0.15	*	*	*	*	*
neglecium	0.17	0.03	0.03	0.10	0.17	0.10	0.14	0.12	0.10	0.16	*	*	*	*
JF /92432.1 M. aacquen	0.17	0.02	0.02	0.18	0.17	0.18	0.14	0.13	0.18	0.10	T	Ŧ	Ť	Ŧ
MN526206.1 M. esculentum	0.14	0.18	0.18	0.15	0.16	0.13	0.18	0.15	0.16	0.14	0.17	*	*	*
ON753700 1 M equidens	0 16	0 17	0 17	0.15	0 16	0 17	0 16	0.18	0.16	0 16	0 17	0.16	*	*
MW845507 1 M	0.10	0.17	0.17	0.10	0.10	0.17	0.10	0.10	0.10	0.10	0.17	0.10		
latidactylus	0.15	0.18	0.18	0.17	0.18	0.17	0.19	0.17	0.19	0.16	0.17	0.17	0.18	*
MXXQA5A77 1 M														
191 99 0434 / 2.1 191. foreingtum	0.17	0.18	0.18	0.17	0.18	0.17	0.20	0.19	0.19	0.17	0.18	0.18	0.18	0.17
յուսրաստ														



→ 0.02

Figure 3. Phylogenetic tree of giant freshwater shrimp downstream of Serayu River, Indonesia. The red box line shows that *M. rosenbergii* is the same as *M. dacqueti*.

Habitat

M. spinipes and M. rosenbergii downstream of Serayu River show similar locations and characteristics. Both shrimp migrate to brackish water to spawn, and juvenile return upstream (57). Rivers with muddy and sandy

characteristics dominate habitat characteristics in freshwater downstream of Serayu River. However, the characteristics of Serayu River estuary are dominated by sandy and muddy waters, as shown in Figure 4.



Figure 4. Habitat of *M. rosenbergii* and *M. spinipes*. A. Downstream of Serayu River with fresh water; B. Downstream of Serayu River with brackish water.

The distribution of M. spinipes requires detailed exploration, as previous investigations focus on the pattern by dividing M. rosenbergii and M. spinipes based on Wallace Line (50). Wallace region is an essential biogeographic zone that includes Sulawesi, Lombok, Sumbawa, and Flores. It also includes other small islands located between Sundaland such as Malay Peninsula, Sumatra, Kalimantan, Java, and Bali, as well as the Sahul region comprising Australia and Papua New Guinea (21). Wallace Line passes through the Lombok Strait, between Bali and the Lombok Islands, and through Makassar which separates Kalimantan and Strait. Sulawesi. The boundary of Wallacea is

M anininaa

marked by Lombok Strait, the deep, and wide Makassar Strait in the west, with Australo-Papuan Shelf in the east (26). This region comprises several islands, including Sulawesi, Lesser Sunda, Halmahera, Buru, Seram, and other small islands. *M. rosenbergii* is found west of Wallace Line and *M. spinipes* is east. The discovery of *M. spinipes* downstream of Serayu River remains complex due to the location of the river in Central Java, west of Wallace Line, compared to the expected pattern based on line separation. Based on previous research, the distribution of *M. rosenbergii* and *M. spinipes* is listed in <u>Table 5</u> and figure 5.

Mi. spinipe	63	M. Tosenbergu		
Research Location	Reference	Research Location	Reference	
Australasia Eastern Sh	ort (49); <u>De Bruyn</u>	Coastal freshwaters throughout	Lindenfelser (29)	
Wallace Line and the(18)	<u>8);</u> Ng and Wowor	India, northern Australia, and		
Philippines (35	5)	Southeast Asia		
Daly River in No	ovak (38); Novak (36);	Western Pakistan, southern	De Bruyn (18); De	
Australia's Northern No	ovak (37)	Vietnam, Southeast Asia,	Bruyn (17)	
Territory		southern Asia, northern		
		Australia, New Guinea, Pacific		
		and Indian Oceans		
Papua New Guinea Pic	ckering (41)	Halmahera Island, Maluku	Cai (11)	
_		archipelago, Indonesia		
Fitzroy River, Be	eesley (9)	East Africa	Kuguru (26)	
Kimberley, Western				
Australia				
Fitzroy River, northern Th	orburn (53)	Myanmar	Cai (12)	
Australia		-		
Fitzroy River, Daly Jac	ckson (25)	South, Southeast Asia, and	Wowor (62); Wowor	
River, Mitchell River		Indochina	(64)	
Northern Australia				
Serayu, Central Java Th	nis research	Serayu, Central Java	This research	

 Table 5. Distribution of M. spinipes and M. rosenbergii results

Figure 5. Distribution of Macrobrachium. Red: M. rosenbergii; Green: M. spinipes of Wallace Line, which includes Java Island,

Discussion

Giant freshwater shrimp have gained significant attention across various countries due to the great potential in aquaculture (24). Recent developments show that freshwater shrimp farming contributes to global fisheries in quality and value (34). Morphological identification showed the presence of M. rosenbergii and M. spinipes downstream of Serayu River. Specifically, M. spinipes juvenile have very different coloration and chromatophore patterns, namely red rostrum, transparent body with several longitudinal black lines on the carapace, and black spots on the abdomen. The difference in color patterns in juvenile in these two shrimp has also been described by Shy (50). Based on the results, M. spinipes juvenile from Papua Indonesia (male, 46.9 mm cl) and Sulawesi showed identical color patterns, with several longitudinal dark brown lines on the carapace, a red rostrum, and a belly decorated with dark brown spots. As described by Shy (50) in Taiwan, the body coloration of adult M. spinipes varied depending on individual size. The body is blue-green in more prominent individuals (carapace length \geq 50 mm), while yellowgreen is observed in smaller individuals (carapace length about ≤ 35 mm). The downstream of Serayu River at the west of Wallace Line have waters with a rocky, sandy bottom, while muddy waters are observed in the estuary. M. rosenbergii is distributed west showing the natural presence in the river. Wowor and Ng (64) stated that M. spinipes should be found east of Wallace Line due to natural dispersal or human intervention such as trade in ornamental fish and the release of wild species (5, 33). De Bruyn (18) also stated that significant genetic differences between species in the Philippines, Indonesia, New Guinea, and were influenced during Australia the Pleistocene. The global climate change caused large sea-level fluctuations, which had a significant impact on the geography and ecology of the Indo-Australian region. At the end of the Pleistocene, approximately 20,000 years and 10,000 years after sea level rise began in the Pacific Ocean, the current level increased in 6000 - 7000 years ago. This event occurred around 7500 - 10,000 years ago for populations in Western and Eastern Indonesia. Sunda region had a shallow continental shelf. which was repeatedly to large islands due to eustatic changes in sea level (60). Therefore, the migration of marine crustaceans to and from the Indonesian Archipelago probably occurred during the last ice age (10,000 -70,000 years ago) (21). Due to the complications of geology and climate history of transition areas such as Wallace Line regions, it is difficult to conduct an investigation (42), including several smaller biogeographic units (20). Despite the fact that this research indicated currents had an impact

on the dispersal of *M. spinipes* juvenile, it was to consider the great distance to the west Wallace Line. The results showed a strong correlation between larval dispersal as well as distance and duration of each species (23). Research has demonstrated that certain taxa can cross the Wallace Line in both directions. Notable examples include fanged frogs (19) and rodents (43) from the west, as well as avian butterflies (15), beetles (54), and more recently borer beetles (16) from the east. Pelagic larvae of marine organisms living in open water have been found to be crucial to the migration and dispersal of ocean species (45). This is due to the fact that larvae's longdistance capacity facilitates migration to new regions, which facilitates marine biodiversity. The ITF travels from the Pacific to the Indian Ocean, passing through the Makassar Strait and into Indonesia water (46). A tiny amount of the ITF also enters the South China Sea, the Karimata Strait, and the Lifamatola Strait into the Indian Ocean through Lombok, Ombai, and Timor Straits and exits through Sunda Strait in summer. Investigation has observed that the ITF reaches Sumatra (31), showing the significance of the currents on larval dispersal. Previous research established that giant freshwater shrimp larvae were capable of surviving high salinity conditions. According to Novak (38), *M. spinipes* larvae possess greater survival than other Macrobrachium except M. which survive amazonicum. can for approximately 11 days. Although the juvenile of M. rosenbergii and M. spinipes thrive in brackish habitats, the adults of these species are found downstream of the Serayu River in freshwater environments. Amphiboly is a stage in the life cycle of shrimp, where adults in freshwater release juvenile that float to estuarine areas. As part of a life history that is frequently observed in tropical and subtropical fish (1, 39), including crustaceans in rivers (9), these shrimp will transform into juvenile and travel upstream (48). According to Novak (37), recent research has shown that shrimp can release juvenile to approximately 400 km downstream, with the highest concentrations found downstream of Serayu River. M. seasonally, spinipes spawn having reproductive activity from November to April and peak in January, with gonad maturation starting at 80 mm in length. Downstream of Serayu River, there are 435 M. rosenbergii and 30 M. spinipes recorded during the one-year research. The presence of M. spinipes in Segera Anakan River, Central Java, was supported by the results of Winarni et al. (61), which identified Macrobrachium at the site. This river has mangrove forests and is adjacent to Serayu River, with similar ecological characteristics. Sequences can be identified as the same species when a specimen is 98-100% similar to species found in GenBank (7). M. rosenbergii of Serayu River is a valid species because it has 100% similarity but not M. spinipes, as shown in Table 4. Since this research is the first to register M. spinipes in GenBank, there is difficulty in the similarity comparison with M. spinipes found east of Wallace Line. According to the standards the genetic distance between the shrimp of the genus Macrobrachium used for comparison with the shrimp from this research was classified as low (0.01-0.09) to medium (0.1-0.99) (Table 6). These genetic distance values showed that partial sequences of CO1 gene could provide a fast and efficient DNA barcoding method for species identification, particularly in overcoming confusion or ambiguity related to morphology (8). Genetic variation has become a source of evolutionary innovation that can facilitate reproductive isolation. thereby forming parapatric speciation (32). The existence of a significant genetic structure in giant freshwater shrimp downstream of Serayu River with other Macrobrachium genera is also influenced by the main ocean currents in each region, such as Kuroshio, Mindanao, Cross Indonesia, and East Australia Current (22). Significant differences were also observed in fiddler crabs (Austruca perplexa) (22) on Wallace Line explored reef fish (Casio cuning) (2) and found a significant genetic difference between populations in Sumatra and Java in the western part of Indonesian archipelago and populations in Maluku Islands located on the eastern side Wallace. Previous investigations of of Polycarpa aurata on coral reefs showed that genetic variation spread across Wallacea region. This research used microsatellite markers and suggested that genetic differences were due to the highly diverse environment

and the short duration of pelagic larvae (58). Significant population patterns have also been observed in other crustaceans in Wallacea region. For example, microsatellite analysis on ribbon crayfish (Stenopus hispidus) showed substantial population differences despite a relatively long pelagic larval duration of approximately 210 days (59). Several biological characteristics, such as typical reproductive mode, egg type (benthic or pelagic), variation in reproductive success, as well as the ability of adults to disperse, have been shown to influence patterns of population genetic structure in species (40). Additionally, local adaptation to specific environmental or habitat factors can lead to the evolution of genetically distinct lineages in a species without considering the potential influence of external oceanographic factors and pelagic larval duration (4). Giant freshwater shrimps often release the larvae in the water column which are frequently washed away by the current. The larvae will move and highly depend on the speed of the current, thereby causing minimal genetic differentiation in a population. The wide dispersal of larvae from different marine taxa is a significant factor that can have a major impact on genetic variation, structure, and connectivity (22). Phylogenetic relationships in the genus Macrobrachium remain unexplored (30). This research found that *M. rosenbergii* was similar to *M. dacqueti* and closely related to *M. malcolmsonii* (Figure 3). The result was consistent with the research conducted by Kuguru (26) in East Africa using COI and 16S markers. Wowor (63) also observed a close relationship between M. rosenbergii and M. dacqueti, M. malcolmsonii, and M. lanchesteri, while M. spinipes formed a distinct Claude closest to M. suphanense. The phylogenetic analysis also showed that M. suphanense was closely related to М. sintangense, as observed by Saengphan (44) in Thailand.

CONCLUSIONS

In conclusion, this research found two species of giant freshwater shrimp downstream of Serayu River were found, namely *M. rosenbergii* and *M. spinipes*. The result showed that mtDNA fragment of *M. rosenbergii* was 631 bp, while *M. spinipes* was 616 bp. The presence of *M. spinipes* downstream of Serayu River was different from previous theories because Serayu River was located west of Wallace Line. However, there were no matching data based on the match results with GenBank and BOLD. Genetic distances performed on 15 species of the genus Macrobrachium showed that giant freshwater shrimp had a matrix of 0.09 - 0.18. Long genetic distances suggested natural dispersal in M. spinipes juvenile through currents across Wallace Line. M. rosenbergii was similar to M. dacqueti and closely related to M. malcolmsonii while M. spinipes was closest to M. suphanense. Additionally, it was observed that natural dispersal in M. spinipes juvenile was caused by larval reversal. To strengthen these results, further research was recommended on the presence of M. spinipes in the western part of Java Island, Indonesia. REFERENCES

- Al-Khshali, M. S., and S. A. Faisal. 2023. Study of the Chemical Properties of the Meat of some Fish Imported to Iraq. IOP Conference Series: Earth and Environmental Science, 1262 (7), 0720892.
- Ackiss, A.S., S. Pardede, E.D. Crandall, M. Carmen A. Ablan-lagman, and N. Romena. 2013. Pronounced genetic structure in a highly mobile coral reef fish, caesio cuning, in the coral triangle. Marine Ecology Progress Series, 480(4):185–97.

https://doi.org//10.3354/meps10199.

3. Akinwunmi, M.F., and R. O. Moruf. 2021. Aspects of the bionomics of brackish water prawn (*Macrobrachium macrobrachion*: Herklots 1851) in the Interconnecting Lagoons of South-Western Nigeria. Egyptian Journal of Basic and Applied Sciences, 8(1):222–34. https://doi.org//10.1080/2314808X.2021.1956 186.

4. Alam, M.M. Mahbub, K.M. Westfall, and S. Pálsson. 2017. Historical demography and genetic differentiation of the giant freshwater prawn *Macrobrachium rosenbergii* in Bangladesh Based on mitochondrial and DdRAD sequence variation. Ecology and Evolution, 7(3):4326–35.

https://doi.org//10.1002/ece3.3023.

5. Al-Khshali, M.S. and A. B. Ibrahim. 2019. Study of the chemical composition of common carp fish reared in different culture systems. Plant Archives, 19, 1816–1818 6. Alsaadoon, D.W.K., F. M. Hassan, W. M. Mahdi. 2023. Assessement of water quality of Diyala River using overall index of pollution (OIP) in Iraq. Iraqi Journal of Agricultural Sciences 54(3):682–690.

https://doi.org/10.36103/ijas.v54i3.1748.

7. Astuti, S.S, A.M Hariati, W.E. Kusuma, A. Yuniarti, N. Kurniawan, and D.G.R Wiadnya. 2023. Anthropogenic introduction of the *Spotted barb*, *Barbodes binotatus*, across the Wallace Line in Western Sulawesi, Indonesia. Biodiversitas, 24(3):1916–25.

https://doi.org//10.13057/biodiv/d240369.

8. Basith. A, Abinawanto, E. Kusrini, and Yasman. 2021. Genetic diversity analysis and phylogenetic reconstruction of groupers *Epinephelus Spp*. from Madura Island, Indonesia based on partial sequence of Co1 Gene. Biodiversitas, 22(10):4282–90. https://doi.org//10.13057/biodiv/d221020.

9. Beesley, L.S., S. Killerby-Smith, D.C. Gwinn, B.J. Pusey, M.M. Douglas, P.A. Novak, T.C. Tayer, C.S. Keogh, M.J. Kennard, C.A. Canham, and S.A. Setterfield. 2023. longitudinal Modelling the distribution. abundance, and habitat use of the giant freshwater shrimp (Macrobrachium spinipes) in a large intermittent, tropical Australian River to inform water resource policy. Freshwater Biology, 68(1):61-76. https://doi.org//10.1111/fwb.14009.

10. Buckley, S. James, C. Brauer, P.J. Unmack, M.P. Hammer, and L.B. Beheregaray. 2021. The roles of aridification and sea level changes in the diversification and persistence of freshwater fish lineages. Molecular Ecology, 30(19):4866–83. https://doi.org//10.1111/mec.16082.

11. Cai, Y., and P.K.L. Ng. 2001. The freshwater decapod crustaceans of Halmahera, Indonesia. Journal of Crustacean Biology, 21(3):665–95. https://doi.org//10.1651/0278-0372(2001)021[0665:tfdcoh]2.0.co;2.

12. Cai, Yixiong, and Peter K.L. Ng. 2002. The freshwater palaemonid prawns (Crustacea: Decapoda: Caridea) of Myanmar. Hydrobiologia, 487(2):59–83.

https://doi.org//10.1023/A:1022991224381.

13. Castelin, M, V. Mazancourt, G. Marquet, G. Zimmerman, and P. Keith. 2017. Genetic and morphological evidence for cryptic species in *Macrobrachium australe* and

resurrection of *M. Ustulatum* (Crustacea, Palaemonidae). European Journal of Taxonomy, 2017(289):1–27.

https://doi.org//10.5852/ejt.2017.289.

14. Chan, T.Y. 1998. Shrimps and prawns. FAO species identification guide for fishery purposes. in: carpenter ke, Niem VH (Eds).

15. Condamine. F.L., E.F.A. Toussaint, A.L Clamens, G. Genson, F.A.H. Sperling, and G.J. Kergoat. 2015. Deciphering the evolution of birdwing butterflies 150 years after alfred russel Wallace. Scientific Reports, 5:1–11. https://10.1038/srep11860

16. Detsch, H, M. Balke, E.F.A. Toussaint, R.P. Narakusumo, K. Fiedler, and A. Riedel. 2020. Transgressing Wallace's Line brings hyperdiverse weevils down to earth. Ecography, 43(9):1329–40.

https://10.1111/ecog.05128

17. De Bruyn, M., E. Nugroho, M.M Hossain, J.C. Wilson, and P.B. Mather. 2005. Phylogeographic evidence for the existence of an ancient biogeographic barrier: the isthmus of kra seaway. Heredity, 94(3):370–78. https://doi.org//10.1038/sj.hdy.6800613.

18. De Bruyn, M, J.A. Wilson, and P.B. Mather. 2004. Huxley's Line demarcates extensive genetic divergence between Eastern and Western forms of the giant freshwater prawn, *Macrobrachium rosenbergii*. Molecular Phylogenetics and Evolution, 30(1):251–57.

https://doi.org//10.1016/S1055-7903(03)00176-3.

19. Evans. B.J., R.M. Brown, J.A. McGuire, J.Supriatna, N. Andayani, A. Diesmos, D. Iskandar, D.J. Melnick, and D.C. Cannatella. 2003. Phylogenetics of fanged frogs: testing biogeographical hypotheses at the interface of the Asian and Australian faunal zones. Systematic Biology, 52(6):794–819. https://10.1080/10635150390251063.

20. Ferro, I, and J.J. Morrone. 2014.

Biogeographical transition zones : a search for conceptual synthesis. Biological Journal of the Linnean Society, 113:1–12.

21. Hall, Robert. 2013. The palaeogeography of sundaland and wallacea since the late jurassic. Journal of Limnology 72(S2):1–17. https://doi.org//10.4081/jlimnol.2013.s2.e1.

22. Hardianto E, D.P. Wijayanti, J.Y. Shy, P. Mather, J. Hughes, and H. Imai. 2021.

Molecular ecology of the fiddler crab *Austruca perplexa* (H. Milne Edwards, 1852): genetic divergence along a major biogeographical barrier, Wallace's Line. Biological Journal of the Linnean Society, 135(2):310–21. https://doi.org//10.1093/biolinnean/blab142.

23. Hoarau, P.E., C.R.M. Treilhes, and P.B. Valade. 2018. Reproductive biology and recruitment in an amphidromous prawn *Macrobrachium australe* in Reunion Island. invertebrate biology, 10(10):1–9.

https://doi.org//10.1111/ivb.12222.

24. Ibrahim S, Z. Zhong, X. Lan, J. Luo, Q. Tang, Z. Xia, S. Yi, and G. Yang. 2023. Morphological diversity of different male morphotypes of giant freshwater prawn *Macrobrachium rosenbergii* (De Man, 1879). Aquaculture Journal, 3(2):133–48.

https://doi.org//10.3390/aquacj3020012.

25. Jackson S, M. Finn, and K. Scheepers. 2014. The use of replacement cost method to assess and manage the impacts of water resource development on Australian Indigenous customary economies. Journal of Environmental Management, 135:100–109. https://doi.org//10.1016/j.jenvman.2014.01.01 8.

26. Kuguru, B., J. Groeneveld, S. Singh, and B. McHomvu. 2019. First Record of Giant Freshwater Prawn *Macrobrachium rosenbergii* (De Man, 1879) from small-scale fisheries in East Africa, confirmed with DNA barcoding. BioInvasions Records, 8(2):379–91.

https://doi.org//10.3391/bir.2019.8.2.19.

27. Kusbiyanto. 2009. Bioekologi udang *Macrobrachium Spp*. di Sungai Banjaran Kabupaten Banyumas. Biosfera, 26(1):23–29.

28. Letsch, H., M. Balke, E.F.A. Toussaint, R.P. Narakusumo, K. Fiedler, and A. Riedel. 2020. Transgressing Wallace's Line brings hyperdiverse weevils down to earth. Ecography, 43(9):1329–40.

https://10.1111/ecog.05128

29. Lindenfelser, M. Ellen. 1984. Morphometric and allozyme congruence: evolution in the prawn *Macrobrachium rosenbergii* (Decapoda: Palaemonidae). society of systematic biologists morphometric, 33(2):195–204.

30. Liu, M., Y. Cai, and C. Tzeng. 2007. Molecular systematics of the freshwater prawn Genus Macrobrachium Bate, 1868 (Crustacea: Decapoda: Palaemonidae) inferred from MtDNA sequences, with emphasis on East Asian species. Zoological Studies, 46(3):272– 89.

31. Makarim, S., J. Sprinta, Z. Liu, W. Yu, A. Sant, X. Yan, and R. Dwi Susanto. 2019. Previously unidentified indonesian throughflow pathways and freshening in the Indian ocean during recent decades. Scientific Reports, 9(May):1–13.

https://doi.org//10.1038/s41598-019-43841-z.

32. Mohammed, K.S., and Y. M. S. Al-Barzinji. 2022. Genetic characterization of body weight traits in rocky partridge chicks. Iraqi Journal of Agricultural Sciences, 53(6):1289–1297.

https://doi.org//10.36103/ijas.v53i6.1643.

33. Mohammed, A.M. and M. S. ,Al-Khshali. 2023. Effect of fertilization on growth characteristics of cyprinus carpio cultured in rice fields in iraq. Iraqi Journal of Agricultural Sciences, 54(2):447–454.

https://doi.org/10.36103/ijas.v54i2.1719

34. New, M. Bernard. 2010. History and global status of freshwater prawn farming.

35. Ng, P.K.L., and D. Wowor. 2011. On the nomenclature of the palaemonid names *Palaemon spinipes* desmarest, 1817, *Palaemon spinipes* Schenkel, 1902, and *Macrobrachium wallacei* Wowor & Ng, 2008 (Crustacea: Decapoda: Caridea). Zootaxa, 68(2904):66– 68. https://doi.org//10.11646/zootaxa.2904.1.3. 36. Novak, P.A., P. Bayliss, E.A. Garcia, B.J. Pusey, and M.M. Douglas. 2017. Ontogenetic shifts in habitat use during the dry season by an amphidromous shrimp in a tropical Lowland River. Marine and Freshwater Research, 68(2):2275–88.

37. Novak, P.A., M.M. Douglas, E.A. Garcia, P. Bayliss, and B.J. Pusey. 2015. A Lifehistory account of *Macrobrachium spinipes* (Schenkel, 1902) (Cherabin) in a large tropical Australian River. freshwater science, 34(2):620–33.

https://doi.org//10.1086/681526.

38. Novak, P.A., E.A. Garcia, B.J. Pusey, and M.M. Douglas. 2016. Beesley. marine and freshwater research, 4(2):1–13.

39. Noori, A.A., and M. S. AlKhshali , J. Abuelheni. 2023. Effects of dietary addition of natural and nano zinc oxide to the diet on the physiological state of common carp. IOP

Conference Series: Earth and Environmental Science, 1252(1), 012126

40. Pascual, M., B. Rives, C. Schunter, and E. MaCpherson. 2017. Impact of life history traits on gene flow: a multispecies systematic review across oceanographic barriers in the Mediterranean Sea. PLoS ONE, 12(5):1–20. https://doi.org//10.1371/journal.pone.0176419.

41. Pickering, T., G. Pama, A. Singh, J. Varawa, R. Mana, and R. Totome. 2014. SPC activities creating an aquaculture industry for a new type of freshwater prawn in Papua New Guinea hatchery technique. 9–10.

42. Rao, T., G. Leishe, P. Candice, and M.J. Peter. 2020. Discovery of populations endemic to a marine biogeographical transition zone. Biodiversity Research, 26(June):1825–1832. https://doi.org//10.1111/ddi.13162.

43. Rowe, K.C., A.S. Achmadi, P.H. Fabre, J.J. Schenk, S.J. Steppan, and J.A. Esselstyn. 2019. Oceanic islands of wallacea as a source for dispersal and diversification of murine rodents. Journal of Biogeography, 46(12):2752–2768. https://10.1111/jbi.13720

44. Saengphan, N., B. Panijpan, S. Senapin, P. Laosinchai, A. Suksomnit, and K. Phiwsaiya. 2021. Morphology and molecular phylogeny of *Macrobrachium prachuapense Sp.* Nov. (Decapoda: Palaemonidae) from Southern Thailand. Zootaxa, 4966(4):428–42. https://doi.org//10.11646/zootaxa.4966.4.2.

45. Sanvicente-a, L., J. Zavala-hidalgo, E. Allende-arandı, and M. Hermoso-salazar. 2018. Larval dispersal in three coral reef decapod species : Influence of Larval duration on the metapopulation structure. Plos One, 13(3):1–22.

46. Sartimbul, A., H. Nakata, E.Y. Herawati, E. Rohadi, D. Yona, L.I. Harlyan, A.D.R. Putri, V.A. Winata, R.I. Khasanah, Z. Arifin, R.D. Susanto, and F.M. Lauro. 2023. Monsoonal variation and its impact on the feeding habit of Bali Sardinella (S. lemuru Bleeker, 1853) in Bali Strait. Deep-Sea Research Part II: Topical Studies in Oceanography, 211(August 2022):105317. https://doi.org//10.1016/j.dsr2.2023.105317.

47. Sastranegara, M.H., Kusbiyanto, and A.E. Pulungsari. 2020. Species richness and longitudinal distribution of crustaceans in the Logawa River, Banyumas, Indonesia. Biodiversitas, 21(11):5322–30. https://doi.org//10.13057/biodiv/d211137.

48. Shiao, J.C, C.S. Tzeng, P.C. Li, and K.N.I. Bell. 2015. Upstream migration and marine early life history of amphidromous gobies inferred from otolith increments and microchemistry. Environmental Biology of Fishes, 98(3):933–50.

https://doi.org//10.1007/s10641-014-0329-5.

49. Short, J.W. 2004. A revision of Australian River Prawns, Macrobrachium (Crustacea: Decapoda: Palaemonidae). Hydrobiologia, 525(1–3):1–100.

https://doi.org//10.1023/B:HYDR.0000038871 .50730.95.

50. Shy, J., D. Wowor, and Peter K.L. Ng. 2013. A new record of the giant freshwater prawn, *Macrobrachium spinipes* (Schenkel, 1902) (Crustacea: Decapoda: Palaemonidae) from Taiwan, with Notes on Its Taxonomy. Zootaxa, 3734(1):1925–28.

51. Siregar, A.S., T.P. Sinaga, and Setijanto. 2001. Studi ekologi fauna benthik (*Macrobrachium Spp*) pada Sungai Banjaran, Pelus dan Logawa di Kabupaten Banyumas. Biosfera, 3(Mei):1–11.

52. Suryaningsih, S., D. Bhagawati, S. Sukmaningrum, Sugiharto, and I.G.A.A. Ratna Puspitasari. 2020. Freshwater fish diversity in three tributary streams in Serayu Basin, Central Java, Indonesia. Biodiversitas, 21(12):5811–17.

https://doi.org//10.13057/biodiv/d211244.

53. Thorburn, D.C., H.S. Gill, and D.L. Morgan. 2014. Predator and Prey interactions of fishes of a tropical Western Australia River revealed by dietary and stable isotope analyses. Journal of the Royal Society of Western Australia, 97(1):363–87.

54. Toussaint, E.F.A., R. Tänzler, C. Rahmadi, M. Balke, and A. Riedel. 2015. Biogeography of Australasian Flightless Weevils (*Curculionidae, celeuthetini*) suggests permeability of Lydekker's and Wallace's Lines. Zoologica Scripta 44(6):632–44. https://10.1111/zsc.12127

55. Tran, D.D., and J. Weger. 2018. Barriers to implementing irrigation and drainage policies in an Giang Province, Mekong Delta, Vietnam. Irrigation and Drainage, 67:81–95. https://doi.org//10.1002/ird.2172.

56. Udayasuriyan, R., P. Saravana, C. Vadivalagan, and G. Rajkumar. 2015.

Efficiency of different COI markers in DNA barcoding of freshwater prawn species. Journal of Entomology and Zoology Studies, 3(October 2016):98–110.

57. Van Day. P., H. K. Huong, P.T.T.Truc, N.T.H. Nhi, D.T. Toan, L.P. Son, H.K. Nam, P.T.B. Nguyen, P.C.Hieu, D.X. Diep, and L.T.P. Mai. 2022. Effects of salinity and alkalinity on growth and survival of all-male giant freshwater prawn (*Macrobrachium rosenbergii* De Man, 1879) juveniles. Israeli Journal of Aquaculture – Bamidgeh, 74(4):1– 14. https://doi.org//10.46989/001c.56682.

58. Wainwright, B.J., I.S. Arlyza, and S.A. Karl. 2020a. Population genetics of the banded coral shrimp, Stenopus hispidus (Olivier, 1811), in the Indonesian archipelago. Regional Studies in Marine Science, 39:101430. https://doi.org//10.1016/j.rsma.2020.101430.

59. Wainwright, B.J., I.S. Arlyza, and S.A. Karl. 2020b. Population genetics of the banded coral shrimp, stenopus hispidus (Olivier, 1811), in the Indonesian Archipelago. Journal of Experimental Marine Biology and Ecology, 525(December 2019):151325.

https://doi.org//10.1016/j.jembe.2020.151325.

60. William B.L., and L.A. Rocha. 2015. Shifting seas: the impacts of Pleistocene Sealevel fluctuations on the evolution of tropical marine taxa. Journal of Biogeography, 42:25– 38.

61. Winarni, E.T., S.B.I. Simanjuntak, and A.

Nuryanto. 2023. DNA barcoding of crustacean larvae with two new records of *Caridina gracilipes* and *Ptychognathus altimanus* in the Western Region of Segara Anakan Lagoon in Indonesia. Biodiversitas, 24(1):341–48. https://doi.org//10.13057/biodiv/d240141.

62. Wowor, D., Y. Cai, and P.K.L. Ng. 2004. Crustacea: Decapoda, Caridea. Freshwater invertebrates of the Malaysian Region, 4(2):337–57.

63. Wowor, D., V. Muthu, R. Meier, M. Balke, Y. Cai, and P.K.L. Ng. 2009. Evolution of life history traits in asian freshwater prawns of the Genus Macrobrachium (Crustacea: Decapoda: Palaemonidae) based on multilocus molecular phylogenetic analysis. Molecular Phylogenetics and Evolution, 52(2):340–50. https://doi.org//10.1016/j.ympev.2009.01.002.

64. Wowor, D., and P.K.L. Ng. 2007. The giant freshwater prawns of the *Macrobrachium rosenbergii* species group (Crustacea: Decapoda: Caridea: Palaemonidae). Raffles Bulletin of Zoology, 55(2):321–336.

65. Wowor, D., and P.K.L. Ng. 2008. Case 3428 *Palaemon rosenbergii* De Man, 1879 (Currently Macrobrachium Rosenbergii; Crustacea, Decapoda): proposed conservation of usage by designation of a neotype . The Bulletin of Zoological Nomenclature, 65(4):288–93. doi: 10.21805/bzn.v65i4.a12.