

THE PREVALENCE OF BLANDM, BLAVIM GENES AMONG *ENTEROBACTER CLOACAE* BACTERIA

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ABSTRACT

This study was aimed to detect the prevalence of *blaVIM* and *blaNDM* genes among *Enterobacter cloacae* isolates that were isolated from Iraqi patients. About 50 bacterial isolates were collected from different hospitals in Baghdad city and all these isolates were diagnosed using biochemical tests and CHROMagar culture media and conformed using the Vitek II system. The antibiotic susceptibility for *E.cloacae* isolates was determined using the disk diffusion (Kirby Bauer) method and the results showed that these bacteria were showed resistance to Cefepime and Meropenem antibiotics in percentage (50%), (40%) respectively. In addition it is resistant to Amikacin, ampicillin/sulbactam, Piperacillin, and aztreonam was (20%), (50%), (40%), and (55%) respectively. Also it has been noted that all isolates showed resistance to Cefixime in percentage (100%). EDTA combined disc test was used to detect the ability of *E. cloacae* isolates to produce carbapenemase and the results were showed that only 8 isolates were gave positive results , While the prevalence of *blaVIM* and *blaNDM* was determined using the conventional Polymerase chain reaction technique. And the result showed that only four isolates harbored *blaNDM* while *blaVIM* was present in 3 isolates

Keywords: Enterobacteriaceae, carbapenem resistance, metallo β lactamase, EDTA combined disc test

محمود

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انتشار جينات *blaNDM* و *blaVIM* بين بكتريا *Enterobacter cloacae*

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المستخلص

هدفت هذه الدراسة الى تحديد انتشار جينات *blaVIM* و *blaNDM* بين عزلات بكتريا *Enterobacter cloacae* والتي عزلت من المرضى العراقيين. مايقارب خمسين عزلة بكتيرية تم جمعها من مختلف مستشفيات مدينة بغداد وتم تشخيص كل العزلات باستعمال الاختبارات الكيموحيوية ووسط CHROMagar ومن ثم تم تأكيد التشخيص باستعمال نظام Vitek II. تم تحديد الحساسية البكتيرية لعزلات *E.cloacae* باستعمال طريقه انتشار القرص (Kirby Bauer) وظهرت النتائج بان البكتريا اظهرت مقاومة لمضادات Cefepime Meropenem, بنسبة مئوية (50%) و(40%) على التوالي. بالاضافة, كانت البكتريا مقاومة ل Amikacin, ampicillin/sulbactam, Piperacillin, and aztreonam ب (20%), (50%), (40%), (55%) بالتعاقب. كذلك, لوحظ بان كل العزلات مقاومة ل Cefixime بنسبة مئوية (100%). لقد استخدم اختبار EDTA combined disc لتحديد قدرة عزلات *E. cloacae* على انتاج انزيم carbapenemase وظهرت النتائج بان فقط 8 عزلات اظهرت نتائج ايجابية, بينما انتشار جينات *blaVIM* و *blaNDM* قد حددت باستعمال تقنية تفاعل البلمرة المتسلسل التقليدي. وظهرت النتائج بان فقط اربع عزلات تحمل *blaNDM* بينما ظهر *blaVIM* في ثلاث عزلات فقط.

كلمات مفتاحية: العائلة المعوية, مقاومة الكاربينيم, انزيمات البيتا لكتام المعدنية, اختبار EDTA combined disc

INTRODUCTION

Enterobacter is a rod-shaped, gram-negative bacteria belonging to the genus Enterobacteriaceae, which is facultative anaerobic and is not capable of spore production (6). The most clinically important species of this genus is *Enterobacter cloacae*, which is a bacterial strain usually present in the gut microbiota of healthy humans (12). As well as these bacteria is also recognized as the most common pathogen in nosocomial infections (13). This pathogen capable of producing a wide range of infections, such as urinary tract infections, pneumonia, and septicemia (1). This pathogen has recently emerged as a drug-resistant bacterial species; in both human and veterinary medicine, the production of antimicrobial resistance across *Enterobacter* spp., including resistance against -spectrum cephalosporins, is of serious importance (9). There is a rising understanding that *E.cloacae* behind *Escherichia coli* and *Klebsiella pneumonia* is also the third main genus of Enterobacteriaceae implicated in nosocomial infections. Carbapenems are commonly used to treat infections due to multidrug-resistant (MDR) Gram-negative rods containing cephalosporins or ESBL, such as *Enterobacter* species. However, carbapenemase-producing bacteria have prevented the use of carbapenems in medical practice (18, 11). CRE emergence, particularly with *Enterobacter* spp., is linked to the development of carbapenemases. A wide variety of carbapenemases was found in these bacteria which include ambler class A blaKPC, Metallo- β -lactamases (MBL) class B such as *blaVIM*, *blaIMP*, *blaNDM*, etc. (3, 20). CRE-caused nosocomial infections are serious challenges worldwide, and this is due to the quick spread of these infections across the globe. The mortality rate of CRE-induced infections is high, varying from 30-44 percent (3, 17). There are only a few studies regarding the abundance of carbapenem-resistant *Enterobacter cloacae* that were discovered in Baghdad This study was aimed to detect the prevalence of *blaVIM* and *blaNDM* genes among *Enterobacter cloacae* isolates that were isolated from Iraqi patients Hospital, Iraq, this research was performed.

MATERIALS AND METHODS

Collection and identification of the specimen: About fifty bacterial specimens were collected from patients admitted to Baghdad hospitals as well as these bacteria were collected from wounds, UTI, and burned skin. All these 50 bacterial specimens were identified using biochemical tests and CHROMagar culture media, as well as all of them, were confirmed using ViteK II system.

Antibiotic susceptibility

The antibiotic susceptibility for *E.cloacae* was determined using Kirby Bauer method. This method was used to determine the susceptibility of *E.cloacae* to Meropenem, Cefepime, ampicillin/ sulbactam, Amikacin, Piperacillin, aztreonam, and Cefixime, and the result was interpreted according to the guideline of CLSI (2020).

EDTA combined disc test

This test was performed depends on the way provided by Galani *et al* (7). In which bacterial specimen was transferred from overnight growth of *E.cloacae* in brain heart infusion agar to 5 ml of normal saline and the inoculum was adjusted to 0.5 McFarland then the bacterial specimen was transferred from inoculum to the Muller Hinton agar. After the spreading of bacteria on the agar, Imipenem and Imipenem+ EDTA were placed on the agar surface and incubated for 18 hr at 37 C, and the results were interpreted according to the guideline of CLSI (2020).

Molecular detection of carbapenemase

DNA was extracted from *E.cloacae* isolates using WizPrep gDNA Mini Kit (Korea). The prevalence of carbapenem resistance genes (*bla* NDM and *bla* Vim) was detected using Conventional PCR technique. The primer used for detection of these genes was designed using (Primer 3 program <https://primer3.ut.ee/>) and was provided by alpha DNA company (Canada) and the sequence of the primer was illustrated in the table (1). The detection of these genes was performed by amplifying these genes in Conventional PCR technique with final concentration volume 25 μ l which was prepared by mixing 12.5 μ l of master mix with 2 μ l of primers, 3 μ l of DNA and the volume was completed to 25 μ l by the addition 7.5 μ l of nuclease free water. The mixture was placed in the thermo-cycler (Bioneer Korea),

and the steps of amplification was as following: one cycle at 95°C for 5mins ,then 35 cycles as 30s at 95 °C ,annealing step at 58

°C for VIM and 52 for NDM for 30 S and extension for 10 mins at 72 °C and final extension at 72°C for 10 mins.

Table 1. Sequence of the Primers (Vim and Imp)

GENE		SEQUENCE 5' → 3'	Reference
Vim	F	GATGGTGTGGTTCGCATA	44
	R	CGAATGCGCAGCACCAG	
	F	ATGGAATTGCCCAATATTATGC	
NDM	R	CGAAAGTCAGGCTGTGTTG	45

RESULTS AND DISCUSSION

From 50 bacterial specimen that are collected from different clinical source of Baghdad hospitals (UTI, wounds and burned skin) only 20 bacterial specimen was conformed using biochemical test and CHROMagar and the result of identification was reconfirmed using Vitek II system while the remaining of collected specimens were related to another pathogenic bacteria.

Antibiotics susceptibility

The antibiotic susceptibility test showed 10 isolates of *E.cloacae* were resistant to

Cefepime (50%) and 8 isolates were resistant to Meropenem (40%) as well as only 4 isolates were resistant to Amikacin (20%). While another 10 isolates were resistant to ampicillin/sulbactam (50%), in addition 11 isolates were found to be responsible for providing resistance to aztreonam (55%), while resistance to Piperacillin was provided by only 8 isolates (40%) of *E.cloacae* and finally, all isolates were responsible for providing resistance to Cefixime (100%). The percentages of antibiotic susceptibility were showed in Figure (1).

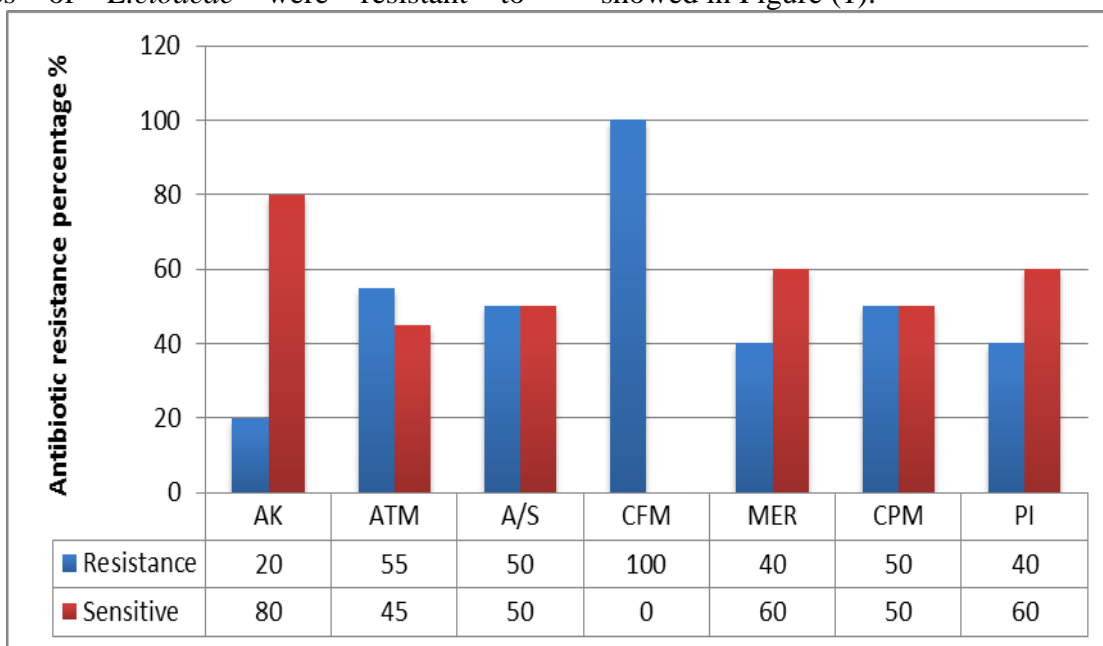


Figure 1. The antibiotic susceptibility test of *E.cloacae*. In which AK (Amikacin), ATM (Aztreonam), A/S (Ampicillin/sulbactam), CFM (Cefixime), MER (Meropenem), CPM (Cefepime), and PI (Piperacillin)

Result of EDTA combined disc test

This test was conducted only on 8 isolates because these isolates were resistant to all tested antibiotics and the result shows these 8 isolates were resistant to Imipenem and

become sensitive after the addition of Imipenem + EDTA as shown in Figure (2), Which indicate these bacteria produce carbapenem-resistant genes.

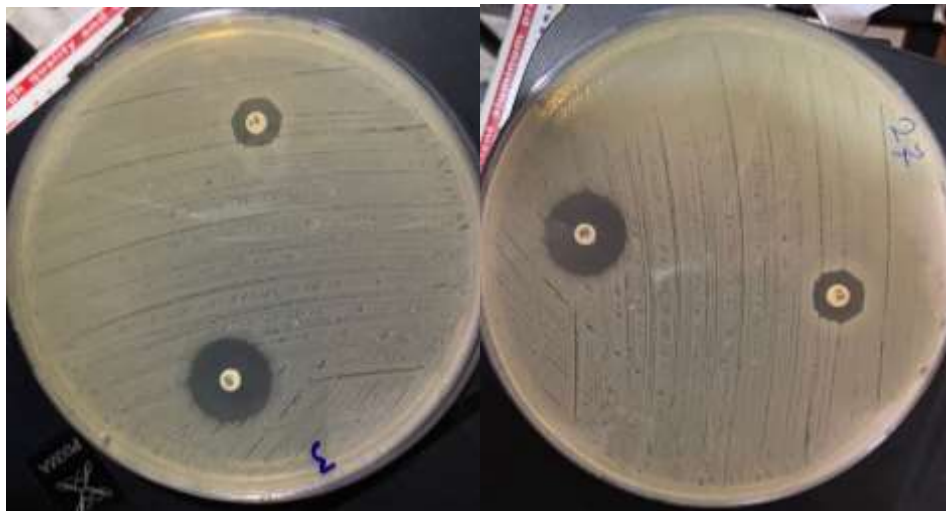


Figure 2. EDTA combined disc test

Molecular detection of Carbapenem resistance genes: This test was also conducted only on 8 isolates of *E. cloacae* and the result show out of 8 isolates only 4 isolates was

carried *blaNDM* with molecular size of 490 bp while *blaVIM* was found only on three isolates with molecular size 390 bp as shown the following Figures (3A) and (3B).

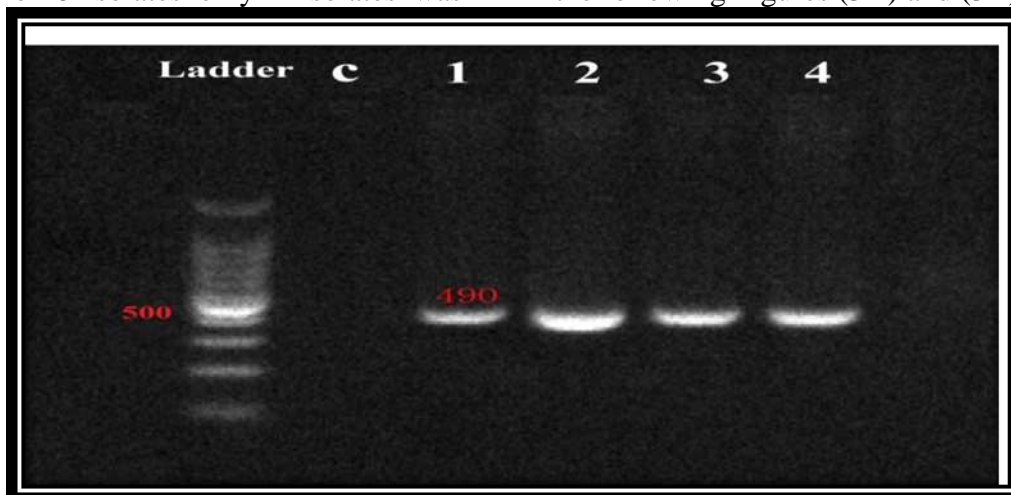


Figure (3a) Detection the prevalence of *blaNDM* gene in *E.cloacae* using gel electrophoresis in which agarose concentration was 2% and DNA ladder from 100bp to 1500 bp

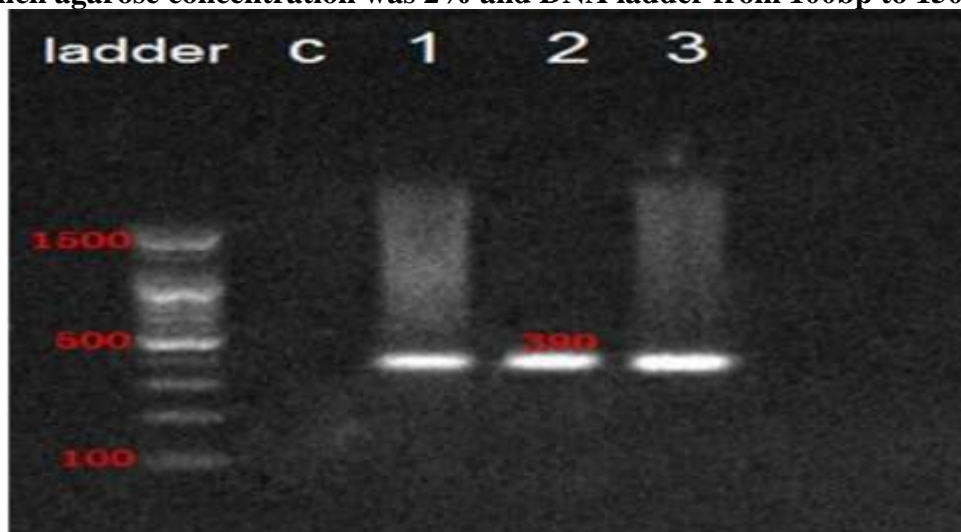


Figure (3b) Detection the prevalence of *blaVIM* gene in *E. cloacae* using gel electrophoresis in which agarose concentration was 2% and DNA ladder from 100bp to 1500 bp

There are few therapeutic possibilities with a rise in the production of MDR carbapenemases developing *Enterobacter cloaca*. Therefore, early detection is important to reduce the mortality of nosocomial infections caused by these species. (10, 18). *E. cloacae* are responsible for a wide range of nosocomial infections, especially wound infections, pneumonia, and bacteremia (15, 5). In the current study, most isolates (100%) were recovered from different clinical sources. In Consistent with our work, different researchers were documented isolation of these bacteria from different clinical sources included respiratory tract samples (17), skin (9). In contrast, in several studies from different countries like Brazil (18), China (22), a global surveillance program (14), and Korea (11). Furthermore, the most common sites for *Enterobacter* isolation were abdominal and blood samples. According to the results of susceptibility testing, 60% of our isolates showed MDR phenotype, this makes them a threat to public health in our area. This observation is not consistent with two research previously performed in Iran with a prevalence of 17.5% and 47.5% respectively (8, 16). Carbapenem resistance was characterized as resistance to one or more carbapenems in compliance with the CLSI guidelines (18). In the current analysis, 40% of isolates were not susceptible to Imipenem (carbapenem-resistant) and 8.7%, 29.2%, 25.7%, 35.1%, respectively, were recorded in many other reports from different regions (11, 5, 8, 2, 21). In this study, the pattern of antimicrobial resistance and the existence of two carbapenemase genes were characterized among 8 *E. cloaca* clinical isolates. Which were Imipenem resistance and EDTA combined test positive that were recovered from an Iraqi population. However, Carbapenemase-producing *E. cloacae* has been reported in many countries and in current study, among evaluated carbapenemase genes only *bla*NDM (n=4/8, 50%) and *blavim* (n=3/8, 37%) were detected. This study is the first reported presence of *bla*NDM among clinical isolates of *Enterobacter cloaca* in Iraq. While, studies published from other countries as Iran, China (5), Spain (4), Korea (11), and Mexico (3) with frequencies of 2.8%, 0%, 0%,

and 100%, respectively. Indeed, an *E. cloaca*, the species which produces VIM-1, has been frequently recorded in Europe and has been a major nosocomial pathogen in southern Europe and Asia (21, 4). In this study, 3/8 of the isolates examined harbored the *bla*VIM-gene. An analysis in Spain showed that 52% and 100% of the *E. cloacae* isolates were found to be *bla*VIM producers. On the other hand, in Far Eastern studies, the rate of *bla*VIM (0.25%) was noted too close to our results. In addition, in an Iranian study, no carbapenemase gene was present in *Enterobacter* spp isolates Combined, the varying distributions between regions and strains in genetic diversity are more definitely attributable to the differences in results. Since this study not evaluate the existence of other carbapenemase genes from different groups of β -lactamases, we had to set the study's limitations to one being the comparatively limited sample size and the other being the other limitation of the work which is that to properly determine beta-lactam tolerance in our isolates, we could not evaluate the presence of different carbapenemase genes from different classes of β -lactamases.

Conclusion: In this study, we found for the first time the prevalence of *bla*NDM in *E. cloaca* from an Iraqi patients. Due to the rise in carbapenem-producing MDR *Enterobacter cloacae* in our country, hygiene regulations for hospitals and other healthcare facilities in developed countries will be required to monitor the fast spread of MDR *Enterobacter* isolates.

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REFERENCES

1-Annavaajhala, M. K. Gomez-Simmonds, A.Macesic, N. Sullivan ,S.B. Kress , A.Khan , S.D.Giddins , M.J.Stump , S.Kim , G.I. Narain , R.E.C.Verna and A.C. Uhlemann, 2019. Colonizing multidrug-resistant bacteria and the longitudinal evolution of the intestinal microbiome after liver transplantation. *Nature communications*, 10(1), 4715.

<https://doi.org/10.1038/s41467-019-12633-4>

- 2-Armin, S., F. Fallah, , L. Azimi, H. S. Kafil, H. Ghazvini, K. Hasanzadeh, and A. Karimi, 2018. Warning: spread of NDM-1 in two border towns of Iran. *Cellular and molecular biology* (Noisy-le-Grand, France), 64(10), 125–129
- 3-Bocanegra-Ibarias, P., E. Garza-González, R. Morfín-Otero, H. arrios, L.Villarreal-Treviño, E.Rodríguez-Norieg, U.Garza-Ramos ,S. Petersen-Morfín and J. Silva-Sanchez 2017. Molecular and microbiological report of a hospital outbreak of NDM-1-carrying Enterobacteriaceae in Mexico. *PloS one*, 12(6), e0179651.
<https://doi.org/10.1371/journal.pone.0179651>
- 4-Coelho A., N. Piedra-Carrasco, R. Bartolomé, J. N.Quintero-Zarate , N.Larrosa, , T.Cornejo-Sánchez, G. Prats, M. P.Garcillán-Barcia, F.de la Cruz and J. J.González-Lopéz 2012. Role of IncHI2 plasmids harbouring blaVIM-1, blaCTX-M-9, aac(6)-Ib and qnrA genes in the spread of pneumoniae multiresistant Enterobacter cloacae and Klebsiella strains in different units at Hospital Vall d'Hebron, Barcelona, Spain. *International journal of antimicrobial agents*, 39(6), 514–517.
<https://doi.org/10.1016/j.ijantimicag.2012.01.006>
- 5-Dai, W., S. Sun, P. Yang, ,S. Huang, X.Zhang and L. Zhang, 2013. Characterization of carbapenemases, extended spectrum β -lactamases and molecular epidemiology of carbapenem-non- susceptible Enterobacter cloacae in a Chinese hospital in Chongqing. *Infection, genetics and evolution : journal of molecular epidemiology and evolutionary genetics in infectious diseases*, 14, 1–7.
<https://doi.org/10.1016/j.meegid.2012.10.010>
- 6-Davin-Regli, A., and J. M. Pagès, 2015. Enterobacter aerogenes and Enterobacter cloacae; versatile bacterial pathogens confronting antibiotic treatment. *Frontiers in microbiology*, 6, 392.
<https://doi.org/10.3389/fmicb.2015.00392>
- 7-Galani, I., P.D.Rekatsina, D.Hatzaki, , D.Plachouras, , M.Souli, and H.Giamarellou, (2008). Evaluation of different laboratory tests for the detection of metallo- β -lactamase production in Enterobacteriaceae. *Journal of antimicrobial chemotherapy*, 61(3), 548-553
- 8-Ghanavati, R., M. Emaneini, ,D. Kalantar-Neyestanaki, A.S.Maraji ,M. Dalvand, R.Beigverdi and F.Jabalamel, 2018. Clonal relation and antimicrobial resistance pattern of extended-spectrum β -lactamase- and AmpC β -lactamase-producing Enterobacter spp. isolated from different clinical samples in Tehran, Iran. *Revista da Sociedade Brasileira de Medicina Tropical*, 51(1), 88–93.
<https://doi.org/10.1590/0037-8682-0227-2017>
- 9-Harada, K., T.Shimizu , Y.Mukai, K.Kuwajima, T. Sato, A.Kajino, M. Usui, Y Tamura, Y. Kimura, T. Miyamoto, Y.Tsuyuki, A. Ohki and Y. Kataoka, 2017. Phenotypic and molecular characterization of antimicrobial resistance in Enterobacter spp. isolates from companion animals in Japan. *PloS one*, 12(3), e0174178.
<https://doi.org/10.1371/journal.pone.0174178>
- 10-Hoffmann, H., E. Stürenburg, J. Heesemann, and A. Roggenkamp 2006. Prevalence of extended-spectrum beta-lactamases in isolates of the Enterobacter cloacae complex from German hospitals. *Clinical microbiology and infection : the official publication of the European Society of Clinical Microbiology and Infectious Diseases*, 12(4), 322–330.
<https://doi.org/10.1111/j.1469-0691.2006.01360.x>
- 11-Lee, J. Y., Y.K.Hong, H.Lee and K.S. Ko 2017. High prevalence of non-clonal imipenem-nonsusceptible Enterobacter spp. isolates in Korea and their association with porin down-regulation. *Diagnostic microbiology and infectious disease*, 87(1), 53–59.
<https://doi.org/10.1016/j.diagmicrobio.2016.10.004>
12. Leong, L., D. Shaw, L.Papanicolas, D. Lagana, I. Bastian and G.B. Rogers 2017. Draft Genome Sequences of Two Enterobacter cloacae subsp. cloacae Strains Isolated from Australian Hematology Patients with Bacteremia. *Genome announcements*, 5(33), e00756-17.
<https://doi.org/10.1128/genomeA.00756-17>
13. Moradigaravand, D., C. J. Boinett, V. Martin, J. S. Peacock and J. Parkhill, 2016. Recent independent emergence of multiple multidrug-resistant Serratia marcescens clones within the United Kingdom and Ireland.

- Genome research, 26(8), 1101–1109. <https://doi.org/10.1101/gr.205245.116>
14. Peirano, G., Y. Matsumura, M. D. Adams, P. Bradford, M. Motyl, L. Chen, B. N. Kreiswirth, and J. Pitout, 2018. Genomic Epidemiology of Global Carbapenemase-Producing *Enterobacter* spp., 2008-2014. *Emerging infectious diseases*, 24(6), 1010–1019. <https://doi.org/10.3201/eid2406.171648>
15. Pérez, A., M. Poza, A. Fernández, M. Fernández, S. Mallo, M. Merino, S. Rumbo-Feal, M. P. Cabral, and G. Bou 2012. Involvement of the AcrAB-TolC efflux pump in the resistance, fitness, and virulence of *Enterobacter cloacae*. *Antimicrobial agents and chemotherapy*, 56(4), 2084–2090. <https://doi.org/10.1128/AAC.05509-11>
16. Peymani, A., T. N. Farivar, R. Sanikhani, A. Javadi and R. Najafipour 2014. Emergence of TEM, SHV, and CTX-M-extended spectrum β -lactamases and class 1 integron among *Enterobacter cloacae* isolates collected from hospitals of Tehran and Qazvin, Iran. *Microbial drug resistance (Larchmont, N.Y.)*, 20(5), 424
17. Rosa, J. F., C. Rizek, A. P. Marchi, T. Guimaraes, L. Miranda, C. Carrilho, S. A. Levin, and S. F. Costa, 2017. Clonality, outer-membrane proteins profile and efflux pump in KPC- producing *Enterobacter* sp. in Brazil. *BMC microbiology*, 17(1), 69. <https://doi.org/10.1186/s12866-017-0970-1>
18. Shahid, M., A. Malik, M. Akram, L. M. Agrawal, A. U. Khan, and M. Agrawal 2008. Prevalent phenotypes and antibiotic resistance in *Escherichia coli* and *Klebsiella pneumoniae* at an Indian tertiary care hospital: plasmid-mediated cefoxitin resistance. *International journal of infectious diseases : IJID : official publication of the International Society for Infectious Diseases*, 12(3), 256–264. <https://doi.org/10.1016/j.ijid.2007.08.008>
19. Tação, M., A. Correia, and I. S. Henriques, 2015. Low Prevalence of Carbapenem-Resistant Bacteria in River Water: Resistance Is Mostly Related to Intrinsic Mechanisms. *Microbial drug resistance (Larchmont, N.Y.)*, 21(5), 497–506. <https://doi.org/10.1089/mdr.2015.0072>
20. Villa, J., E. Viedma, P. Brañas, M. A. Orellana, J. R. Otero, and F. Chaves 2014. Multiclonal spread of VIM-1-producing *Enterobacter cloacae* isolates associated with In624 and In488 integrons located in an IncHI2 plasmid. *International journal of antimicrobial agents*, 43(5), 451–455. <https://doi.org/10.1016/j.ijantimicag.2014.02.006>
21. Wang, S., S. Z. Xiao, F. F. Gu, J. Tang, X. K. Guo, Y. X. Ni, J. M. Qu, and L. Z. Han. 2017. Antimicrobial susceptibility and molecular epidemiology of clinical *Enterobacter cloacae* bloodstream isolates in Shanghai, China. *PloS one*, 12(12), e0189713. <https://doi.org/10.1371/journal.pone.0189713>