

TRADITIONAL AND MOLECULAR STUDY OF CRYPTOSOPORIDIUM SPP. IN DOMESTIC DOGS IN BAGHDAD CITY,IRAQ

A. A. Faraj*

Assist. Prof.

*Department of Parasitology, College of Veterinary Medicine, University of Baghdad , Iraq
aazhar888@yahoo.com

ABSTRACT

The aim of this study to investigate the prevalence of *Cryptosporidium* spp. in domestic dogs in Baghdad city, by using microscopic examination (flotation and staining) and molecular techniques. The results revealed that the rate of *Cryptosporidium* infection was 15% by microscopic examination and 28.6% using PCR, our study showed that the infection rate was 8.5% in males and 21.42% in females by microscopic examination, while the infection rate by using PCR in males and females was 21.42%, 35.71% respectively. The percentage of infection in the age groups <1 year and >1 year was 20%, 10% respectively by microscopic examination, while by using molecular assay the rate of infection in the age groups <1 year was 40% and 17.14% in >1 year. This is the first study in Iraq to detect *Cryptosporidium* spp. in domestic dogs using molecular technique.

Key words: Cryptosporidiosis, microscopic examination, PCR, phylogenetic tree, dogs.

فرج

مجلة العلوم الزراعية العراقية - 2019: 50(4): 1094-1099

الدراسة التقليدية والجزيئية لطيفلي البوغ الخبيث في الكلاب المنزلية في بغداد , العراق

ازهار علي فرج*

استاذ مساعد

*فرع الطفيليات - كلية الطب البيطري - جامعة بغداد - العراق

المستخلص

هدفت الدراسة الحالية للتحري عن انتشار الابواغ الخبيثة في الكلاب المنزلية في مدينة بغداد ، وذلك باستخدام الفحص المجهرى (التطويف والتصبيغ) والتقنيات الجزيئية. أظهرت النتائج أن معدل الإصابة بطيفلي البوغ الخبيث كان 15% بالفحص المجهرى و 28.6% باستخدام تفاعل تسلسل البلمرة ، اظهرت نتائج الفحص المجهرى معدل الإصابة في الذكور 8.5% و 21.42% في الإناث ، في حين كان معدل الإصابة باستخدام تفاعل تسلسل البلمرة في الذكور والإناث 21.42% ، 35.71% على التوالي. اظهرت نسبة الإصابة بالفحص المجهرى في الفئات العمرية < 1 سنة كانت 20% و < 1 سنة 10% ، بينما كان معدل الإصابة باستخدام الفحص الجزيئي في الفئات العمرية < 1 سنة 40% و > 1 سنة 17.14% في الاعمار < 1 سنة. تعد هذه الدراسة الاولى في العراق للكشف عن الابواغ الخبيثة في الكلاب المنزلية باستخدام التقنيات الجزيئية.

الكلمات المفتاحية: الابواغ الخبيثة ، الفحص المجهرى ، تفاعل تسلسل البلمرة، الشجرة الوراثية، الكلاب .

INTRODUCTION

Cryptosporidiosis is one of the important parasitic diseases, caused by *Cryptosporidium*. It infects wide range of vertebrates, including, humans, livestock, companion, and wildlife animals. Infection is occur via the fecal–oral route after the ingestion of contaminated food or water with oocysts (17, 3), Diarrhea is the most important clinical signs of disease humans and animals (14). and dogs also act as reservoirs for a large number of parasitic zoonoses, including cryptosporidiosis (20,11). Diagnoses *Cryptosporidium* of infection in fecal samples of dogs is based by identifying the oocysts using microscopic techniques, identifying the parasite antigen by ELISA, or by pathogen DNA using PCR and the basic interest of last one compared with other assays is that the amplification products can be analyzed by sequencing analysis to define the species of the parasite (13,2). There is little information on the *Cryptosporidium* spp. present in infected dogs in Iraq. This study was

conducted to determine the prevalence and molecular diagnosis *Cryptosporidium* species in domestic dogs in Baghdad city, Iraq.

MATERIALS AND METHODES

Detection of *Cryptosporidium* oocysts

One hundred and forty fecal samples (10 g) from domestic dogs, of different sexes and ages referred that a small animal private clinics in Baghdad, Iraq. During the period from 1/1/2018 to 31/5/2018, analyzed by flotation methods and modified Ziehl-Neelsen staining (4,6). Finally the samples were saved at 2.5% potassium dichromate (K₂Cr₂O₇) (12), and stored at (-20 C °) until DNA was extracted.

DNA Extraction and PCR

DNA extraction procedure was performed using DNA stool kit (ABM, Canada) based on the manufacturer’s guideline. By using for 18S ribosomal gene *Cryptosporidium*. design this primer by the (NCBI)gene-Bank data according to program(Primer3 plus online) (Table1,Figure 1)

Table 1. Oligonucleotide primers used to amplify and sequence parasite 18S rRNA genes

Primers	Primer sequence (5' to 3')		Product size (bp)
<i>18S rRNA Gene</i>	F	GAGGTAGTGACAAGAAATAACAATACACC	300
	R	CTGCTTTAAGCACTCTAATTTTCTCAAAG	

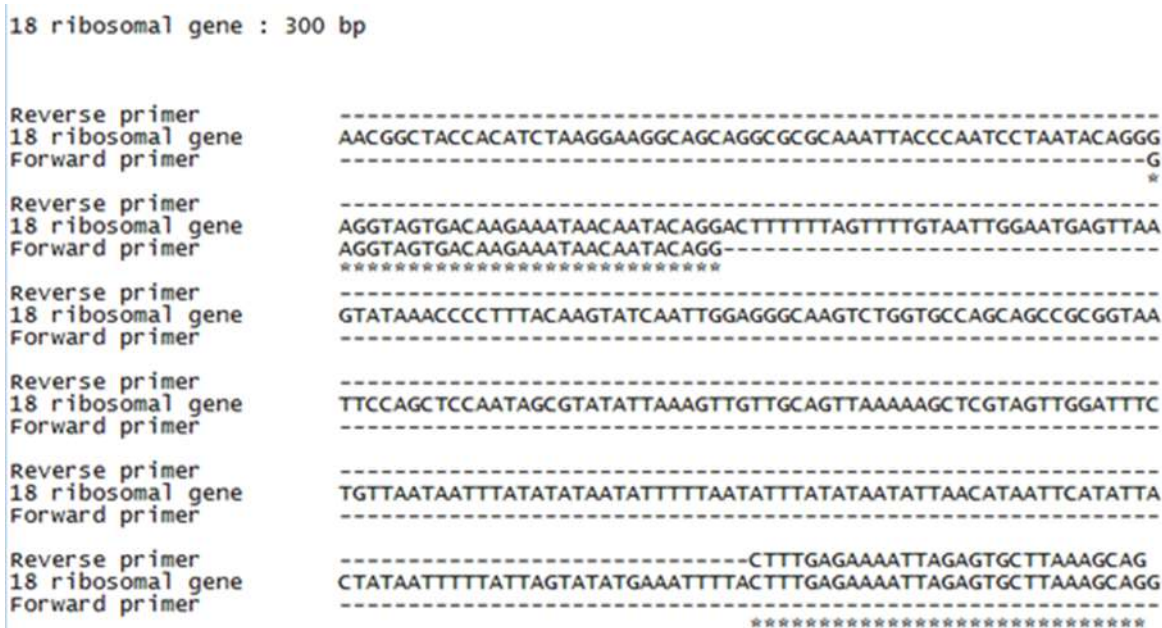


Figure 1. Amplification and sequence analysis of 18s(300bp) of the rRNA gene

Statistical analysis: Statistically analyzed of the data was done by using SAS software(Statistical Analysis System –Version 9.1) (15).

RESULTS AND DISCUSSION

The results showed the infection rate of *Cryptosporidium* spp. of domestic dogs by microscopic examination revealed 21(15.0%) positive sample and 119(85.0%) negative sample and PCR positive samples 40(28.6%)

and negative samples 100 (71.4%). Also, PCR was done and showed 300 bp band which confirmed only 40 samples as *Cryptosporidium* spp., the specificity was 94.00 and the low sensitivity was(37.50) which means that not useful for con firm the infection also the value of kappa (0.367) reflects a fair agreement between two test (Table 2, Figure 2,3).

Table 2. Infection rate of *Cryptosporidium* spp. by microscopic and PCR technique.

Observer A	PCR		
Observer B	Microscopic		
Observer B	Observer A		
	0	1	
0	94	25	119 (85.0%)
1	6	15	21 (15.0%)
	100 (71.4%)	40 (28.6%)	140
Weighted Kappa ^a	0.367		
Standard error	0.087		
95% CI	0.197 to 0.538		
Sensitivity		37.50	
Specificity		94.00	

- Poor agreement = Less than 0.20
- Fair agreement = 0.20 to 0.40
- Moderate agreement = 0.40 to 0.60
- Good agreement = 0.60 to 0.80
- Very good agreement = 0.80 to 1.00

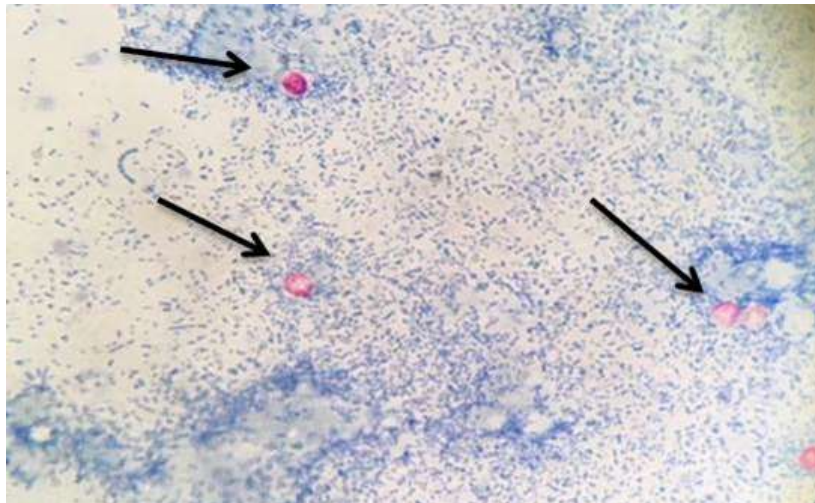


Figure 2. *Cryptosporidium* oocysts stained by Modified Ziehl Nielsen stain (X100).

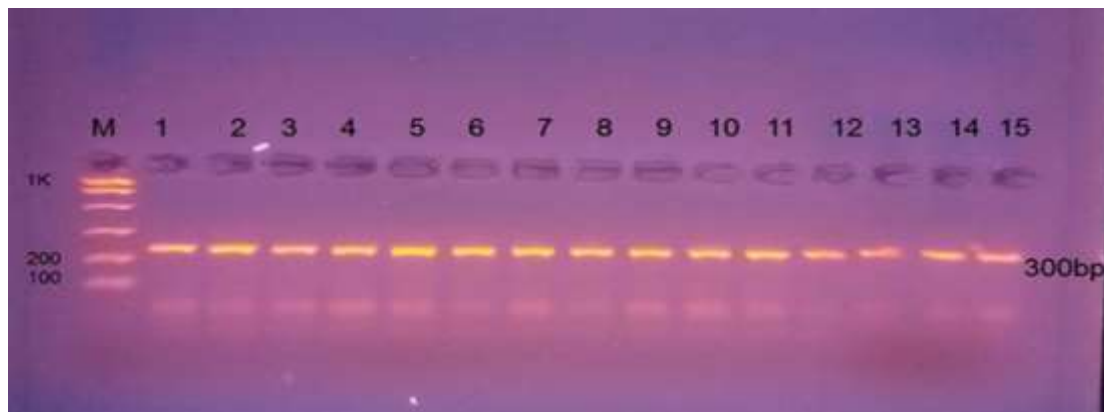


Figure3. Gel electrophoresis of PCR product of 18S rRNA (300bp), for *Cryptosporidium parvum* using 2% agarose gel at 60volt for 1 hour. Lane 1- 15: PCR product positive for 18S rRNA genes

Table 3 shows that the infection rate in males was 8.5% and in females 21.42% by microscopic examination, while the infection rate by using PCR in males and females was 21.42%, 35.71% respectively. The association between males and females was not significant

Table 3. Prevalence of *Cryptosporidium spp.* in domestic dogs by microscopic examination and PCR according to sex

Sex	No. of fecal samples examined	Microscopic Examination No. infected (%)	PCR No. infected (%)
Male	70	6(8.5)	15 (21.42)
Female	70	15(21.42)	25(35.71)
Total	140	21 (15.0)	40(28.6)
Chi square value			3.50
P			0.06

Results revealed that the percentage of infection in the age group <1 year and >1 year was 20%, 10% respectively by microscopically examination, while by using

the molecular assay the rate of infection in the age group <1 year was 40% and 17.14% in age group >1 year with statistical differences at(<0.01) (Table ,4).

Table 4. Prevalence of *Cryptosporidium spp.* in domestic dogs by microscopic examination and PCR according to age

Age year	No. of fecal Samples examined	Microscopic Examination No. infected (%)	PCR No. infected (%)
<1	70	14(20)	28 (40)
>1	70	7(10)	12(17.14)
Total	140	21 (15.0)	40(28.6)
Chi square value			8.96
P			<0.01

Phylogenetic Analysis

The BLASTIN analysis of the 18S rRNA gene to the our sample that isolated and sequences with the recorded strains that described in the databases using the MEGA 6.06. program. The evolutionary relationship was designed using molecular sequences in sequences in determining cluster due to variation in 18SrRNA monochromatic sequence. It can be stretched between 0.1.when analysis and compare with the NCBI-Gen bank *Cryptosporidium parvum* isolates that based on offered information with local isolate of *Cryptosporidium parvum* that

recorded under accession number (MH716021.1) (Figure 4). Figure 4: Show the phylogenetic tree inferred the degree of relatedness between 18SrRNA sequence deposited in the international oligonucleotide primers used to amplify and sequence parasite 18S rRNA genes. Bank sequence database(NCBI) of *Cryptosporidium parvum* as isolated from infected dogs under accession number (MH716021.1), sample based on a partial sequence of 18S rRNA Gene GenBank *Cryptosporidium* is a coccidian, intracellular protozoan parasite pathogen, which causes diarrheal illness of animals and

humans (18). The present study showed that the rate of infection *Cryptosporidium* spp. oocysts in dogs fecal was 30% and 57.40% by using staining and molecular methods, respectively. This rate is a high than the results of some previous studies, Bahrami *et al.* (1) reported 7.04% infection in stray dogs of Ilam using the Ziehl Neelsen staining method. in Bahía Blanca of Argentina Sala *et al.* (9) was recorded 14.7% by using by ZiehlNeelsen staining. . in Thailand Koompapong *et al.* (8) who recorded 2.1% by using PCR and in Japan Yoshiuchi *et al.* (20) who recorded 3.9% by PCR.. Results revealed that the microscopic test has a low sensitivity %37. This is attributed to the experience of the examiner to observation of oocysts in the feces specimens , also microscopic examination does not distinguish between different species of parasites.; thus, molecular assay like PCR have been used with high sensitivity, specificity, and rapidly features are capable of differentiating between species and genotypes in different samples, although they are expensive. Previously, genotyping of *Cryptosporidium* spp. has been done successfully by PCR method according to (*I8S rRNA*) gene (12, 10 ,19). Our results indicate that the age may affect the rates of infection through the significant high prevalence of *Cryptosporidium* in the young dogs than in adult dogs using both methods. Similar research were reported in dogs (5, 20). This could be attributed to lower resistance in the young compared to older animals. In otherwise, no statistical differences relationship was between the infection of *Cryptosporidium* and the sex of the dogs. Agree with two studies conducted in China and Brazil recorded similar findings (16,7). This refers that sex is not a major affecting Cryptosporidiosis infection in dogs. In the present study, a first phylogenetic analysis of *C. parvum* isolates in Iraq from infected domestic dogs, based on the *I8S rRNA* gene sequence showed that there were relationships between *C.parvum* local isolate (MH716021.1) and other global isolates the local isolates were closely related to France isolate under accession number EF158462.1 with nucleotide sequence identity 91%. Furthermore, showed gradual similarities with

the other isolates from the high grade to the lower indicated on identity between (91-92%).

REFERENCES

1. Bahrami, A.; A. Doosti ; H. Nahravanian ; A.M. Noorian and S.A. Asbchin. 2011. Epidemiological survey of gastrointestinal parasites in stray dogs and cats. Australian. J. Basic. App. Sci. 5:32-35
2. Bouzid, M.; P.R. Hunter and R.M.T. Chalmers. 2013. "Cryptosporidium pathogenicity and virulence," Clin. Microbiol. Rev., 26: 115–134.
3. Feng, Y. and L. Xiao. 2011. Zoonotic potential and molecular epidemiology of *Giardia* species and giardiasis. Clin. Microbiol. Rev. 24: 110-140.
4. Fujino, T.; T. Matsuo; M. Okada and T. Matsui. 2006. Detection of a small number of *Cryptosporidium parvum* oocysts by sugar floatation and sugar centrifugation methods. J. Vet. Med. Sci., 68:1191–1193
5. Gil, H.; L. Cano ; A. de Lucio; B. Bailo ; M.H. de Mingo and G.A. Cardona. 2017. Detection and molecular diversity of *Giardia duodenalis* and *Cryptosporidium* spp. in sheltered dogs and cats in Northern Spain. Infect Genet Evol., 50:62–9
6. Henriksen, S.A. and J.F.L. Pohlenz. 1981. Staining of *Cryptosporidia* Acta, Vet. Scand., 22: 594-596
7. Huber, F. and T.C.B. Bomfim. 2005. Gomes RS: Comparison between natural infection by *Cryptosporidium* sp. *Giardia* sp. in dogs in two living situations in the West Zone of the municipality of Rio de Janeiro. Vet. Parasitol., 130: 69–72
8. Koompapong, K. ; H. Mori ; N. Thammasonthijarern; R. Prasertbun ; A.R. Pintong; and S. Popruk. 2014. Molecular identification of *Cryptosporidium* spp. in seagulls, pigeons, dogs, and cats in Thailand. Parasite., 21:52-53
9. La Sala, L.F.; A. Leiboff ; J.M. Burgos and S.R. Costamagna. 2015. Spatial distribution of canine zoonotic enteroparasites in Bahía Blanca, Argentina. Rev. Argent. Microbiol., 47:17–24
10. Mahami, O.M.; E. Fallah and M. Ahmadi. 2014. Molecular and parasitological study of *Cryptosporidium* isolates from cattle in

- Ilam, west of Iran. Iranian. J. Parasitol., 9: 435–440.
11. Molloy, S.F.; H.V. Smith; P. Kirwan; R.A. Nichols ; S.O. Asaolu; L. Connelly and C.V. Holland. 2010. Identification of a high diversity of *Cryptosporidium* species genotypes and subtypes in a pediatric population in Nigeria. Am. J. Trop. Med. Hyg., 82 : 608-613.
12. Pirestani, M.; J. Sadraei, ; A. Dalimi.; M. Zavvar and H. Vaeznia. 2008. "Molecular characterization of *Cryptosporidium* isolates from human and bovine using 18s rRNA gene in Shahriar county of Tehran, Iran," Parasitol. Res., 103: 467–472.
13. Plutzer, J. and P. Karanis. 2009. Genetic polymorphism in *Cryptosporidium* species: an update. Vet. Parasitol., 165: 187–199.
14. Ryan, U.; R. Fayer and L. Xiao. 2014. *Cryptosporidium* species in humans and animals: current understanding and research needs. Parasitol., 141: 1667–1685
15. SAS. 2010/STAT Users Guide for Personal Computer. Release 9.1. SAS Institute, Inc., Cary, N.C., USA
16. Wang, J.; P. Li ; X. Xue ; X. Chen; S. Jiang ; X. Qiu and G.L. Yin. 2008. Investigation on the infection situation of *Cryptosporidium* in dogs in Hefei city (in Chinese). Chin. J. Vet. Parasitol., 16: 20–23
17. Xiao, L. 2010. Molecular epidemiology of cryptosporidiosis: An update. Experimental Parasitol., 124(1): 80-89
18. Xiao, L. and Y. Feng. 2008. Zoonotic cryptosporidiosis. FEMS Immunol. Med. Microbiol., 52: 309-323
19. Xiao, L.; U.M. Ryan and T.K. Graczyk . 2014. Genetic diversity of *Cryptosporidium* spp. in captive reptiles, Applied and Environmental Microbiology., 70: 891–899.
20. Yoshiuchi, R.; M. Matsubayashi; I. Kimata; M. Furuya; H. Tani and K. Sasai, 2010. Survey and molecular characterization of *Cryptosporidium* and *Giardia* spp. in owned companion animal, dogs and cats, in Japan. Vet. Parasitol., 174: 313–6
21. Yoshiuchi, R.; M. Matsubayashi; I. Kimata; M. Furuya; H. Tani and K. Sasai. 2010. Survey and molecular characterization of *Cryptosporidium* and *Giardia* spp. in owned companion animal, dogs and cats, in Japan. Vet. Parasitol., 174: 313–6.