STUDYING THE EFFICIENCY OF SOME NANOPARTICLES ON SOME PLANT PATHOGENIC FUNGI AND THEIR EFFECTS ON HYPHAL MORPHOLOGY

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ABSTRACT

This study was investigated the effect of iron NPs and essential oils of clove in normal and nanoemulsion forms for controlling *Fusarium oxysporum, Fusarium solani*, and *Alternaria solani* compared with Azoxystrobin fungicide under laboratory conditions. Data revealed that all tested compounds are capable of inhibiting the growth of mycelial of *F. oxysporum F. solani*, and *A. solani* from 0 to 84.4%, 0 to 88.9 %, and 0 to 61.1 %, respectively. There is relationship was found between the tested concentration of all treatments and their percentages of inhibition of mycelium. In addition, the use of Azoxystrobin, iron NPs and essential oils of clove in normal and nanoemulsion increased the activity of polyphenol oxidase (PPO). The fungal hyphae morphology was investigated by using scanning electron microscopy (SEM). The fungi hyphae without treatments are regular branching, linearly and normal morphology shaped, the surface of the hyphae is smooth and apical tapered. Treatments caused loss of linearity, irregular branching of the terminal hyphae, deformations of the hyphal shape, and the lysis cytoplasm of the hyphal.

Keywords: Fusarium oxysporum, Fusarium solani, Alternaria solani. Azoxystrobin, iron NPs, clove oil, clove oil NPs, PPO, SEM.

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لنباتية وتأثيرها على الشكل المظهري للهيفات.	, بعض الفطريات المسببة للأمراض اا	دراسة كفاءة بعض الجسيمات النانوية على
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قسم وقاية النبات، كلية الزراعة، جامعة دمنهور

المستخلص:

تهدف الدراسة الى معرفة تاثير كلا من زيت القرنفل فى صورتة الطبيعية والنانوية والحديد النانوي والمبيد الفطري ازوكسيستروبين على فيوزاريوم أوكسيسبوروم و فيوزاريوم سولانى و التيرناريا سولانى فى المعمل. اظهرت النتائج ان جميع المركبات المختبرة قادرة على تثبيط الفطريات المختبرة. وكلما زاد تركيز المركبات المختبرة ادي الى زيادة نسبة التثبيط مقارنة بالكنترول. ومن خلال البيانات المتحصل عليها وجد أن فيوزاريوم سولانى هو الأكثر حساسية ، بينما أقلهم حساسية هو التيرناريا سولانى لجميع المركبات المختبرة. كما وجد أن ميد ازوكسيستروبين أعطى أفضل النتائج ضد جميع الفطريات المختبرة. وايضا ادت المعاملات المختبرة. كما وجد أن مبيد ازوكسيستروبين أعطى أفضل النتائج ضد جميع الفطريات المختبرة. وايضا ادت المعاملات الى زيادة نشاط انزيم البولي فينول اوكسيديز مقارنة بالكنترول. كما اظهر استخدام المكرسكوب الماسح للالكترونات ان هيفات الفطريات فى الكنترول ذات تفرعات منتظمة وشكلها طبيعى وملساء بينما هيفات الفطريات بعد المعاملة اصبحت ذات تفرعات غير منتظمة وادى الى حدوث تشوهات وتحلل الستيتوبلازم للهيفات الفطريات. الكلمات المفتاحية: فيوزاريوم أوكسيسبوروم وفيوزاريوم سولانى والتيرناريا سولانى، الماستحد النانوي، زيت الفطريات بعد المعاملة الى زيادة الفطريات فى الكنترول ذات تفرعات منتظمة وشكلها طبيعى وملساء بينما هيفات الفطريات العاملية الميتوبلازم المحترة الفريات إلى منتظمة وادى الى حدوث تشوهات وتحلل الستيتوبلازم للهيفات الفطريات. الفطريات المفتاحية: فيوزاريوم أوكسيسبوروم وفيوزاريوم سولانى والتيرناريا سولانى، ازوكسيستروبين، الحديد النانوي، زيت

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INTRODUCTION

Due to the harmful effects the synthetic fungicides on the environment and humans. In addition to targeting pathogens, pesticides could kill various beneficial organisms, and their toxic forms can persist in soil (13) and their frequent use has led to the emergence of resistant strains. Additionally, in recent years public pressure to reduce the use of synthetic fungicides in agriculture has increased (2). The world begins to use plant-derived products as disease control agents as they had fewer environmental effects and low mammalian toxicity (7) and (6). Essential oils have a long history of use as natural microbial agents and are used in several foods, pharmaceuticals, and cosmetic products. Essential oils inhibited the growth of a wide range of microorganisms, with fewer side effects than synthetic fungicides (28). Clove (Syzygium aromaticum L) used for centuries for medicinal purposes or food preservatives. Flower buds had many medicinal properties as an antifungal. antimicrobial. carminative. antiviral. hypertensive aphrodisiac, light stomachic, and anesthetic (18) and (21). Clove essential oil had the antifungal properties to exhibit strong inhibitory effects against the mycelial growth of many fungi and is widely investigated due to its availability, popularity, and high essential oil content (27) and (22). Nowadays nano-biotechnology has emerged as one of the fastest-growing areas of research in different technology and science (35). The nanocapsules, prepared interfacial by deposition of the preformed polymer (nanoprecipitation), represent an effective method to obtain robust nanosystems suitable for applications in various fields, ranging from medicine, health, and agri-food to the environment (9). Nanoparticles have high against efficiency plant pathogens viz. nematodes, bacteria, viruses, and fungi, and easy application to soil, foliage, and seeds (17). The new trend of nanotechnology is safe eco-friendly antifungal agents such as plantbased oils. Essential oils (EOs) are lipophilic compounds, easily degradable by the effects of oxygen, light, moisture, and, temperature. Nanoencapsulation is a valid strategy to overcome these obstacles. This technology allows for the protection the essential oils from thermal and photodegradation phenomena, solubility increasing their in aqueous environments, masking their flavor, and bioaccessibility improving their and bioavailability (12). Nanoemulsion oil of Eugenol showed fungicidal activity against F. oxysporum (3). Nanoemulsion essential oil of citronella had high efficacy against S. rolfsii and R. solani (5). Nanoemulsions of clove, orange black seed, and lemon oils significantly decrease the mycelial growth of F. solani, A. tenuissima, and G. candidum. Nanoemulsion (25). The present study aimed to evaluate the antifungal activities of clove oil, clove oil (NPs), Fe (NPs), and Azoxystrobin fungicide controlling Fusarium oxysporum, for Fusarium solani, and Alternaria solani, as well as the determination PPO activities, and their effects on F. oxysporum, F. solani, and R. solani hyphae by using scan electronic microscope.

MATERIALS AND METHODS

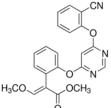
1. Treatments

Common name: Azoxystrobin :**Trade name**: Amistar 25 %SC

Chemical formula: C₂₂H₁₇N₃O₅

Chemical name: Methyl (E)-2-[2-(6-(2cyanophenoxy) pyrimidin-4-yloxy) phenyl] =-3methoxyacrylate

Chemical structure



Clove oil: obtained from International Favors and Plant oils Inc., Giza., Egypt.

Clove oil (NPs): nanoemulsion for natural oil synthesized at College of Biotechnology, Misr University for Science and Technology, Egypt **Iron (NPs):** obtained from the Agriculture Research Center of Kafr Eel-Sheikh

2. Fungi strains used

Fusarium solani (*F. solani*): Obtained from the Department of Plant Diseases, Faculty of Agriculture, Damanhur University

Alternaria solani (*A. solani*): obtained from the Department of Plant Pathology, Agricultural Research Station, Itay El Baroud

oxysporum (*F*. oxysporum): Fusarium brought from Plant Institute, Pathology Agricultural Research Center

2.1. In vitro antifungal activity assay

Used to essential clove oil at concentrations of 200 and 400 μ g/ml, clove oil nanoemulsion (NPs) at concentrations of 50 and 100 μ g/ml, iron NPs at concentrations of 50 and 100 μ g/ml, and fungicide Azoxystrobin at 250 μ g/ml were tested in vitro against F. oxysporum, F. solani, and A. solani by Poisoned Food Technique. Added separately to get the required concentrations then mixed with 50ml of sterilized PDA medium and transferred equally into three Petri dishes (9 cm). The media allowed solidifying. Control without tested compounds. These plates were inoculated with an added disc 5cm diameter of the culture of the test fungi. All dishes were incubated at 27±2°C for 7 days and radial growth of the colony was measured when the mycelia of control were complete with the colonization of the fungi. Each treatment was in triplicate. performed The inhibition percentages were calculated according to the method of (33) by using the formula: $MGI\% = \frac{R-r}{R} \times 100$ MGI%: mycelium growth

inhibition percentage

R: the growth of mycelium of fungi in the control plate

r: the growth of mycelium of fungi in the treatment plate

3. Determination of polyphenol oxidase activity (PPO).

The activity of polyphenol oxidase was assayed according to the method of (10) using a spectrophotometer as follows: The sample of each fungus was homogenized in 0.1 M phosphate buffer (pH 7.0) and centrifuged at 4000 rpm for 10 min at 4°C. The final mixture contained 1 mL enzyme, 2 mL 0.1 M phosphate buffer (pH 7.0), and 1 mL 0.1 M methyl catechol, and an increase in the absorbance was measured at 495 nm. The activity of PPO was expressed as units'/mg tissue.

4. Effects of different antifungal treatments on hyphae morphology: The changes in the morphology of fungal hyphae using different antifungal treatments were characterized by scanning electron microscopic (SEM). The SEM observation (JOEL, JSM 5300) with high resolution at an accelerating voltage of 120 Kev. An aliquot of each material was coated with a copper grid and scanned for its shape and size. The tested cells were subjected to Xray Electron Dispersive Analysis (EDA) using an X-ray Oxford detector unit (model 6697, England)- Faculty of Science, Alexandria University.

5. Statically analysis

Data were analyzed using ANOVA. Analysis of variance (ANOVA) using software 1998 -2005 cost at 6.3111 and Duncan, multiple range test at level P < 0.05 used method of Winer 1971.

RESULTS AND DISCUSSION

Comparing obtained data using Essential oil (EOs) of clove in normal and nanoemulsion, iron NPs and Azoxystrobin fungicide at used concentrations as μ g/ml in Table (1). Through the data obtained, it was found that F. solani is the most sensitive, while the least sensitive is A. solani to all tested compounds. It also found that the fungicide Azoxystrobin gave the best results against all tested fungi. The highest level of inhibition was obtained using the fungicide Azoxystrobin, while the lowest level of inhibition was by iron NPs at 50 μ g/ml. The growth of mycelium decreased with increasing the concentrations in all treatments. Data revealed that all tested were compounds capable of inhibiting the growth of mycelial of F. oxysporum F. solani, and A. solani from 0 to 84.4%, 0 to 88.9 %, and 0 to 61.1 %, respectively. The inhibition growth of mycelium increased with increasing the concentrations in all treatments. These results agree with (41) who found that the antifungal activities of Azoxystrobin fungicide and clove were able to inhibition of spore oil germination of the fusarium isolated. In addition, Azoxystrobin and Sarfun 500 µg/ml were regarded as a prospective means of limiting the development and inhibiting mycelium growth of Colletotrichum gloeosporioides and protecting Hypericum perforatum L. (11). Also, essential clove oil had stronger antifungal activities against Penicillium citrinum and Aspergillus flavus than Rhizopus nigricans (39). The previous research showed that microbialsynthesized iron nanoparticles by Fusarium

oxysporum exhibited antimicrobial activity (32) and ZnO NPs enhanced the antimicrobial effect (30). The activity of Azoxystrobin fungicide compared with essential oils extracted from Eberm linaloolifera, Eucalyptus staigeriana, Cinnamomum camphora, and Eucalyptus globulus L. against Alternaria solani and all tested compounds controlled early blight disease in vitro assays (36). Essential oil clove had antifungal activity and strongly inhibited the mycelial growth of fungi (8). Essential oils play an important role in controlling plant diseases and pests. Nanoemulsion containing lemongrass and clove oils had antifungal agents against F. oxysporum and was rapidly affected as fungicidal (29). Many authors explained the antimicrobial activity of plant oils due to

passing oils through the cell wall and membrane cytoplasmic of the fungus depending on partition and hydrophobicity in the plasma membrane. The clove oil exhibited inhibitory activity against the mycelial growth of F. oxysporum and R. solani. The concentrations used significantly reduced the mycelial growth compared with the control (34). The biosynthesized iron nanoparticles exhibited inhibitory activity against different microorganisms (22). The efficiency of nanoemulsions of clove, orange black seed, and lemon oils against F. solani, A. tenuissima and G. candidum. Nanoemulsion of clove and black seed oils showed a significant decrease in the mycelial growth of fungal isolates at 5000 ppm (25)

Table 1. Effects of Azoxystrobin, iron NPs and essential oil of clove in normal andnanoemulsion forms on mycelial growth (cm) and inhibition (%) of F. oxysporum, F. solani,and A solani after 7 days of incubation at 27+2 °C

Treatments	Мус	Mycelial growth (cm)		Inhibition (%)		
(µg/ml)	F.o	F.s	A.s	F.o	F.s	A.s
*Control	9.0a	9.0a	9.0a	0.0	0.0	0.0
Clove oil 200	4.6d-f	3.7f-i	5.9c	48.9j	58.9gh	34.4 l
Clove oil 400	3.9e-h	3.1g-k	5.3cd	56.7hi	65.6e	41.1k
Clove oil (NPs) 50	4.1d-g	2.6h-k	5.1с-е	54.4i	71.1d	43.3k
Clove oil (NPs) 100	3.3f-j	2.2i-k	4.6d-f	63.3ef	75.6c	48.9j
Fe (NPs) 50	6.1 bc	4.1d-g	7.1b	32.21	54.4i	21.1m
Fe (NPs) 100	5.3cd	3.5f-j	8.1a	41.1k	61.1fg	10.0m
Azoxystrobin 250	1.4jk	1.0k	3.5f-j	84.4b	88.9a	61.1fg

F.o: F. oxysporum, F. s: F. solani, and A. s: A. solani

Each number represents the mean of 3 replicates.

*Control without active ingredient (medium free and discs were cut from the pathogen only on PDA).

Different letters in Table (1) indicated significant differences according to Duncan's multiple range test at the level $P \le 0.05$

Determination of polyphenol oxidase enzyme (PPO).

The efficiency of Azoxystrobin, iron NPs and clove oils in normal and nanoemulsion on polyphenol oxidase enzyme showed in Figure (1). Treatment with Azoxystrobin fungicide and nanoparticles led to an increase in polyphenol oxidase enzyme in all tested fungi. The highest increase was when using the fungicide Azoxystrobin, while the lowest was when using Fe NPs at 50 µg/ml compared to the control. The results also showed an increase in polyphenol oxidase enzyme in A. solani, then followed by F. oxysporum, and the least of them F. solani. In addition, the increase in concentrations in all treatments increased the polyphenol oxidase enzyme. The more recent research on polyphenol oxidase reviewed fungi and plants and the main aspects considered properties, structure. location, distribution, and discovered inhibitors (PPO). Most fungi and plants have multiple forms of PPO and their function remains enigmatic. The function of PPO is probably different from that in plants and PPO plays an important role in defense against other pathogens (23). The activity of PPO in response to treatment with compounds indirectly helped in cell wall reinforcement to prevent the penetration and dissemination of compounds within the cell wall of fungi (19). The mechanism of definition against microbial pathogens initiates with the induction of defense enzymes such as PPO, deposition of the cell wall, and hypersensitive response (26). CuO NPs and MgO NPs had a great impact on increasing the enzyme activities of PPO (Ismail 2021). Treated with ZnO NPs increased the activities of PPO enzyme defense (16). The activity of polyphenol

oxidase increased when treated with oil of Vitex agnus-castus L and zinc oxide nanoparticles compared with the untreated (4).

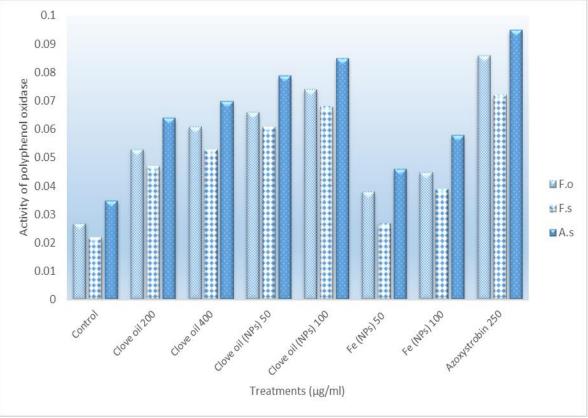


Figure 1. Effects of Azoxystrobin, iron NPs and essential oils of clove in normal and nanoemulsion forms on polyphenol oxidase enzyme (PPO) in *F. oxysporum*, *F. solani*, and *A. solani*

Effects of Azoxystrobin fungicide, iron NPs and clove oils in normal and nanoemulsion on Morphological fungi hyphae: Use the microscope scanning electron at 1500 magnification to know the effect of the Azoxystrobin fungicide, iron NPs and clove oils in normal and nanoemulsion forms on hyphae of F. oxysporum, F. solani, and A. solani. The information obtained from the SEM images referred to the effect on the hyphae of each fungus varies according to the type and concentration of each treatment. In addition, the shape of the hyphae for each fungus differs in control from the treatment. The data from obtained results by SEM revered significant morphological changes in the surface of the cell wall of each fungus. It found that the surface wall of the fungus F. solani was the highest affected, while the least affected was A. solani. The hyphae on control regular branching, linearly and normal morphology shaped, the surface of the hyphae is smooth and apical tapered. Treatments

the terminal hyphae, deformations of the hyphal shape, and lysis cytoplasm of the hyphal. The use of the pesticide Azoxystrobin caused the rupture of hyphae, irregular branching in the apical of hyphae, and the loss of linearity and morphological shape. The effect of the Azoxystrobin on the rupture and deformation of the hyphae in the fungus F. solani, while the fungus A. solani was the least deformation and rupture in its hyphae. It also found that clove oil in the nanoemulsion form caused more rupture and distortion of the hyphae than clove oil in its normal form. Iron nanoparticles cause less tearing and deformation of hyphae than Azoxystrobin and clove oil. Effects of Azoxystrobin, iron NPs and essential oils of clove in normal and nanoemulsion forms on F. oxysporum hyphae in Figure (2). Through the results obtained from the scanning electron microscope, these treatments affected the hyphae of the fungus and led to the occurrence of deformations and

caused loss of linearity, irregular branching of

ruptures, and the most obvious of them were Azoxystrobin, then followed by nanoemulsion of clove oil, clove oil, and the least effect by iron NPs. Effects of Azoxystrobin, iron NPs and essential oils of clove in normal and nanoemulsion forms on the hyphae of F. solani in Figure (3). Treatment with these compounds leads to distortions and ruptures, and these distortions in F. solani hyphae were more than what happened to the hyphae of A. solani and F. oxysporum. The occurrence of distortions may be due to its sensitivity to the tested compounds over other fungi. Effects of Azoxystrobin, iron NPs and essential oils of clove in normal and nanoemulsion forms on A. solani hyphae in Figure (4). It observed that the hyphae of A. solani fungi were the least affected, perhaps due to the least sensitivity to the tested treatments over other tested fungi. Treatment with oil of clove on the P. digitatum caused irregular branching of hyphae, loss of linearity, and effects on the morphology of hyphae (40). Cavities and pores observed on the treated surface of hyphae with AgNPs and pits may be due to the creation of pores on the

membranes of fungus. The reaction of AgNPs sulfur and phosphorous-containing with materials outside or inside of cells possible cause. In addition, positive charges of AgNPs maybe bind with the negative charge of fungal membranes destroying the lipid bilayer of the membrane, leading to inducing intracellular ion efflux resulting in the death of the cell (15). The treatment with AgNPs caused morphological changes in the cell wall surface of hyphae of A. solani compared to the control (2). The effect of clove oil on fungi morphological features showed disruption and conidial malformation as well and in the case of F. oxysporum, clove oil increased the production of chlamydospores but decreased the conidial numbers (34). AgNPs caused changes in the structure of a mycelial and deformation of the hyphal in F. solani (37). The hyphae of *R. solani* treated with ZnO NPs lost their smooth surface and formed unusual bulges on the surface of hyphae opposite of untreated hyphae with a smooth surface may be back to ZnO NPs deforming hyphae structure and inhibited R. solani growth (16).

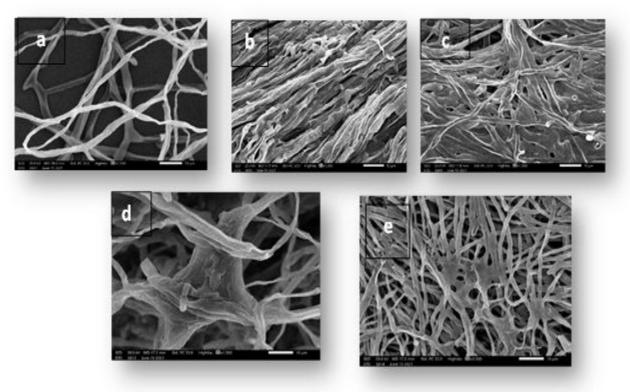


Figure 2. SEM of *F. oxysporum* hyphae after a week of incubation at 27 °C. (a) Control, (b) treated with Azoxystrobin, (c) treated with clove oil, (d) treated with clove oil nanoemulsion, (e) treated with iron NPs

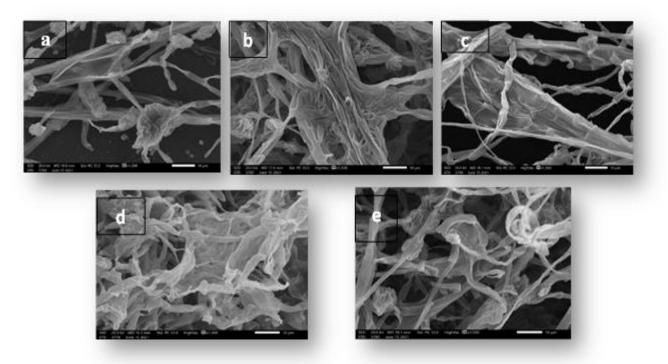


Figure 3. SEM of *F. solani* hyphae after a week of incubation at 27 °C. (a) Control, (b) treated with Azoxystrobin, (c) treated with clove oil, (d) treated with clove oil nanoemulsion, (e) treated with iron NPs

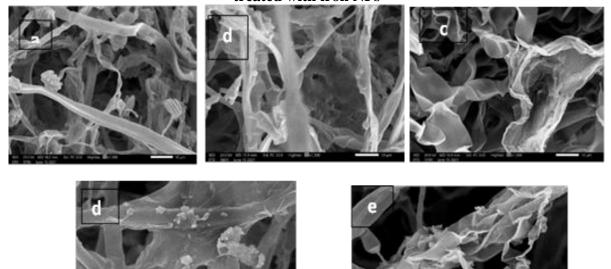


Figure 4. SEM of *A. solani* hyphae after a week of incubation at 27 °C. (a) Control, (b) treated with Azoxystrobin, (c) treated with clove oil, (d) treated with clove oil nanoemulsion, (e) treated with iron NPs

CONCLUSION

Azoxystrobin, iron NPs and essential oils of clove in normal and nanoemulsion forms had antifungal activities against *F. oxysporum, F. solani*, and *A. solani*. Treatments with tested compounds led to an increase in polyphenol oxidase enzyme in all tested fungi. SEM

indicated the shape of the hyphae for each fungus differs in control from the treatment. **ACKNOWLEDGMENT**

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REFERENCES

1. Abad, M.J., M. Ansuategui and P. Bermejo, 2007. Active antifungal substances from natural sources. Arkivoc., 7, 116-145

2. Abdel-Hafez, S. I. I., N. A. Nafady, I. R. Abdel-Rahim, A. M. Shaltout, J. Daro's and A. M. Mohamed, 2016. Assessment of protein silver nanoparticles toxicity against pathogenic *Alternaria solani*. 3 Biotech, 6:199

3. Abd-Elsalam, K. A. and A. R. Khokhlov, 2015. Eugenol oil nanoemulsion: antifungal activity against Fusarium oxysporum f. sp. vasinfectum and phytotoxicity on cottonseeds. Applied Nanoscience, 5: 255-265 4. Abu-Tahon, M. A., A. M. ogazy and G.S. 2022. Resistance assessment and Isaac. enzymatic responses of common bean (*Phaseolus vulgaris* L) against Rhizoctonia solani damping-off in response to seed presoaking in Vitex agnus-castus L. oils and foliar spray with zinc oxide nanoparticles. South African Journal of Botany, 146: 77-89

5. Ali, E. O., N. A. Shakil, V. S. Rana, D. J. Sarkar, S. Majumder, P. Kaushik, B. B. Singh and J. Kumar, 2017. Antifungal activity of nano emulsions of neem and citronella oils against phytopathogenic fungi, *Rhizoctonia solani* and *Sclerotium rolfsii*. Industrial Crops and Products, 108:379-387

6. Batiha, G. E., L. M. Alkazmi, L. G. Wasef, A. M. Beshbishy, E. H. Nadwa, E. K. Rashwan, 2020. *Syzygium aromaticum* L. (Myrtaceae): traditional uses, bioactive chemical constituents, phar- mycological and toxicological activities. Biomolecules, 10:202

7. Benincasa, M., F. Buiarelli, G. P. Cartoni and F. Coccioli, 1990. Analysis of lemon and bergamot essential oils by HPLC with microbore columns. Chromatographia, 30: 271-276

8. Chee, H. Y. and H. L. Min 2018. Antifungal activity of clove essential oil and its volatile vapor against dermatophytic fungi. Mycobiology, 35(4): 241-243

9. Ephrem, E., H. Greige-Gerges and H. Fessi, 2014. Charcosset C. Optimisation of rosemary oil encapsulation in polycaprolactone and scale-up of the process. *J. Microencapsul.* 31:746–753.

10. Esterbaner, H., E. Schwarzl and M. Hayn, 1977. A rapid assay for catechol oxidase and laccase using 2-nitro-5-thio benzoic acid. Anal. Biochem., (77): 486–494

11. Filoda, G., 2008. Impact of some fungicides on mycelium growth of *Colletotrichum gloeosporioides* (Penz.) Penz. & Sacc. Pestycydy/Pesticides, 3(4): 109-116

12. Gupta, S. and P. S. Variyar, 2016. Nanoencapsulation of essential oils for sustained release: Application as therapeutics and antimicrobials, pp. 641–672.

13. Hayes, W.J. and E. R. Laws, 1991. Handbook of pesticide toxicology, (1) 55-56

14. Ismail, A. M., 2021. Efficacy of Copper Oxide and Magnesium Oxide Nanoparticles on Controlling Black Scurf Disease on Potato. Egyptian Journal of Phytopathology, 49 (2):116-130.

15. Kanmani, P. and S. T. Lim, 2013. Synthesis and structural characterization of silver nanoparticles using bacterial exopolysaccharide and its antimicrobial activity against food and multidrug-resistant pathogens. Process Biochem; 48(7):1099– 1106

16. Khan, M. R. and Z. A. Siddiqui, 2021. Role of zinc oxide nanoparticles in the management of disease complex of beetroot (Beta vulgaris L.) caused by *Pectobacterium betavasculorum*, *Meloidogyne incognita*, and *Rhizoctonia solani*. Horticulture, Environment, and Biotechnology; 62: 225–241

17. Kim, Y., R. G. Bernard, B. Yoav and R. Choong-Min, 2012. Enhancement of Plant Drought Tolerance by Microbes. Plant Responses to Drought Stress; 383-413

18. Koba, K., A. Y. Nenonene, C. Raynaud, J. P. Chaumont and K. Sanda, 2011. Antibacterial Activities of the Buds Essential Oil of *Syzygium aromaticum* (L.). Journal of Biologically Active Products from Nature; 1: 42-51

19. Lin, J., D. Gong, S. Zhu, L. Zhang, and L. Zhang, 2011. Expression of PPO and POD genes and contents of polyphenolic compounds in harvested mango fruits about Benzothiadiazole-induced defense against anthracnose. Sci. Hortic.; 130: 85–89

20.LuciaGalovičová, P.Borotová, V.Valková, H.Ďúranová, P.Ł.Kowalczewski, H.A.H.S.Ahl, W.M.

Hikal, M. Vukic, T. Savitskaya, D. Grinshpan, N. L. Vukovic, 2021. Chemical Composition, *In Vitro* and *in vivo Situ* Antimicrobial and Antibiofilm Activities of *Syzygium aromaticum* (Clove) Essential Oil. Plants (Basel). 15, 10(10):2185

21. Machado, M., A. M. Dinis, L. Salgueiro, J. B. A. Custódio, C. Cavaleiro and M. C. Sousa, 2011. Anti-Giardia activity of *Syzygium aromaticum* essential oil and eugenol effects on growth, viability, adherence, and ultrastructure. Experimental Parasitology; 127: 32-39

22. Manikandan, G., and R. Ramasubbu 2021. Biosynthesis of Iron Nanoparticles from *Pleurotus florida* and its Antimicrobial Activity against Selected Human Pathogens. Indian J Pharm Sci.; 83(1):45-51

23. Mayer A. M., 2006. Polyphenol oxidases in plants and fungi: Going places? Phytochemistry; 67(21): 2318-2331

24. Mohamed, Y. M., A. M. Azzam, B. H. Amin and N. A. Safwat, 2015. Mycosynthesis of iron nanoparticles by Alternaria alternate and its antibacterial activity. Academic Journal; 14(4): 1234-1241

25. Mossa, A. H., M. M. M. Samia, H. E. El-Sayed, S. A. Ziedan, Ibrahim and F. S. Ahmed, 2021. Development of eco-friendly nanoemulsions of some natural oils and evaluating their efficiency against postharvest fruit rot fungi of cucumber. Industrial Crops and Products; 159: 113049

26. Nandhini, M., S. Rajini, A. Udayashankar, S. Niranjana, O. S. Lund and H. Shetty, 2019. Biofabricated zinc oxide nanoparticles as an eco-friendly alternative for growth promotion and management of downy mildew of pearl millet. Crop Prot.; 121: 103–112

27. Park, M. J., K. S. Gwak, I. Yang, W.S. Choi, H. J. Jo, J. W. Chang, E. B. Jeung and I. G. Choi, 2007. Antifungal activities of the essential oils in *Syzygium aromaticum* (L.) Merr. Et Perry and *Leptospermum petersonii* Bailey and their constituents against various dermatophytes. J. Microbiol.; 45: 460-465

28. Rana I. S.; Aarti S. R. and Ram C. R. (2011). Evaluation of antifungal activity in the essential oil of *Syzgium aromatium* L. by extraction, purification, and analysis of its main component eugenol. Brazilian Journal of Microbiology; 42: 1269-1277

29. Sharma, A., K. S. Naveen, S. Ankit, K. Arti, D. Saurabh, S. Satyawati and K. Bishwajit, 2018. Clove and lemongrass oilbased non-ionic nanoemulsion for suppressing the growth of plant pathogenic Fusarium oxysporum f.sp. lycopersici. Industrial Crops and Products; 123: 353-362

30. Sharma, N., S. Jandaik, S. Kumar, M. Chitkara and I. S. Sandhu, 2016. Synthesis, characterisation and antimicrobial activity of manganese- and iron-doped zinc oxide nanoparticles. Journal of Experimental Nanoscience; 11(1): 54-71

31. Soffan, A., S. S. Alghamdi and A. S. Aldawood, 2014. Peroxidase and Polyphenol Oxidase Activity in Moderate Resistant and Susceptible *Vicia faba* Induced by Aphis craccivora (Hemiptera: Aphididae) Infestation. Journal of Insect Science; 14(1): 285

32. Sunitha, A., R. S. R. Isaac, G. Sweetly, S. S. kshmi, R. Arsula and P. K. Praseetha, 2013. Evaluation of the antimicrobial activity of biosynthesized iron and silver Nanoparticles using the fungi *Fusarium oxysporum* and *Actinomycetes* sp. on human pathogens. Nano Biomed., 5(1): 39-45

33. Taisan, W. A., A. H. Bahkali, A. M. Elgorban and M. A. El – Metwally, 2014. Effective influence of essential oils and microelements against *Sclerotinia Sclerotiorum*. International Journal of Pharmacology, 10 (5): 275 – 281

34. Thabet, M. and K. Walaa, 2018. Antifungal activities of clove oil against root rot and wilt pathogens of tomato plants. American-Eurasian J. Agric. & Environ. Sci., 18 (3): 105-114

35. Thakkar, K. N., S. S. Mhatre and R. Y. Parikh, 2010. Biological synthesis of metallic nanoparticles. Nanomedicine, 6(2):257–262

36. Tomazoni, E. Z., G. F. Pauletti, R. T. S. Ribeiro, S. Moura and J. Schwambach, 2017. *In vitro* and *in vivo* activity of essential oils extracted from *Eucalyptus staigeriana*, *Eucalyptus globulus*, and *Cinnamonum camphora* against *Alternaria solani* Sorauer causing early blight in tomato. Scientia Horticulturae, 223: 72-77.

37. Villamizar-Gallardo, R., J. F. O. Cruz and O. O. Ortíz-Rodriguez, 2016. Fungicidal effect of silver nanoparticles on toxigenic fungi in

cocoa. Pesq. agropec. bras., Brasília; 51(12): 1929-1936

38. Winer, B.J. 1971. Statistical Principles in Experimental Design. 2nded. New York: McGraw Hill, USA

39. Xing, Y., X. Qingliian, L. Xihong, C. Zhenmin and Y. Juan, 2011. Antifungal activities of clove oil against *Rhizopus nigicans*, *Aspergillus flavus*, and *Penicillium citrinum in vitro* and in wounded fruit test. Journal of Food Safety, 32: 84–93

40. Yahyazadeh M., R. Omidbaigi, Z. R. Rasoul and H. Taheri, 2008. Effect of some essential oils on mycelial growth of *Penicillium digitatum* Sacc. World J Microbiol Biotechnol, 24:1445–1450

41. Yulia, E., 2005. Antifungal activity of plant extracts and oils against fungal pathogens of pepper (*Piper nigrum L.*), cinnamon (*Cinnamomum zeylanicum Blume*), and turmeric (*Curcuma domestica* Val.). M.Sc. (Research) thesis.