EFFECT OF CALCIUM AND COLE VIT D₃ IN OVO INJECTION ON HATCHABILITY, BONE AND BLOOD BIOCHEMICAL DEVELOPMENT AT POSTHATCH

M. AG. Mustafa¹ N. AG. Mustafa² R. S. Rasheed³ Assist. professor Lecturer Assist. professor ^{1, 2} Dept. of Animal Resources. Coll. of Agric.. Salahaddin University-Erbil-Iraq ³ Dept. of Animal Prod.. Coll. of Agric. Engin. Sci. University of Baghdad ¹ mahbuba.mustafa@su.edu.krd

ABSTRACT

The aim of this study to investigate the role of in ovo injection in broiler breeder (Ross-308) fertile eggs with calcium, vitamin D_3 and their mixture at 18 d of incubation on hatchability, tibia bone and blood properties post-hatching, included five groups: (non-injected and in ovo injected 100 µL sterilized distill water) controls as well as eggs that were injected with the 100 µL sterilized distill water either 0.8 mg calcium (Ca), 0.8 mg Cole vit D_3 [colecalciferol or 25-OHD₃] and mix of 0.8 mg Ca + 0.8 mg Cole vit D_3 . The results showed that an in ovo injection with calcium, Cole vit D_3 and their mix presented significantly higher in hatchability (%), body performances, tibia bone properties [length, breaking strength, Ca and P concentrations, ash %] and blood biochemical analysis [Ca and P minerals concentrations, vit D_3 and parathyroid hormone (PTH)] at 0 and 35 d posthatch. While, blood calcitonin hormone (CT) at 0 and 35 d posthatch and blood Ca: P ratio had recorded lower concentration at 0 posthatch, however had seen non-significant differences among all the groups in tibia width and Ca:P ratio in tibia bone at 0 and 35 d posthatch and also non-significant among all the groups of in ovo injection and control in blood Ca: P ratio at 35 d posthatch. The in ovo injection with Cole vit D_3 and mixture of Ca and Cole vit D_3 showed more effectiveness in most characteristics.

Keyword: in ovo, calcium, Vit D₃, hatching, minerals, hormones.

المستخلص

تهدف الدراسة الى كشف دور حقن بيض أمهات فروج اللحم (Ross-308) بالكالسيوم وفيتامين D3 ومزيجهما عند عمر 18 يوما من حضن البيض على قابلية الفقس، وزن الأفراخ، طول عظم الساق وخصائص الدم.بعد الفقس وشملت حمسة مجاميع: (مجموعة السيطرة: دون حقن للبيض، حقن البيض100 ميكرولتر من الماء المقطر المعقم، حقن البيض بـ 0.8 ملغم كالسيوم مذاب بـ 100 ميكرولتر من بالماء المقطرالمعقم، 0. حقن البيض بـ 0.8 ملغم كولي فيتامين د3 مذاب بـ 100 ميكرولتر من بالماء المقطرالمعقم، حقن البيض بـ 108 بالماء المقطرالمعقم، 0. حقن البيض بـ 0.8 ملغم كولي فيتامين د3 مذاب بـ 100 ميكرولتر من بالماء المقطرالمعقم، حقن البيض بالكالسيوم، كولي فيتامين د3 ملغم كولي فيتامين د3 مذاب بـ 100 ميكرولتر من بالماء المقطرالمعقم، حقن البيض بالكالسيوم، كولي فيتامين د3 ومزيجها تفوقا في نسبة الفقس، الأداء الأنتاجي، خصائص عظمة الساق (الطول، قوة امقاومة الكس، تراكيزالكالسيوم والفوسفور ونسبة الرماد) والتحليل البيوكيميائي للدم [تراكيز (عنصري الكالسيتونين في الم و 3.0 و 3.5 يوم بعد الفقس تراكيزالكالسيوم، كولي فيتامين د3 ومزيجها تفوقا في نسبة الفقس، الأداء الأنتاجي، خصائص عظمة الساق (الطول، قوة امقاومة الكسر، تراكيزالكالسيوم والفوسفور ونسبة الرماد) والتحليل البيوكيميائي للدم [تراكيز (عنصري الكالسيتونين في الدم بعمر 0 و 35 يوم بعد الفقس تراكيزالكالسيوم: الفسفور وسبطت تركيز أقل عند 0 بعد الفقس ، بينما كل مجاميع الدراسة لم تختلف معنويا في عرض الساق ونسبة ونسبة الكالسيوم: الفسفور وسبطت تركيز أقل عند 0 بعد الفقس ، بينما كل مجاميع الدراسة لم تختلف معنويا في عرض الساق ونسبة ونسبة الكالسيوم: الفسفور في عظمة الساق بعمر 0 و 35 يوما ونسبة الكالسيوم: الفسفور بالدم عند عمر 35 يوما بعد الفقس حالكالسيوم: الفسفور في عظمة الساق بعمر 0 و 35 يوما ونسبة الكالسيوم: الفسفور بالدم عند معر 3.5 يوما بعد الفقس. العلق مراسة م تختلف معنويا في عرض الساق ونسبة حفن البيض بـ كولي فيتامين د 3 ومزيج من (الكالسيوم و كولي فيتامين د 3) فعالية أكبر في معظم الصفات المدروسة.

الكلمات المفتاحية: حقن البيض، الكالسيوم، كولى فيتامين د، الفقس، المعادن، الهرمونات.

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INTRODUCTION

Vitamin D plays an essential role in bone mineralization and calcium homeostasis via regulation of parathyroid hormone (PTH) secretion (20). More recently, there has been growing evidence for a role of vitamin D in extra skeletal health (8). Vitamin D plays an important role in maintaining an adequate level of serum calcium and phosphorus. Without vitamin D, only 10 to 15% of dietary calcium and about 60% of phosphorus is absorbed (10). Therefore vitamin D has a great effect in forming and maintaining strong bones. It has also recently been found that vitamin D receptors exist in a variety of cells thus it has a biological effect on more than mineral metabolism (21). The commercial in ovo injection of 25(OH)D3 was reported to improve the hatchability of fertilized broiler hatching eggs without having detrimental effects on hatchling quality (1). Also, in a later related study, it was shown that in ovo injection of up to 1.20 µg of 25-OHD₃ had no detrimental effects on survival or overall posthatch performance, including body weight gain, of broilers (2). The presence of cholecalciferol in eggs is very important to support the embryo Ca metabolism during incubation. Therefore, deficiency of this vitamin can lead to reduced hatchability, which can be specially related to late embryo mortality (11). Avian embryos assimilate large amounts of calcium in their bones in a short time. The chicken embryo, for instance, accumulates over 100 mg of calcium from the egg shell across the chorio-allantoic membrane from days 10-12 of embryonic life until hatching day (13). (14) conducted to in ovo vitamin D3 at ages 15, 16, 17 days of incubation increased the rate of calcium mobilization from the egg shell to the embryo and increased calcium concentration in all embryonic compartments. The objective of this study was to investigate the effects of the in ovo injection of 25(OH)D3 on d 18 of incubation on hatchability, posthatch performance. tibia mineralization and hormones concentrations in blood plasma.

MATERIALS AND METHODS

Experimental design

This experiment was conducted at Taqtaq broiler breeder and hatchery farm Erbil/Iraq, 1500 eggs were collected from commercial broiler breeder (Ross 308) hens at age 58 wks. the eggs were randomly allocated to 5 groups each group contain 3 replicates (were equally represented on each of 3 tray levels (blocks) of the incubator). The average weight of egg is 65 ± 2 g. The treatment groups on each tray level were randomly arranged with respect to their arrangement on the other tray levels.

In ovo injection

In ovo injection solution preparation, egg handling, and use of a manual in ovo injection in chorioalantoic membrane (CAM) by insulin syringe, eggs of each group application on d 18 of incubation in this study. The treatment groups included: (non-injected and in ovo injected 100 µL sterilized distill water) controls as injected with the 100 µL sterilized distill water of carrying either 0.8 mg calcium (Ca), 0.8 mg Cole vit D₃ (colecalciferol or 25-OHD₃) produced by Sterling UK. and mix of $0.8 \text{ mg Ca} + 0.8 \text{ mg Cole vit } D_3$ respectively. After the injection, the pinhole site was sealed with sterile paraffin wax and eggs were returned to the incubator. On the 19th day of incubation, eggs were shifted to the hatchery and kept in the respective pedigree hatching boxes Immediately.

Hatchability measurements

On the hatching day, chicks were weighed and hatching of fertile and total eggs, hatch window, total dead embryo and culled chicks percentages were recorded.

Broiler experimental design

sixty chicks of each group replicate were randomly selected and placed in floor pens in a light-controlled research facility. Chicks were placed in pens that corresponded to their respective treatment replicate groups in the hatcher units. Brooding and rearing conditions. The lighting schedule was 22 h light / 2 h darkness at 32-30°C at the first day to 1st wk. Pellet diets and fresh water offered ad libitum. all chicks in all groups served the same diets: starter (0-11d) [3100 kcal/kg metabolic energy (ME), 23% crud protein (CP), 4.0% crud fiber (CF)], the grower (12-25d) [2900 kcal/kg ME, 20.5% CP, 4.15% CF] and the finisher (26-35d) [2920 kcal/kg ME, 21.3% CP, 4.45% CF].

Bone measurements

Three chicks/birds from each replicate of treatment groups were randomly selected on d

0 and 35 posthatch they weighed and measured for body length (**BL**) as described by (16) before being euthanized and necropsied. Dried tibias were subjected to breaking strength analysis using the method described by Shim (19). Left tibia bones from birds were subsequently weighed and were dried in a forced-air oven for 24 h at 105°C and weighed. All tibias for chicks at both ages were ether extracted for 12 h extraction before ash obtaining in a muffle furnace at 480°C for 16 h. The mineral contents (Ca and P) of the tibia bone samples were determined by HPLC.

Blood minerals and vitamins determination

blood was sampled from jugular vein of 0 d chicks and the brachial vein of 35 d broiler, for determining blood plasma concentration of total calcium and inorganic phosphorus using kits by spectrophotometer. vitamin D $(1,25(OH)_2D_3)$ using kit by HPLC, hormones [parathyroid (PTH) and calcitonin (CT) concentrations were determined using kits by Radio immunoassay- RIA, all kits for chickens produced by Bioscience UK.

Statistical analysis

All data were analyzed for normal distribution using the normal option procedure of SAS software (18). Data were analyzed as a completely randomized design by the GLM procedure of SAS software. Statistical differences were established using a Duncan's Multiple Range Test at the level of $P \le 0.05$ (4).

RESULTS AND DISCUSSIONS

Table 1. The effect of in ovo injection with calcium, Cole vit D₃ and their combination or mix on hatchability, the results presented significantly ($P \le 0.05$) higher in hatchability of fertile egg (%), hatchability of total egg (%) and hatch window (h) in the groups of in ovo injection with Ca, Cole vit D₃ and their mix compared with the control (without in ovo injection) and the group of DW (desterilized water in ovo injection). Broiler breeder fertile eggs that were in ovo injected with Ca, Cole vit D_3 and their mix had significantly (P \leq 0.05) higher body weight, width of breast and length of shank length of chick at 0 posthatch compared with the control group and eggs that were injected with SDW, also length of broiler was higher in the groups Cole vit D_3 and their mix compared with the other groups of the study. As well as at age 35 d posthatch the groups that were in ovo injected with Ca, Cole vit D_3 and their mix had significantly (P \leq 0.05) higher body weight, length of broiler, width of breast and length of shank length compared with the control group and eggs that were injected with SDW (Table 2)

Traits	Treatments					
	С	SDW	Ca	Cole Vit D ₃	Mix	
Hatchability of fertile eggs (%)	84.7 °	82.9 °	86.0 ^b	87.9 ^{ab}	89.8 ^a	3.11
Hatchability of total eggs (%)	74.3 °	73.6 °	79.5 ^b	81.8 ^{ab}	83.6 ^a	2.74
Hatch window (h)	21:00 ^c	21:24 °	16:10 ^b	15:47 ^{ab}	14:05 ^a	0.93
Total dead embryos (1-21) d (%)	18.30 ^a	19.55 ^a	16.27 ^{ab}	14.65 ^b	13.15 ^b	1.02
Culled chicks (%)	7.40 ^a	6.85 ^a	4.23 ^b	3.55 ^b	3.25 ^b	0.385

Table 1. Effect of In ovo Injection with Calcium and Cole vit D₃ on some hatching traits

C: control (without in ovo injection); SDW: 100 μ l sterilized distill water in ovo injection; Ca: 0.8 mg calcium dissolved in 100 μ l SDW; Cole Vit D₃: 0.8 mg 25--OHD₃ dissolved in 100 μ l SDW; Mix: 0.8 mg Ca + 0.8 mg Cole vit D₃ dissolved in 100 μ l SDW.

Table 2. Effect of In ovo Injection with Calcium and Cole vit D3 on body weight and length,width of breast and length of shank at ages 0 and 35 days of posthatch

	Treatments						
Age Traits	С	C SDW		Cole Vit D ₃	Ν	ſix	
<u>0 day</u>	43.27 ^b	43.10 ^b	44.08 ^a	44.30 ^a	44.85 ^a	1.67	
Body weight (g)							
Length of chick (cm)	19.81 ^b	19.75 ^b	19.80 ^b	20.83 ^a	20.97 ^a	1.20	
Width of breast (cm)	11.45 ^b	11.56 ^b	12.55 ^a	12.70 ^a	12.67 ^a	0.81	
Length of shank (cm)	2.25 ^b	2.10 ^b	2.45 ^a	2.52 ^a	2.58 ^a	0.25	
<u>35 day</u>	2529 °	2546 °	2689 ^b	2710 ^{ab}	2780 ^a	141	
Body weight (g)							
Length of broiler (cm)	43.75 °	42.59 °	48.90 ^b	50.77 ^{ab}	52.91 ^a	1.92	
Width of breast (cm)	22.13 ^b	21.96 ^b	23.82 ^a	24.10 ^a	24.38 ^a	0.89	
Length of shank (cm)	81.33 °	80.42 °	84.10 ^b	85.93 ^{ab}	87.35 ^a	2.11	

C: control (without in ovo injection); SDW: 100 µl sterilized distill water in ovo injection; Ca: 0.8 mg calcium dissolved in 100 µl SDW; Cole Vit D₃: 0.8 mg 25--OHD₃ dissolved in 100 µl SDW; Mix: 0.8 mg Ca + 0.8 mg Cole vit D₃ dissolved in 100µl SDW.

Table 3. as to tibia length had significantly ($P \le 0.05$) higher in the groups of in ovo injected with Cole vit D_3 and their mix at both 0 and 35 d posthatch, so breaking strength had increased in Ca, Cole vit D_3 and their mix groups at both ages compared with the control group and eggs that were injected with SDW.

However had seen non-significant differences among all the groups in tibia width.

Table 4. shows that tibia bone affected by in ovo injection with Ca, Cole vit D_3 and their mix, it had significantly (P ≤ 0.05) raised in Ca and P concentrations, also in ash percentage at 0 and 35 d posthatch, while non-significant differences among all the groups in Ca:P ratio.

Table 3. Effect of In ovo Injection with Calcium and Cole vit D3 in different age on tibiaweight and length %, and strength at ages 0 and 35 d of posthatch

	Treatments					
Age Traits	C	SDV	V Ca	Cole vit	D ₃ Mix	
<u>0 day</u>	2.65 ^b	2.62 ^b	2.88 ^{ab}	2.92 ^a	2.97 ^a	0.281
Tibia length (cm)						
Tibia width (cm)	1.78 ^a	1.76 ^a	1.85 ^a	1.89 ^a	1.95 ^a	0.122
Breaking strength (kg/cm ²)	0.890 ^b	0.847 ^b	1.303 ^a	1.308 ^a	1.325 ^a	0.301
35 da <u>y</u>	8.89 ^b	8.24 ^b	9.48 ^{ab}	9.66 ^a	9.73 ^a	0.783
Tibia length (cm)						
Tibia width (cm)	8.13 ^a	8.02 ^a	8.09 ^a	8.13 ^a	8.10 ^a	0.690
Breaking strength (kg/cm ²)	8.205 ^b	8,100 ^b	10.067 ^a	10.843 ^a	10.892 ^a	0.755

C: control (without in ovo injection); SDW: 100 μ l sterilized distill water in ovo injection; Ca: 0.8 mg calcium dissolved in 100 μ l SDW; Cole Vit D₃: 0.8 mg 25--OHD₃ dissolved in 100 μ l SDW; Mix: 0.8 mg Ca + 0.8 mg Cole vit D₃ dissolved in 100 μ l SDW. The same superscripts within rows means non-significant. ^{a-c} Means within rows with different superscripts differ significantly at (P≤ 0.05).

 Table 4. Effect of In ovo Injection with Calcium and Cole vit D3 on tibia bone chemical analysis at ages 0 and 35 d of posthatch.

	Treatments					
Age Traits	С	SDW	Ca	Cole Vit D ₃	Mix	
0 day	10.33 ^b	10.09 ^b	12.13 ^a	12.42 ^a	12.53 ^a	0.726
Ca (g/g tibia ash)						
P (g/g tibia ash)	6.47 ^c	6.55 ^c	7.69 ^b	8.03 ^a	8.11 ^a	0.703
Ca: p ratio	1.597 ^a	1.540 ^a	1.577 ^a	1.547 ^a	1.545 ^a	0.077
Ash %	37.75 ^c	37.16 °	41.29 ^b	40.88 ^b	44.96 ^a	1.402
35 day	11.54 ^b	11.35 ^b	15.00 ^a	14.72 ^a	15.29 ^a	0.810
Ca (g/g tibia ash)						
P (g/g tibia ash)	6.92 ^b	6.89 ^b	8.39 ^a	8.55 ^a	8.83 ^a	0.674
Ca: p ratio	1.668 ^a	1.647 ^a	1.789 ^a	1.722 ^a	1.732 ^a	0.083
Ash %	34.65 ^b	34.02 ^b	39.48 ^a	40.7 ^a	40.10 ^a	1.633

Table 5 clarify posthatch blood minerals, vitamin D_3 and hormones were impacted with the in ovo injection with Ca and Cole vit D_3 . The groups of Ca, Cole vit D_3 and their mix were significantly (P \leq 0.05) higher in Ca and P minerals concentrations, vit D_3 [1,25(HO)₂D₃] and parathyroid hormone (PTH) at 0 and 35 d posthatch. However, calcitonin hormone was significantly ($P \le 0.05$) lower in the groups of Ca, Cole vit D₃ and their mix at 0 and 35 d posthatch, also and Ca: P ratio was decreased in the same groups at 0 posthatch. While, there wasn't any significant differences among the non-injected and injected groups in Ca: P ratio at 35 d posthatch

		p	osthatch				
	Treatments						
Age Traits	С	SDW	Ca	Cole Vit D ₃	Mix		
<u>0 day</u>							
<u>Minerals</u>							
$Ca^{+2}(mg/dL)$	7.08 ^b	7.35 ^b	8.07 ^a	8.19 ^a	8.31 ^a	0.633	
P (mg/dL)	4.36 ^b	4.26 ^b	5.61 ^a	5.45 ^a	5.68 ^a	0.425	
Ca: P ratio	1.624 ^b	1.725 ^a	1.439 °	1.503 ^c	1.463 °	0.106	
<u>Vitamine</u>							
$\overline{\mathbf{D}_3}$ (ng/ml)	125 °	89.5 ^d	162 ^b	217 ^a	208 ^a	17.20	
Hormones							
CT (ng/ml)	1.074 ^a	1.127 ^a	0.895 ^b	0.781 ^c	0.759 °	0.093	
PTH (ng/ml)	1.18 °	1.03 ^c	2.31 ^b	2.34 ^b	2.77 ^a	0.204	
<u>35 day</u>							
Minerals	8.10 °	7.82 ^c	10.18 ^b	10.35 ^{ab}	10.93 ^a	0.852	
$\overline{\text{Ca}^{+2}}$ (mg/dL)							
P (mg/dL)	5.11 ^b	5.02 ^b	6.23 ^a	6.57 ^a	6.89 ^a	0.441	
Ca: P ratio	1.585 ^a	1.558 ^a	1.634 ^a	1.575 ^a	1.586 ^a	0.102	
<u>Vitamine</u>							
$\overline{\mathbf{D}_3}$ (pg/ml)	55.4 °	37.2 ^d	74.2 ^b	121.0 ^a	124.9 ^a	3.69	
Hormones							
CT (ng/ml)	2.35 ^a	2.47 ^a	1.16 ^b	1.02 ^b	0.91 ^b	0.082	
PTH (ng/ml)	1.95 °	1.63 ^c	2.79 ^b	3.18 ^{ab}	3.47 ^a	0.205	

Table 5. Effect of In ovo Injection with Calcium and Cole Vit D₃ on blood minerals (Ca & P), vit D₃ [1,25(OH)₂D₃] and hormones [calcitonin (CT) & parathyroid (PTH)] at 0 and 35 nosthatch

C: control (without in ovo injection); SDW: 100 μ l sterilized distill water in ovo injection; Ca: 0.8 mg calcium dissolved in 100 μ l SDW; Cole Vit D₃: 0.8 mg 25--OHD₃ dissolved in 100 μ l SDW; Mix: 0.8 mg Ca + 0.8 mg Cole vit D₃ dissolved in 100 μ l SDW. The same superscripts within rows means non-significant. ^{a-c} Means within rows with different superscripts differ significantly at (P≤ 0.05).

The hatchability percentage and body weight of hatched chicks was increased in groups in ovo injected with calcium, phosphorus, and vitamin D complex (6). The functional form of vitamin D in biology is 1,25-(OH)₂D₃, the production is very carefully regulated by parathyroid hormone (PTH) in response to serum calcium and phosphate (PO4⁻³) concentra-tions (5). 1,25-(OH)₂D₃ is a critical factor in the maintenance of sufficient maternal calcium for transport to the embryo and may play a role in normal skeletal development of the neonate (10). Two hormones, calcitonin and parathyroid function in a delicate relationship with $1,25-(OH)_2D_3$ to control blood calcium and phosphorus levels (7 & 15). Production rate of 1,25-(OH)2D is under physiological control as well as dietary control. Calcitonin, contrary to the other two, regulates high serum calcium levels by (10) depressing gut absorption, (17) halting bone demineralization. and (10)depressing reabsorption in the kidney. Vitamin D elevates plasma calcium and phosphorus by stimulating specific ion pump mechanisms in the intestine, bone and kidney. These three sources of calcium and phosphorus provide reservoirs

that enable vit D to elevate calcium and phosphorus in blood to levels that are necessary for normal bone mineralization and for other functions ascribed to calcium. In the target tissue, the hormone enters the cell and binds to a cytosolic receptor or a nuclear 1,25-(OH)2D regulates receptor. gene expression through its binding to tissuespecific receptors and subsequent interaction between the bound receptor and the DNA (3). The results reported confirms the importance and essentiality of vit D3 and its active metabolites for normal embryonic development by providing the necessary Ca and P needed for normal skeletal development. calcium used for skeleton-genesis comes primarily from the shell, whereas phosphorus derived mainly from the volk. is Ca mobilization from the shell commences later in incubation and proceeds at a rapid rate resulting in well mineralized embryos. In ovo treatments with vit D3 or metabolites increased the rate of calcium mobilization from the egg shell to the embryo and increased concentration in all embryonic calcium compartments (13). Feeding and in ovo injection of 1a-OHD3 increased breaking strength, calcium and phosphorous percentage in tibia bone (9). Research has shown that a 1 cm chick length advantage at day of hatch can result in 264 grams more body weight with 45 grams more breast meat yield at 38 days of age, (16). This, and the fact that an optimally developed chick will have a better feed conversion rate.

In a previous study in which Cole Vit D3 and its mix with calcium were used as in ovo injection, it was shown that at a 0.8 mg able to embryonic improve the development, hatchability, body weight at 0 and 35 posthatch of broilers, also increased bone characteristics and breaking strength, thus improved Ca and P in bone and blood plasma also vit D₃, calcitonin and parathyroid concentrations, all hormone these improvement positively reflected in better formation of skeleton and body formation well.

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REFERENCES

1. Bello, A., W. Zhai, P. D. Gerard, and E. D. Peebles. 2013. Effects of the commercial in ovo injection of 25-hydroxycholecalciferol on the hatchability and hatching chick quality of broilers. Poult. Sci. 92:2551–2559

2. Bello, A., W. Zhai, P. D. Gerard, and E. D. Peebles. 2014. Effects of the commercial in ovo injection of 25-hydroxycholecalciferol on broiler post-hatch performance and carcass characteristics. Poultry Sci. 93:155–162

3. Collins, E.D., and A.W. Norman. 1991. Vitamin D. In "Handbook of Vitamins" (L.J. Machlin, ed.) Marcel Dekker Inc., New York. pp;59.

4. Duncan, D.B. 1955. Multiple range and Multiple F-test. Biometrics. 11:1-42

5. DeLuca, H.F. 2008. Evolution of our understanding of vitamin D. Nutr. Rev. 66 (Suppl 2): 573-587

6- Ebrahimi, H.; F. Shariatmadari & M.A. Karimi Torshizi. 2016. Dietary supplementation and in ovo injection of 1α -OHD3 in a low-calcium and low-phosphorous diets for broilers. Journal of Applied Animal Research, 44(1), 113-117

7. Engstrom, G.W., and E.T. Littledike. 1986. Vitamin D metabolism in the pig. In: Swine in Biomedical Research, (M.E. Tumbleson, ed.). Plenum Press, New York pp:1091-1112..

8. Geleijnse, J. 2011. Vitamin D and the prevention of hypertension and cardio-vascular diseases: a review of the current evidence. American journal of hypertension, 24 (3) 253-262

9. Ghobadi, N.; and Hamid Reza Hemati Matin, 2015. Response of Broiler Chicks to in ovo Injection of Calcium, Phosphorus, and Vitamin D Complex (CaDPhos). Global Journal of Animal Scientific Research. 3 (2), 536-543

10. Holick, MF. and M. Garabedian. 2006. Vitamin D: photobiology, metabolism, mechanism of action, and clinical applications. In Primer on the metabolic bone diseases and disorders of mineral metabolism. 6th edition. Edited by: Favus MJ. Washington, DC: American Society for Bone and Mineral Research; 6: 129-137

11. Khan, S.H. Shahid, R. Mian, A.A.R. Sardar and M.A. Anjum. 2010. Effect of the level of cholecalciferol supplementation of broiler diets on the performance and tibial dyschondroplasia. Journal of Animal Physiology and Animal Nutrition, 94: 584-593

12. Lester, G.E. 1986. Cholecalciferol and placental calcium transport. Fed. Proc. Fed. Am. Soc. Exp. Biol. 45:2524

13. Matthiesen, C.F., D. Blache, P.D. Thomsen, N.E. Hansen, and A.H. Tauson. 2010. Effect of late gestation low protein supply to mink (Mustela vison) dams on reproductive performance and metabolism of dam and offspring. Arch. Anim. Nutr, 64: 56– 76.

14. Mansour, S. Doaa, Y.A. El-Senosi, M. I. Mohamed, M.M. Amer and M.A. Elaroussi1. 2017. Effects of injecting vitamin D3 or an active metabolite in-ovo on chick embryonic development and calcium homeostasis. World J. of Pharmacy and Pharmaceutical Sci., 6 (12):1454-1467

15. McDowell, L. R. 2000. Vitamins in Animal and Human Nutrition. Iowa State University Press, Ames, IA. pp: 793 16. Molenaar, R., I. A. M. Reijrink, R. Meijerhof, and H. van den Brand. 2008. Relationship between hatchling length and weight on later productive performance in broilers. World's Poult. Sci. J. 64:599–604 17. Prentice A, GR. Goldberg and I. Schoenmakers. 2008. Vitamin D across the lifecycle: physiology and biomarkers. Am J Clin Nutr. 88:500S–506S

18. SAS, Statistical analysis system. 2005. User's Guide for Personal Computer. Release 8.2 SAS Institute Inc. Cary. NC, USA

19. Shim, M. Y., A. B. Karnuah, A. D. Mitchell, N. B. Anthony, G. M. Pesti, and S.

E. Aggrey. 2012. The effects of growth rate on leg morphology and tibia breaking strength, mineral density, mineral content, and bone ash in broilers. Poultry Sci. 91:1790–1795

20. Veldurthy, Vaishali; Ran Wei; Leyla Oz; Puneet Dhawan; Yong Heui Jeon and Sylvia Christako. Vitamin D, calcium homeostasis and aging. 2016.Bone Research J. 4, 16041; doi:10.1038/boneres. 41

21. Zhang, R. and D. P. Naughton. 2010. Vitamin D in health and disease: Current perspectives. Nutrition Journal, 9:65.