NUTRITIVE VALUE OF BARLEY SILAGE (Hordeum vulgare L.) WITH DIFFERENT LEVELS OF SACCHAROMYCES CEREVISIAE AT DIFFERENT ENSILING LENGTHS

J. L. Contreras^{1*} R. G. Quichca¹ A. G. Cordero² Y. C. Rojas¹ J. Curasma¹ U. Dickhoefer³ J. Castro.³

¹Laboratory of Animal Nutrition and Fedstuffs Evaluation (LUNEA), Faculty of Engineering Sciences Zootecnia. National University of Huancavelica, Huancavelica, Peru.

² Postgraduate school, Faculty of Engineering Sciences. National University of Huancavelica, Huancavelica, Peru.

³ Institute of Agricultural Sciences in the Tropics, Animal Nutrition and Rangeland Management in the Tropics and Subtropics, University of Hohenheim, Germany

*E-mail: jose.contreras@unh.edu.pe mob: +51948133886 https://orcid.org/0000-0003-4591-3885 ABSRTACT

The objective of this study was to evaluate the effects of yeast (Saccharomyces *cereviceae*) as additive for barley silage preparation at different ensiling times. Small scale silages were prepared from barley forage and four yeast levels were evaluated (0 (control), 5, 10 and 15 g/kg FM). Silos were opened at 6, 12 and 24 days. Three silage-replicates were prepared for each yeast level × ensiling time combination. Silage quality in terms of CP and NDF and ADF concentration appeared to improve when yeast was added at 5 and 10 g/kg, compared with the control, but the quality decreased again when 15 g yeast/kg FM were aded to the silage. However, gas production, metabolisable energy and organic matter digestibility linearly decreased by yeast addition. This indicates a likely negative effect of yeast over the fermentation process during fermentation of barley silage. No time effects were observed for most of the parameters with the exception of ether extract concentration. Based on the results of this study, yeast addition is detrimental to the nutritional quality of barley silage. Mechanisms for this still remain unknown, but an undesirable fermentation provoked by yeast addition might be an explanation for our findings.

Keywords: Digestibility, yeast, forage, neutral detergent fibre

كانتريروز وأخرون

مجلة العلوم الزراعية العراقية -2020 :51 (5):1356

القيمة الغذائية لسايلج الشعير (Hordeum valgare L) المعامل بمستويات مختلفة من الخميرة (SACCHAROMYCES CEREVISIAE) في سايلوات السايلج المختلفة كانتريروز كواجي كورديرو روجس كوراسما ديكو فور كاسترو

المستخلص

الهدف من الدراسة تقييم تأثير الخميرة (SACCHAROMYCES CEREVISIAE) كمضاف الى مواعيد السايلج المختلفة . ثم استعمال سايلوات سايلج صغيرة مع استعمال اربعة مستويات من الخميرة (صفر للمقارنة , 5 , 10 , 15 غم/ كغم المختلفة . ثم استعمال سايلوات بعد 6 و 12 و 24 يوم . استعمل ثلاثة مكررات لكل معاملة . كانت نوعية السايلج كغم MT . فتحت السيايلوات بعد 6 و 12 و 24 يوم . استعمل ثلاثة مكررات لكل معاملة . كانت نوعية السايلج المايلج معاملة . كانت نوعية السايلج معاملة . معاملة . كانت نوعية السايلج معاملة . كانت نوعية السايلج معاملة . كانت نوعية السايلج معاملة . معاملة المعارنية مع معاملة . معاملة

كلمات مفتاحية : قابلية الهضم, خميرة , علف , الالياف

*Received:22/9/2019, Accepted:13/12/2019

INTRODUCTION

Barley (Hordeum vulgare L.) silage production has been proposed as an option for conserved forage in areas where maize cultivation is limited by e.g. climate, soil conditions, irrigation or machinery (13). Barley ensiling, however, may be challenging due to the high moisture content at the ideal time of harvest and a low concentration of water soluble carbohydrates, compared with other grasses (9), making it imperative to investigate the potential of additives to enhance the ensilability of this material. Even though yeasts are generally recognized as undesirable microorganisms in the silage, because they are the primary initiators of aerobic spoilage during ensiling and opening (12), their ability to consume oxygen has made researchers to believe that they could have a positive effect on silage quality (10).particularly to ensure the onset of anaerobic fermentation. Furthermore, when directly fed to ruminants certain yeasts have shown to increase the population of cellulolytic bacteria and to enhance growth rate and milk production, while also excluding zoonotic pathogens from the intestinal tract (8), which would be an additional advantage of using yeasts as silage inoculant if those microorganisms are able to withstand the ensiling. Therefore, the objective of this study was to evaluate the changes that yeast early stages inoculant have over of conservation of barley silage on the nutritional characteristics of the forage.

MATERIALS AND METHODS

The experiment was conducted at the Faculty of Engineering Sciences of the University of Huancavelica (UNH). Barley forage was harvested from a local farms from the Antaccocha municipality in the Huancavelica district, in the Western chain of the Peruvian Andes. Barley forage was manually harvested at an age of 5 months and chopped at a particle length of 2 to 2.5 cm using a mechanical chopper for the ensiling. Small scale silos were prepared by storing 5 kg of fresh forage in polyethylene bags. Saccharomyces cerevisiae yeast (Active Dry Yeast, Fleischmann's, ACH Food Companies, Inc., Oakbrook Terrace, IL) was added to the fresh forage before ensiling at four levels: 0 (Control), 5, 10 and 15 g/kg

fresh matter (FM). The yeast was uniformly spread and thoroughly mixed by hand with the forage material. The silages were stored for 6, 12 and 24 days to study the effects of the yeast during the ensiling time. In total 36 small scale silos were prepared, corresponding to three replicates for each yeast level and time evaluation. At opening, 500 g of sample were collected from each silage, dried at 65 °C for 72 h and ground with a hammer meal. All samples were analysed for crude protein (CP), organic matter (OM) and ether extract (EE) following the procedures of (15). Neutral (NDF) and acid (ADF) detergent fiber were analysed following the procedure of (17). These analyses were done at the facilities of the Laboratory of Animal Nutrition and Feedstuffs Evaluation (LUNEA) of UNH, Peru. Additionally, the Hohenheim in vitro gas test was utilized to determine gas production (GP) in silage samples following the procedure of (7) at the facilities of the Institute of Agricultural Sciences in the Tropics, Section Animal Nutrition and Rangeland Management in the Tropics and Subtropics, of the University of Hohenheim. Metabolisable energy (ME) and organic matter digestibility (OMD) was estimated with the following equations based on GP (2):

 $\dot{ME} = 2.2 + 0.136 \times GP + 0.057 \times CP + 0.00285 \times CL^2$ (Eq. 1)

Where: ME = Metabolisable energy in MJ/kgDM; GP = Gas production in ml/200 mg of substrate dry matter (DM) after 24 h; CP =Crude protein concentration of grass hay in g/kg DM; CL= crude lipid concentration of grass hay in g/kg DM.

 $OMD = 14.88 + 0.889 \times GP + 0.045 \times CP + 0.065 \times CA$ (Eq. 2)

Where: OMD = Organic matter digestibility in g/100 g; GP = Gas production in ml/200 mg of substrate DM after 24 h; CP = Crude protein concentration of grass hay in g/kg DM; CA= crude ash in g/kg DM.

Statistical analysis

Statistical analyses were conducted using the software SAS 9.4 (SAS Institute Inc., Cary, NC, USA). The main effect of yeast level and time and their interaction was tested using the GLM procedure according to:

 $Y_{ij} = \mu + Y_i + T_j + (Y \times T)_{ij} + e_{ij}$

Where: Y_{ij} = dependent variable; μ = overall mean; Y_i = effect of the ith Yeast level; T_j = effect of jth ensiling Time; $(Y \times T)_{ij}$ = the interaction effect of Yeast level and ensiling Time; and eij = residual error of experiment. Significant effects of Yeast level were characterized using orthogonal polynomial contrasts testing the probability of linear or quadratic responses. Differences within means of Yeast level or ensiling Time were estimated by the Tukey test. The values reported are means and standard errors. Differences between treatments were declared significant at p < 0.05, whereas tendencies were declared at p < 0.10.

RESULTS AND DISCUSSION

There were no differences caused by the ensiling length on the nutrients concentration on barley silage, with the exception of ether extract which increased with longer ensiling Conversely, time (Table 1). the supplementation of yeast had an effect (quadratic) on all nutrients other than OM (Table 1). Crude protein was highest and ADF and EE lowest for yeast levels of 5 and 10 g/kg FM; whereas NDF was highest with the yeast level of 15 g/kg FM while all other levels remained unaffected. There was an ensiling Time \times Yeast level interaction effect for CP, EE, NDF and ADF. Where CP increased at yeast levels of 5 and 10 g/kg FM for the length of ensiling of 6 and 12 days, but decreased for the length of ensiling of 24 days; NDF and ADF had the opposite trend; and EE changed the proportional differences between Yeast levels depending on ensiling Time. Gas production, metabolisable energy and organic matter digestibility linearly decreased with increasing yeast level and were always higher in silages with no yeast addition. No time effects were observed for these parameters. In recent years some yeast species have been studied as inoculants because of their ability to inhibit detrimental silage microorganisms (10), and because of the benefits that yeasts may have on microbial ecology of the digestive tract of livestock (10). However, yeast are also lactate-assimilating microorganisms, and are therefore undesired in the silage because they are the primary initiators of aerobic spoilage in silage (12). A number of studies have evaluated the effects of yeast on silage quality.

It is difficult, however, to compare among those studies because of the differences in the type of yeast utilized, including specific strains of Saccharomyces cerevisiae and baker's yeast of different manufacturers. Additionally, the amounts of yeast added to the silage is not always presented in comparable units or it cannot be calculated back to the concentrations per unit of fresh matter. These are important considerations to keep in mind for the remaining of the discussion. Similar to the current study, increases in CP concentration by yeast addition to silages has been reported by Ok et al (11), who found that a strain of S. cereviseae treatment (20 g/kg FM) increased CP concentration of a rice straw silage compared with no yeast addition. Similarly, Saeed (14) found an increase in CP concentration upon addition of a commercial baker's yeast (100 g/kg FM) to reed silage. The main reason for an increase in CP concentration in a silage is related to the disappearance of other substrates (regularly sugars) coupled with a minimal degradation of protein to ammonia-N with consequent reduced losses of N in the effluents. It appeared that adding yeast at 5 and 10 g/kg FM to barley forages prevents the degradation of protein, however, once the concentration of yeasts increases a negative effect on CP appeared, likely reflecting an undesired fermentation where excessive production -and loss- of ammonia occurred. The reasons for this phenomenon cannot be explained with the data available from the current study, but it could be hypothesized that an excessive amount of yeast caused a shift in the microbial population, with the possibility that yeast started consuming lactate and/or caused aerobic spoilage of the silage. The NDF and ADF concentrations followed a pattern that can be similarly explained by the changes in CP concentration, where CP increased upon addition of yeast at 5 and 10 g/kg FM, but decreased when added at 15 g/kg FM, which would indicate an effect of dilution or concentration of the fiber components, when other substrates' concentration change. A study of (16) inoculated king grass with 3 inoculant treatments (untreated, Lactobacillus plantarum, and Lactobacillus plantarum plus S. cerevisiae) along with 3 different levels of rice bran (0, 5, and 10 g/100 g FM). After 21 d of ensiling in vitro ruminal degradation over 48 h measured by gas production was greatest for the L. plantarum treatment and this was higher than that of the L. plantarum plus S. cerevisiae treatment. This might indicate an effect of yeast on lactobacilli, where the presence of yeast hindered some of the positive effects of the lactic acid-producing bacteria. If this theory was truth it would explain the detrimental effects of yeast observed in this study, particularly in terms of in vitro OMD and ME, which strongly decreased upon addition of yeast, partially agreeing with the observations of Sofyan et al. (16). Similarly, GP decreased linearly with increasing yeast level, in agreement with Dunniere et al. (3) who found a numerical decrease in gas production for two strains of S. cerevisiae of up to 7 %. It appears that yeast addition during ensiling is detrimental to the quality of the forage, particularly in terms of rumen degradability. The reasons for the decreased GP, ME and OMD are not clear, but it is obvious that the quality of a forage will decrease with decreasing CP and increasing NDF and ADF. Yeast supplements have demonstrated in vitro and in vivo to increase the populations of cellulolytic bacteria (4,6), which would enhance the OM degradation. This effect, however, was not observed and it must be an evidence that not a significant amount of yeast remained available for rumen fermentation after the ensiling. No effects of time were observed on the quality of the silages, with the exception of EE, indicating that the characteristics of the silage may remain similar once the silage environment has stabilized (roughly after 3-4 days). Finally, there are two further considerations to make about these results. First, the quality of the original forage was much higher than that of the silages, including the control silage. For example GP for the original sample was 54.3 ml/200 mg DM, indicating a greater substrate degradability of the original forage material compared with the silages. A higher overall quality of fresh forages compared with their silages is commonly reported, but such an extreme difference as seen here can only indicate the difficulties to produce high quality barley silage, as argued by McDonald eta 1.(9). A second consideration is that 21 days of ensiling is close to the minimum time a silage would be stored before opening. Therefore, the silage quality that better reflects the forage that an animal would be consuming is that of the 21 days length of ensiling, which does not always correspond to the averages across all ensiling times. It still remains unclear what would the effects be for longer ensiling times, as several studies have reported increases in e.g. OM and starch degradability with advancing ensiling length (1,5), and how those longer conservation lengths would interact with the addition of yeast.

Table 1. Nutrients concentration of barley silage at different ensiling times with increasing levels of yeast addition.

1										
Item	Ensiling		Yeast level (g/kg FM)			Ensiling	a	p-values		
	time (days) ¹	0	5	10	15	time average ²	SEM	Yeast	Time	Yeast*Time
Organic matter	6	87.9	87.6	87.7	86.5	87.4	0.224			
	12	87.7	86.8	86.1	88.0	87.2		0.51	0.79	0.56
	24	87.9	87.0	87.0	88.3	87.6				
	Yeast level average ¹	87.8	87.2	86.9	87.6					
Crude protein	6	15.4	18.7	16.9	15.0	16.5	0.245	<0.01, Q	0.11	<0.01
	12	14.4	15.9	18.2	14.6	15.8				
	24	16.7	15.2	15.8	16.2	16.0				
	Yeast level average ¹	15.5 ^b	16.6 ^a	17.0 ^a	15.2 ^b					
Ether extract	6	3.37	2.64	2.10	2.30	2.6 ^b	0.110	<0.01, Q	<0.01	<0.01
	12	3.37	2.78	3.39	3.94	3.4 ^a				
	24	4.08	3.30	3.30	3.74	3.6 ^a				
	Yeast level average ¹	3.61 ^a	2.91^b	2.93 ^b	3.33 ^a					
Neutral detergent fiber	6	52.7	56.9	61.6	60.0	57.8	0.634	<0.01, L, Q	0.95	<0.01
	12	60.8	52.7	55.8	62.3	57.9				
	24	58.7	60.4	52.9	60.2	58.0				
	Yeast level average ¹	57.4 ^b	56.7 ^b	56.8 ^b	60.8 ^a					
Acid detergent fiber	6	34.6	33.4	37.7	37.7	35.8	0.428	<0.01, Q	0.65	<0.01
	12	39.1	33.0	35.3	38.5	36.5				
	24	37.7	37.3	33.1	37.0	36.2				
	Yeast level average ¹	37.1 ^a	34.6 ^b	35.4 ^b	37.7 ^a					

¹ Means with different superscripts within a row indicate statistical differences (p < 0.05)

² Means with different superscripts within a column indicate statistical differences (p < 0.05)

 Table 2. Gas production, metabolisable energy and organic matter digestibility of barley silage at different ensiling times with increasing levels of yeast addition

Térrer	Ensiling	Yeast level (g/kg FM)				Ensiling time	SEM	p-values		
Item	(days)	0	5	10	15	average ² S	SEM	Yeast	Time	Yeast*Time
	6	46.1	40.1	37.6	38.7	40.6	0.52	<0.01, L, Q	0.48	0.75
Gas	12	44.1	40.3	38.6	38.4	40.4				
production	24	45.0	39.4	39.0	38.4	40.5				
(ml/200 g DM)	Yeast level average ¹	45.1 ^a	39.9 ^b	38.4 ^b	38.5 ^b					
	6	8.97	8.74	8.29	8.33	8.58	0.071	<0.01, L, Q	0.64	0.51
Metabolisable	12	9.32	8.60	8.53	8.30	8.69				
energy (MJ/kg DM)	24 Yeast	9.32	8.46	8.43	8.39	8.65				
	level average ¹	9.20 ^a	8.60 ^b	8.42 ^{bc}	8.34 ^c					
	6	68.0	67.0	64.0	64.8	65.9	0.472	<0.01, L	0.68	0.45
Organic	12	70.3	66.4	66.5	63.3	66.6				
matter digestibility	24 Veast	70.2	65.2	65.1	63.9	66.1				
(g/100 g)	level	69.5 ^a	66.2 ^b	65.2 ^{bc}	64.0 ^{bc}					

¹ Means with different superscripts within a row indicate statistical differences (p < 0.05)

² Means with different superscripts within a column indicate statistical differences (p < 0.05)

ACKNOWLEDGEMENT

This study was financed by the Camisea Development Fund through the project: Evaluation of oats and barley planting in combination with vicia and conservation of forage for alpaca feeding in the Andean highlands of the Huancavelica province. Granted to the Huancavelica National University, Perú.

CONCLUSIONS

Yeast supplementation to barley silage had overall detrimental effects on silage quality, particularly at yeast level of 15 g/kg FM. Even though some advantage could be seen with lower yeast levels, as CP concentrations increased and NDF concentrations decreased, *in vitro* OMD and ME linearly decreased upon yeast addition. This may indicate negative effects of yeast on the fermentation process during ensiling, likely promoting the excessive degradation of soluble sugars and starch. Further research is needed to elucidate the modes of action of yeast when used as an additives for Barley silage.

REFERENCES

1. Benton, J. R., T. J, Klopfenstein and G.E, Erickson 2005. Effects of corn moisture and length of ensiling on dry matter digestibility and rumen degradable protein. Animal Science Department, Nebraska Beef Cattle Reports.

2. Close, W.H., and K.H, Menke 1986. Evaluation on the basis of metabolizable energy. Deutsche Stiftung für Internationale Entwicklung. In: Zentralstelle für Ernährung und Landwirtschaft (Ed.), Selected Topics in Animal Nutrition: A Manual Prepared for the 3rd Hohenheim Course on Animal Nutrition in the Tropics and Semi-Tropics, second edition, Feldafing, Germany, pp:60–63

3. Dunniere, L., L, Jin., B, Smiley., M, Qi., W, Rutherford., Y, Wang., and T, McAllister 2015. Impact of adding saccharomyces strains on fermentation, aerobic stability, nutritive value, and select lactobacilli populations in corn silage. Journal of Animal Science, 93, 2322–2335

4. Harrison, G.A., R.W, Hemken., K.A, Dawson., R.J, Harmon., and K.B, Barker 1988. Influence of addition of yeast culture supplement to diets of lactating cows on ruminal fermentation and microbial populations. Journal of Dairy Science, 71, 2967–2975

5. Hoffman, P.C., N.M, Esser., R.D, Shaver., W.K, Coblentz., M.P, Scott., A.L, Bodnar., R.J, Schmidt., and R.C, Charley 2011. Influence of ensiling time and inoculation on alteration of the starch-protein matrix in highmoisture corn. Journal of Dairy Science, 94, 2465–2474

6. Kung, L., E.M, Kreck., R.S, Tung., A.O, Hession., A.C, Sheperd., M.A, Cohen., H.E, Swain., and J.A.Z, Leedle, 1997. Effects of a live yeast culture and enzymes on in vitro ruminal fermentation and milk production of dairy cows. Journal of Dairy Science, 80, 2045–2051

7. Menke, K.H., and H, Steingass 1988. Estimation of the energetic feed value obtained from chemical analysis and in vitro gas production using rumen fluid. Anim. Res. Dev. 28, 7–55

8. McAllister, T.A., K.A, Beauchemin., A.Y, Alazzeh., J, Baah., R.M, Teather., and K, Stanford 2011. The use of direct fed microbials to mitigate pathogens and enhance production in cattle. Canadian Journal of Animal Science, 91, 193–211

9. McDonald, P., A.R, Henderson., and S.J.E, Heron 1991. Biochemistry of silage (Second Edition). Marlow, Chalcombe Publications, Journal of Animal and Feed Sciences, 26, 339

10. Muck, R.E., E.M.G, Nadeau., T.A, McAllister., F.E, Contreras-Govea., M.C, Santos., and L, Kung 2018. Silage review: Recent advances and future uses of silage additives, Journal of Dairy Science, 101, 3980–4000

11. Ok, J.U., S.M, Lee., S.J, Lee., J.H, Lim., T.W, Kang., H.Y, Jung., Y.H, Moon., and S.S, Lee 2006. Effect of yeast addition in rice straw silage fermentation, Journal of Animal Science and Technology, 48, 691–698

12. Pahlow, G., R,E, Muck., F, Driehuis., S.J, Elferink., W.H, Oude., and S.F, Spoelstra 2003. Microbiology of ensiling. Silage Science and Technology, Agronomy Monograph, 31– 93

13. Rojas, G. C., S.A, Catrileo., and Y.O, Romero 1997. Ensilaje de cebada en la engorda invernal de novillos hereford. Centro Regional de Investigación Carillanca Instituto de Investigaciones Agropecuarias. Casilla 58 D. Temuco. Chile

14. Saeed, A.A 2015. Effect of addition of baker's yeast Saccharomyces cerevisae and source of nitrogen on fermentation of reed silage and its nutritive value. Euphrates Journal Agriculture Science, 7, 10–24

15. Silva, D.J., and A.C, Quiroz 2002. Food analysis: chemical and biological methods (In Portuguese: Análise de alimentos: métodos químicos e biológicos). Minas Gerais, Brasil: Universidade Federal de Viçosa, Imprenta Universitaria, Viçosa, Brasil

16. Sofyan, A., L.M, Yusiati., Y, Widyastuti., and R, Utomo 2011. Microbiological characteristic and fermentability of king grass (Pennisetum hybrid) silage treated by lactic acid bacteria-yeast inoculants consortium combined with rice bran addition. Journal of the Indonesian Tropical Animal Agriculture, 36, 265–272

17. Van Soest, P. J., J.B, Robertson., and B.A, Lewis 1991. Methods for dietary, neutral detergent fiber, and non-starch polysaccharides in relation to animal nutrition. Symposium: carbohydrate methodology, metabolism and nutritional implications in dairy cattle. Journal Dairy Science, 74, 3583 – 3597