EFFICACY OF SILVER ZEOLITE ON HEMATOLOGY AND SURVIVAL RATE IN CYPRINUS CARPIO INFECTED WITH SAPROLEGNIA SPP.

Mussab K. Abdulhameed

Researcher

S. A. Mustafa Prof.

Coll. Vet. Med., University of Baghdad

Correspondence: mosa b8@yahoo.com

ABSTRACT

This study was undertaken to evaluate the effect of silver zeolite (SZ) on hematological parameters and survival rate in common carp, Cyprinus carpio infected with Saprolegnia spp. A total of 120 fish (weight 100 g) were divided into six treated groups in duplicate (10 fish/tank) as follows: T1, T2, T3 and T4 that infected with Saprolegnia spp. (2×10^4) zoospore/ml) and treated with SZ at (100,200 and 300 mg/l) while T4 infected with Saprolegnia spp. and treated with copper sulfate (5g/100 L); C+ group, fish were infected with Saprolegnia without treatment and C- served as control negative group. Following 14 days from infection with Saprolegnia spp. and treatment with SZ, considerable changes were observed in blood indices. In pre-treatment, results showed a significant increased (P≤0.05) in WBC count in all infected fish (C+, T1, T2,T3 and T4) in comparison to C- group. RBC, Hb content and PCV% registered a significantly decreased (P≤0.05) in all infected fish (C+,T1, T2,T3 and T4) relative to the control group (C-). Post treatment, WBC registered significantly decreased in treated groups (T1,T2,T3 and T4) compared to C+ in WBC count. The best survival was recorded in T3 (85%) followed by T2 and T1 (65%, and 60%) respectively. In conclusion Silver zeolite seems to be a valuable disinfectant agent against Saprolegnia spp.; at the dose of 300 mg/l.

Keywords: carp- clinoptilolite- copper sulphate-fungus

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فعالية الزيولايت الفضي على المعايير الدمية ومعدل البقاء في اسماك Cyprinus carpio المصابة تجريبيا بعفن الماء مصعب كامل عبد الحميد سناء عبد العزيز مصطفى باحث

كلية الطب البيطرى، جامعة بغداد

المستخلص

هدفت هذه الدراسة إلى تقييم تأثير الزيولايت الفضي على الصفات الدمية ومعدل البقاء في اسماك الكارب الشائع المصابة تجريبيا بفطر. جريبيا بفطر (10 سمكة/حوض) على النحو التالي: 10 و 12 و 13 المصابة بـ*Saprolegnia spp* والمعالجة بالزيولايت الفضي بمعدل (200 و 300 ملغم/ لتر).) بينما كان 14 مصابًا و 12 و 13 المصابة ب*Saprolegnia spp* والمعالجة بالزيولايت الفضي بمعدل (200 و 300 ملغم/ لتر).) بينما كان 14 مصابًا و 15 همصابة ب*Saprolegnia spp* والمعالجة بالزيولايت الفضي بمعدل (200 و 300 ملغم/ لتر).) بينما كان 15 مصابًا و 10 همصابة بعد بعد يونيت النحاس (5 غم/ 10 لتر) وكذلك (+C) المصابة ب 15 مصابًا محد دريات الدم البيضاء في جميع الأسماك المصابة (+ C و 10 و 200 و 100 هالعلاج زيادة معنوية 15 هماملة السيطرة السلبية (-C). بعد 14 يومًا من الإصابة بالفطر والمعالجة بالزيولايت أظهرت النتائج قبل العلاج زيادة معنوية 16 هماملة السيطرة السلبية (-C). بعد 14 يومًا من الإصابة الفطر والمعالجة الزيولايت أظهرت النتائج قبل العلاج زيادة معنوية 17 هماملة السيطرة السلبية (-C). بعد 14 يومًا من الإصابة المصابة (+ C و 10 و 20 و 30 هر) في جميع الأسماك المصابة 10 هو 20.05 (20.05) في وعدد كريات الدم البيضاء في جميع الأسماك المصابة (+ C و 10 و 20 و 30 هر) في جميع الأسماك المصابة 10 هو 20 و 13 مقارنة بمعاملة –C. بعد العلاج ، أظهرت النتائج أن هناك انخفاضًا معنويًا في المجموعات المعالجة 10 مار 20 و 13 مقارنة بـ +C في عدد كريات الدم البيضاء. مقارنة بمعاملة السيطرة (+C). تم تسجيل أفضل معدل بقاء في 10 معرو 20 هو 60% في 12 و 11 على التوالي. بناء على هذه النتائج، أثبتت الدراسة فاعلية الزيولايت في معدل البقاء 10 معد مرض عفن الماء في اسماك الكارب الشائع بمعدل 300ملغم/لتر.

الكلمات المفتاحية: الكارب - كلينوبتيلولايت- كبريتات النحاس- فطريات

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INTRODUCTION

Around the world, aquaculture now produces 80 million tons of fish (7). In this fast growth, fish are exposed to a wide variety of diseases, notably bacteria, viruses, and fungi (17, 19). In recent years, severe fungal infections have significantly contributed to the increasing morbidity and mortality. Without a doubt, infectious diseases, especially fungi infections, are the biggest contributor to economic losses in aquaculture (14, 29,). Diseases in fish farms and aquaculture are caused by oomycetes like Saprolegniales, which include the Saprolegnia and Achlya species (21). Saprolegniasis in teleost fish is a major problem affecting both wild and farmed freshwater fish productions (2, 6). Saprolegnia spp. are secondary infections, but under certain situations, they may also operate as main pathogens, inflicting severe economic losses to aquaculture (27). Infected fish with Saprolegnia spp. often have lumps on their skin and gills that range in color from cottony white to gray, brown, red, or greenish (29). Zeolites are microporous crystalline hydrated aluminosilicates that have many applications due to their biocompatibility, such as ion exchange and adsorption- desorption properties (1). Silver is recognized to have a broad antibacterial range and to be generally harmless compared to other antibacterial metals. Among various antibacterial metals, silver is known to have a wide antibacterial spectrum and is relatively safe (5). SZ may impart antibacterial and antifungal characteristics to resins or synthetic polymers combined when with zeolite containing silver ions (28). The antifungal activity of SZ has received only marginal attention and only a few studies have been published on this topic (20). Hence, this study was aimed to evaluate the efficiency of SZ on hematology and survival rate in common carp infected with Saprolegnia spp.

MATERIALS AND METHODS Experimental design

The current study was carried out in the Fish Diseases Laboratory at the College of Veterinary Medicine/University of Baghdad for one month. A total of 120 of healthy *C. carpio* (average weight 100 g) were obtained from a commercial farm from Al-Mahawil. Initially, the health status of the experimental fish was inspected, after that the fish were dipped in formalin (37%: (15ml/100L) for 30 min. or until the appearance of the stress on fish, after two weeks of acclimation for the fish were stocked in two bath with dimension of $150 \times 20 \times 40$ cm. Then, fish were randomly distributed into 12 tanks at rate of 10 fish per tank (two replicates/treatment) as follows: T1. T2, T3 and T4 that infected with Saprolegnia *spp.* $(2 \times 10^4 \text{ zoospore/ml})$ and treated with SZ at (100,200 and 300 mg/l) while T4 infected with Saprolegnia spp. and treated with copper sulfate at concentration of 5g/100 L; C+ group, fish were infected with Saprolegnia without treatment and C- served as control negative group. Each of the six treatment groups was fed the formulated diets at a daily rate of 2% body mass during the experimental study. The experimental fish were preserved under a 12 h light/dark cycle. The chemo-physical tests of the water were measured daily during the experimental period as follows: [Temperature: 20.2±0.8 °C, Dissolved oxygen: 6.1±0.3 mg/l, pH: 7.5±0.2] as the optimize condition of carp fish.

Silver- modified- clinoptilolite (zeolite) preparation

A silver modification of clinoptilolite was achieved using the ion exchange method according to the procedure described by Nikawa *et al.* (20). Breifly, 100 g of NZ was placed in 250 ml of 3% (w/v) AgNO₃ liquid at pH 50.2 (to avoid metal precipitation) and then stirred at 300 rpm for 24 hours in a dark environment (due to silver's light sensitivity) to obtain maximal silver exchange onto the zeolites. Filtration was used to extract the zeolites from the liquid, which was then rinsed with deionized water, left to dry at 60 °C and saved ready for use.

Isolation and identification of *Saprolegnia spp.:* For the isolation of *Saprolegnia spp.*, water samples were collected from the Tigris River/Baghdad, Iraq, and the biting method was used to isolate the aquatic fungus (4, 18). In order to get pure colonies from the environment, 15–20ml of stream water was placed into a sterile petri plate with chloramphenicol. At that time, 5-7 sesame seeds, *Sesamum indicum* were placed in each petri dish (3). The plates were kept at 20 °C for 7 days, and every 24 hours, hyphae were looked. The macroscopic appearance included color, shape and other characteristic features of colonies of Saprolegnia spp., which were determined on Sabroud dextrose agar (SDA) and on sesame seeds. Direct microscopic examination included the removal of infected areas of samples and washed 2-3 times with sterile dH₂O and then transmitted to another clean slide and adding 1-2 drops of lactophenol Cotton Blue stain. Then, it was covered with a cover slip and left for 2 min. The examination was conducted using a light microscope under high magnification (x100) to determine the shape of hyphae. The spore suspension was scored in 1 ml of solution by using a "Newbaurhemocytometer chamber."

Hematological parameters

Blood samples were drawn from caudal peduncle (n=6). Then, samples were transferred to heparin anticoagulant tubes for determination of RBCs $(10^6/\mu l)$ and WBCs $(10^3/\mu l)$ count. Hemoglobin content (g/dl) was detected using Cyan-methaemoglobin method (16).

Statistical analysis

Statistical Analysis System (22) was applied to determine the impact of various study

parameters. Least Significant Difference (LSD) test (Analysis of Variance-ANOVA) were applied to compare means significantly. A P value of less than 0.05 was detected as significantly different.

RESULTS AND DISCUSSION

Isolation and identification of Saprolegnia spp.: All isolates were classified as Saprolegnia spp. depend on the morphological features of fungal colonies were appeared after 24-48 h from incubation on SDA as rounded mass of filaments, white in color and brown in the center. Also, these fungal colonies characterized by an extensive and dense mycelium (Figure 1 A). While, the fungal growth colonies in water media appeared after 3-5 days from culturing as small growth, white in color and diameter ranged 1-1.5cm (Figure 1 B). asexual reproduction of fungus isolate was distinguished by the appearance of branched non-septate hyphae as well as masses varying in width and length that are translucent and have a cell membrane. These zoosporangia were densely packed with spores (Figure 1 C and D).

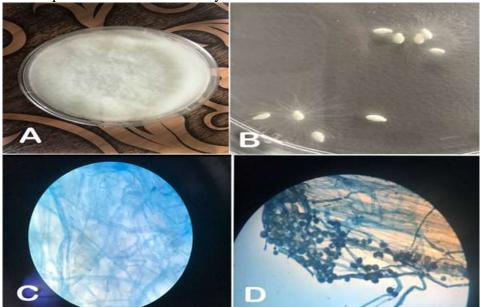


Figure 1. A- *Saprolegnia spp.* colonies on SDA at 20°C for 3-4 days started with cysts of long hairs with white cottony color. B- Wet culture of developing *Saprolegnia spp.* growth on sesame seeds. C- The wet smear showing non-septate hyphae. D- Part of sesame seed colony stained with lactophenol cotton blue showing zoospores.

Hematology: Results of WBCs count showed that at 1 and 3 days pre and post treatment with SZ , WBCs count reported significantly increased (P \leq 0.05) in C+ and in all treated groups (T1, T2, T3 and T4) relative to C-

group. While, all treated groups (T1, T2, T3 and T4) registered significantly decrease (P \leq 0.05) post treatment with SZ pre- treatment with SZ (Table 1).

	Mean ± SE of WBCs count×10 ³ /µl			
Groups	Day 1 pre treatment	Day 3 pre treatment	post treatment	LSD value
C- : healthy fish	24.56 ± 0.08	24.46 ±0.02	24.81 ± 0.01	0.043 *
	Вb	Вс	E a	
C+ infected with	30.42 ± 0.57	31.12 ±0.58	31.08 ± 0.01	1.635 NS
Saprolegnia spp.	A a	A a	A a	
T1: 100 mg\L SZ	28.84 ± 0.63	31.13 ±1.46	26.19 ± 0.02	3.182 *
_	A ab	A a	B b	
T2: 200mg/L SZ	30.21 ± 0.88	30.94 ±0.59	25.74 ± 0.01	2.116 *
0	A a	A a	Сb	
T3: 300mg/L SZ	30.04 ± 0.87	32.26 ±0.57	24.91 ± 0.01	2.092 *
_	A b	A a	Dc	
T4: CuSO ₄ 5g/100L	30.18 ±0.56	31.12 ± 0.58	24.32 ± 0.02	1.631 *
-	A a	A a	Fb	
LSD value	2.025 *	2.353 *	0.035 *	

Table 1. Results of WBCs×1	0³/μl count of <i>C. carpio</i> at 1 and 3 days pre and post treatment
	with silver zeolite

Means with different capital letters in the same column and small letters in the same row are indicated significantly different. * ($P \le 0.05$).

Results of RBCs count reported that at pretreatment (1 and 3 days) and post treatment with SZ, RBCs count stated significantly decreased (P<0.05) in C+ and in all treated groups (T1, T2, T3 and T4) in comparison to C- group. However, all treated groups (T1, T2, T3 and T4) reported significantly increased (P<0.05) post treatment compared to pre-treatment with SZ (Table 2)

Table 2. Results of RBCs×10⁶/µl count of *C. carpio* at 1 and 3 days pre and post treatment with SZ

Groups	Mean ± SE of RBCs×10 ⁶ /µl count			_ LSD value
	Day 1 pre treatment	Day 3 pre treatment	Post treatment	- LSD value
C- : healthy fish	2.87 ± 0.02	2.88 ± 0.01	2.86 ± 0.01	0.046 NS
	A a	A a	A a	
C+ infected with	1.78 ± 0.01	1.67 ± 0.01	1.57 ± 0.01	0.032 *
Saprolegnia spp. without treatment	C a	D b	Fc	
T1: 100 mg\L SZ	1.91 ± 0.02	1.73 ± 0.01	2.18 ± 0.01	0.048 *
-	Вb	Сc	E a	
T2: 200mg/L SZ	1.70 ± 0.01	1.87 ± 0.02	2.72 ± 0.01	0.034 *
-	Dс	Вb	D a	
T3: 300mg/L SZ	1.50 ± 0.01	1.67 ± 0.01	2.76 ± 0.01	0.034 *
5	Fc	Db	C a	
T4: CuSO ₄ 5g/100L	1.59 ±0.01	1.61 ±0.01	2.82 ± 0.01	0.035 *
	Еb	ЕЬ	B a	
LSD value	0.041 *	0.031 *	0.031 *	

Means with different capital letters in the same column and small letters in the same row are indicated significantly different. * (P \leq 0.05).

Hb content pre-treatment (1 and 3 days) and post treatment with SZ recorded that there were significant differences (P<0.05) in C+ and all treated groups (T1,T2, T3 and T4) compared to C- group. But, all treated groups (T1, T2, T3 and T4) recorded significantly increased in Hb content post treatment with SZ relative to these groups in pre-treatment with SZ (Table 3).

		SZ		
	Mean ± SE of Hb content (g/dl)			
Groups	Day 1 pre treatment	Day 3 pre treatment	After treatment	LSD value
C- : healthy fish	8.73 ± 0.02	8.73 ±0.02	8.73 ±0.01	0.046 NS
	A a	A a	A a	
C+ infected with	4.87 ± 0.01	4.96 ±0.01	4.98 ± 0.01	0.034 *
Saprolegnia spp.	B b	B a	C a	
T1: 100 mg\L SZ	4.54 ± 0.02	4.79 ±0.17	7.44 ± 0.33	0.753 *
_	BC b	BC b	B a	
T2: 200mg/L SZ	4.36 ±0.33	4.60 ± 0.01	8.56 ± 0.64	1.449 *
	BC b	BC b	A a	
T3: 300mg/L SZ	4.03 ± 0.22	4.44 ±0.32	8.43 ± 0.01	0.791 *
	СЬ	СЬ	A a	
T4: CuSO ₄ 5g/100L	4.28 ±0.33	4.58 ± 0.01	8.81 ± 0.14	0.726 *
	BC b	BC b	A a	
LSD value	0.653 *	0.465 *	0.934 *	

Table 3. Results of Hb content (g/dl) of C. carpio at 1 and 3 days pre and post treatment with	
SZ	

Means with different capital letters in the same column and small letters in the same row are indicated significantly different. * (P \leq 0.05).

On the other hand, results revealed that the PCV (%) pre-treatment (1 and 3 days) and post treatment with SZ there were significantly decreased (P<0.05) in C+ and all treated groups (T1,T2, T3 and T4) compared to C-

group. However, all treated groups (T1, T2, T3 and T4) verified significantly increased in PCV (%) post treatment with SZ compared to these groups in pre-treatment with SZ (Table 4).

Table 4. Results of Packed Cell Volume (%) of C. carpio at 1 and 3 days pre and post
treatment with SZ

	Mean ± SE of PCV %			
Groups	Day 1 pre treatment	Day 3 pre treatment	After treatment with SZ	LSD value
C- : healthy fish	32.70 ±0.35 A a	31.38 ±0.36 A a	32.73 ±0.01 A a	2.169 NS
C+ : infected with Saprolegnia spp. without treatment	20.46 ±0.47 B a	21.47 ±0.86 B a	21.78 ±0.86 D a	2.621 NS
T1: 100 mg\L SZ	20.70 ±0.95 B b	21.12 ±1.01 B b	27.85 ±0.63 C a	3.042 *
T2: 200mg/L SZ	21.35 ±0.49 B b	20.56 ±0.99 B b	29.75 ±0.19 BC a	2.252 *
T3: 300mg/L SZ	21.94 ±0.89 B b	20.67 ±0.95 B b	30.86 ±0.27 AB a	2.66 *
T4: CuSO ₄ 5g/100L	20.77 ±0.96 B b	21.81 ±0.32 B b	32.38 ±0.34 A a	2.142 *
LSD value	2.257 *	2.480 *	1.910 *	

Means with different capital letters in the same column and small letters in the same row are indicated significantly different. * ($P \le 0.05$).

WBCs are involved in the control of immune activities, and as a protective reaction to stress, their quantity increases in fish. High WBCs counts suggest tissue damage caused by infection, extreme physical stress, and leukemia. Thus, in the current investigation, an increase in WBCs may have been produced by tissue damage-induced immune system activation (23). Results of the current study in pretreatment (1 and 3 days) showed a significantly increased ($P \le 0.05$) in the WBC count of *C. carpio* in C+ compared to C-. These results are in agreement with Jamalzadeh *et al.* (11) who observed that the WBC was higher values in fungal infected fishes than healthy Caspian Salmon, *Salmo ciscaucasicus*. In the present work, the increases in WBC count in infected *C. carpio* (C+) and treated groups (T1, T2, T3, and T4) may be a result of the cellular immune system's reaction to fungal infection. Similar observation in rainbow trout were also

documented as in the current research (24). The rise in WBCs is connected with a stimulation of the immune response and an increase in antibody production, which aids in the survival and recovery of infected fish 26). Following treatment, there was a significantly decreased of WBCs count in treated groups (T1, T2, T3, and T4) when compared to C+. These results are correlated with the results on culture media, which showed inhibition for fungal growth on SDA, and with the clinical signs that appeared after treatment in all treatment groups which showed the fungal hyphae disappeared from the surface of skin, healing all injuries from the infected area, the color of skin returning to normal, stress reduction, and the fish returning to normal behaviors (data not shown). Infected groups, C+ and treated groups (T1, T2,T3, and T4) exhibited substantial reductions in RBC count, PCV value, and Hb content compared to uninfected group (C-) prior to treatment. This decline may be related to anemia induced by hyphae that invade the blood vessels of infected fish and cause bleeding, damage to hematopoietic tissues, haemodilution with osmotic imbalance, and lethargy owing to mucus secretion, these outcomes are consistent with Shah and Altindag (25). Accelerated erythroclasis (i.e., red blood cell fragmentation) as a result of altered membrane permeability and/or enhanced mechanical fragility. These results are consistent with Suhail et al. (26) and Gill and Epple (8). Other explanation. This could be because Saprolegnia mycelia penetrate deeply, causing wounds that result in blood loss (13). Our findings are also in agreement with Zaki (30) whom reported that Tilapia nilotica infected with Saprolegnia parasitica resulted significantly decreased in RBCs count, Hb content and PCV. Post-treatment, results of the current study showed significant increases in RBC count, PCV and Hb in all treatment groups compared to the infected group C+. These results are correlated with the results on culture media, which showed inhibition for fungal growth on SDA, and with the clinical signs that appeared after treatment in all treatment groups T1, T2, T3 and T4, which showed the fungal hyphae disappeared from the surface of skin, healing all injuries from the infected area, the color of skin returning to normal, stress reduction, and the fish returning normal behaviors. Two antifungal to mechanisms are proposed to explain the behavior of AgNP-coated zeolite: first, the fungi are directly killed by the silver ions released from the filters (fungicidal effect); second, by passing through the AgNP-coated zeolite, the fungi are contaminated with silver ions, but still survive; however, they cannot form colonies on the surface of the fish, as the silver ions inhibit their replication and growth ability (fungistatic effect) (12, 15).

Survival rate

Survival registered highest rate in T4 (90%) followed by T3 (85%) then T1 and T2 (65 and 60%) respectively and the survival percentage in C+ was 50%. The highest rate of survival particularly T3 compared to infected group (C+) suggests that SZ was very efficient disinfectant, can inhibit their replication and growth ability of Saprolegnia (fungistatic effect) (12). Similar results reported by Hamad et al. (9) who reported that the ability of ozone to reduce Saprolegnia infection in common carp. Our results are also in consistent with Hlial et al. (10) whom demonstrated the best antifungal activity of curcumin-silver nanoparticles against Saprolegniasis in common carp.

CONCLUSIONS

In conclusion Silver zeolite seems to be a valuable disinfectant agent against *Saprolegnia spp.;* at the dose of 300 mg/l. The results of this study demonstrate the first success using SZ for the treatment of *Saprolegnia spp.* in common carp.

REFERENCES

1. Abdulathem, A. A. and A. J. Al-Rudainy. 2021. Effect of dietary zeolite on ammonia toxicity and on some of blood parameters in common carp *Cyprinus carpio*. Iraqi Journal of Agricultural Sciences. 52(5): 1276–1283. https://doi.org/10.36103/ijas.v52i5.1465

2. Al-Hassani, T.S and S.A. Mustafa. 2022. Efficiency of synbiotic as feed additives on growth performance, survival rate and health status in common carp challenged with *Saprolegnia spp*. Iraqi Journal of Agricultural Sciences. 53(2): 397-405.

https://doi.org/10.36103/ijas.v53i2.1548

3. Al-Rudainy, A.J., S.A. Mustafa, A.A. Ashor and M.T. Bader. 2023. The role of kaolin on hematological, biochemical and survival rate of Cyprinus carpio challenged with Pseudomonas aeruginosa. Iraqi Journal of Agricultural Sciences. 54(2): 472-477.

https://doi.org/10.36103/ijas.v54i2.1723

4. Ashour, A. A., S. A. Mustafa and S. N. Yassein. 2017. Histopathological studies on common carp (Cyprinus carpio L.) infected with Saprolegnia spp. and treated with Virkon®. Mirror Res. Vet. Sci. Anim. 6(1): 19 - 30

A. A., N. M. Salman, S. A. 5. Ashour, Mustafa and R. O. Nemah. 2019. Evaluation hydrogen peroxide on controlling of saprolegniasis in common carp, Cyprinus carpio L. Biochem. Cell. Arch. 19(2): 4247-4252

6. Fadhal, A. A. and S. A Mustafa. 2020. Influence of phytase enzyme on growth performance and survival rate challenged with Saprolegnia spp. in common carp. Iraqi Journal of Agricultural Sciences. 51(5): 1458-1465. https://doi.org/10.36103/ijas.v51i5.1156 7. Faik, H. H. and S.A. Mustafa. 2023. Role of and Ε vitamin С on genotoxicity, hematological and biochemical investigation in Cyprinus carpio following zinc oxide nanoparticals exposure. Iraqi Journal of Agricultural 54(3): 716-723. Sciences. https://doi.org/10.36103/ijas.v54i3.1752

8. Gill, T. S. and A. Epple. 1993. Stressrelated changes in the hematological profile of American eel (Anguilla rostrata). the Ecotoxicol. Environ. Saf. 25(2): 227-235

9. Hamad, S.M. S A. Mustafa and A.M. Kane. 2019. Evaluation The Ozone Treatment to Control the Infection of Saprolegniasis in Cyprinus carpio L. J. Phys.: Conf. Ser. 1294.

10. Hlial, S.M. S. A. Mustafa and E.E. Obodi. 2021. Antifungal activity of curcumin- silver nanoparticles against Saprolegnia spp. in common carp. Plant Ach. 21(1): 148-154.

11. Jamalzadeh, H. R., A. Keyvan, M. R. Ghomi and F. Gherardi. 2009. Comparison of blood indices in healthy and fungal infected Caspian salmon (Salmo trutta caspius). Afr. J. Biotechnol. 8(2): 319-322

12. Johari, S. A., M. R. Kalbassi, M. Soltani, and I. J. Yu. 2016. Application of nanosilvercoated zeolite as water filter media for fungal

disinfection of rainbow trout (Oncorhynchus mykiss) eggs. Aquac. Int. 24(1): 23-38

13.Juncey, K., and B. Ross. 1982. A Guide to Tilapia Feed and Feeding. Institute of aquaculture University of Striling. Scotland. 125 P

14. Madrid, A., P. Godoy, S. González, L. Zaror, A. Moller, E. Werner, M. Cuellar, J. Villena and I. Montenegro. 2015. Chemical characterization and anti-oomycete activity of Laureliopsis philippianna essential oils against Saprolegnia parasitica and S. australis. Molecules. 20(5): 8033-8047

15. McLeay, D. J. 1973. Effects of a 12-hr and 25-day exposure to kraft pulp mill effluent on the blood and tissues of juvenile coho salmon kisutch). J. Fish. Res. B. (Oncorhynchus Can.30(3): 395–400.

16. Mustafa, S.A. 2012. An Integrated Approach to Assess Impact of Environmental in Carp, Cyprinus Stress carpio L.: Biochemical, Genotoxic, Histopathological and Individual Level Effects. Ph.D. Dissertation.. Dept. of Biomed. and Biol. Sci.. Fac. of Sci.. Uniu. Plymouth. 77 P

17. Mustafa, S.A. and A.A. Ashor. 2021. Effect of chronic exposure to silver nanoparticles on histopathological manifestations of common carp, Cyprinus carpio L. Biochem. Cell. Arch. 21(1): 2719-2725

18. Mustafa, S.A., A.J. Al-Rudainy and K.A. AL-Faragi. 2019. Assessment of hydrogen peroxide on histology and survival rate in common carp, Cyprinus carpio L. infected saprolegniasis. with Iraqi Journal of Agricultural Sciences. 50(2):697-704.

ttps://doi.org/10.36103/ijas.v2i50.669

19. Mustafa, S. A. and A. N. Jha. 2022. Impact of Hypoxia and Dead Zones on Habitat Destruction and Fish Population: In: K.K. Jha and M. Campbell (Eds) Dynamics and Interrelations Between Nature, Science, and Society, New York, p. 158-160.

20. Nikawa, H., T. Yamamoto, T. Hamada, M. B. Rahardjo, H. Murata and S. Nakanoda. 1997. Antifungal effect of zeolite incorporated tissue conditioner against Candida albicans growth and/or acid production. J. Oral Rehabil. 24(5): 350-357

21. Okumus, İ. 2002. Rainbow trout broodstock management and seed production in Turkey: Present practices, constraints and the future. Turkish J. Fish. Aquat. Sci. 2(1): 41-56

22. SAS 2018. Statistical Analysis System, User's Guide. Statistical. Version 9', SAS. Institute Inc. USA

23. Seth, N. and K. K. Saxena. 2003. Hematological responses in a freshwater fish *Channa punctatus* due to fenvalerate. Bull. Environ. Contam. Toxicol B. 71(6): 1192–1199

24. Shah, A. F., A. S. Bhat, F. A. Bhat, M. H. Balkhi, A. Abubakr and I. Ahmad. 2015. Alteration in haemato-biochemical profiles of rainbow trout *Oncorhynchus mykiss* affected by *Saprolegnia spp-A* potential constraint for culture of trout in Kashmir Himalaya. Iran. J. Fish Sci.14(4): 970-984

25. Shah, S. L. and A. Altindag 2004. Hematological parameters of tench (*Tinca tinca L.*) after acute and chronic exposure to lethal and sublethal mercury treatments. Bull. Environ. Contam. Toxicol. B. 73(5): 911–918 26. Suhail, R. N., S.A. Mustafa, and E. E. AL-

Obodi. 2022. Effeciency of silver nanoparticales as antibacterial against

Aeromonas hydrophila isolated from infected common carp. Iraqi Journal of Agricultural Sciences. 53(3), 589-597.

https://doi.org/10.36103/ijas.v53i3.1568

27. Trusch, F., L. Loebach, S. Wawra, , E. Durward, A. Wuensch, N. A. Iberahim, , I. De Bruijn, K. MacKenzie, A. Willems and A. Toloczko. 2018. Cell entry of a host-targeting protein of oomycetes requires gp96. Nat. Commun. 9(1): 1–12

28. Uchida, T., N. Maru, M. Furuhata, A. Fujino, S. Muramoto, A. Ishibashi, K. Koshiba, T. Shiba and T. Kikuchi. 1992. Antibacterial zeolite balloon catheter and its potential for urinary tract infection control. Hinyokika Kiyo. Acta Urologica Japonica. 38(8): 973–978

29. Yanong, R. P. 2003. Fungal Diseases in Fish. Black Sea J. Agric. 7–8

30. Zaki, M. S., O. M. Fawzi and J. E. Jackey 2008. Pathological and biochemical studies in *Tilapia nilotica* infected with *Saprolegnia parasitica* and treated with potassium permanganate. J. Agric. Environ Sci. 3(5): 677–680.